Acanthostigma and Tubeufia species, including T. claspisphaeria sp. nov., from submerged wood in Hong Kong

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Abstract: Acanthostigma scopulum, Tubeufia claspisphaeria sp. nov. and T. paludosa were identified from submerged wood collected in a small forest stream on Lantau Island, Hong Kong. The collections of Acanthostigma scopulum and Tubeufia paludosa differed slightly from the original descriptions. Tubeufia claspisphaeria differs from previously described species in that it has hook-shaped setae that form radially around the ostiole. This new species is described and illustrated and compared with the most similar species. A dichotomous key to the 16 accepted species of Tubeufia is provided.

Key words: new species, saprobic fungi, systematics

INTRODUCTION

There are 22 genera in the Tubeufiaceae (Barr 1980, Rossman 1987, Kirk et al 2001). Some genera, such as Melioliaphila and Uredinophila, are hyperparasites on sooty molds and on rust fungi, while Podonecctia is parasitic fungi on scale insects. Acanthostigma and Tubeufia are saprobes, usually found on old, rotten wood. Previous examinations of freshwater fungi occurring on submerged wood in streams in the tropics and subtropics have yielded numerous novel fungi, including hyphomycetes (Hyde et al 2002, McKenzie et al 2002), coelomycetes (Hyde 1993) and ascomycetes (Wong and Hyde 1999, Cai et al 2002). During our survey of fungi occurring on naturally submerged wood, we collected an Acanthostigma and two Tubeufia species, one being a species new to science, from a small forest stream in Hong Kong. The three species are described, illustrated and compared with known taxa. A key to the 16 accepted species of Tubeufia is provided.

MATERIALS AND METHODS

Submerged wood was retrieved from a stream near the Trappist Monastery, Lantau Island, Hong Kong, in Apr 2002 and returned to the laboratory, where it was incubated in zip-lock plastic bags at room temperature. Humidity was maintained by adding moistened paper towels. Samples were examined within 3 d and periodically over 1 mo for the presence of sporulating structures. Cultures of fungi were obtained where possible from single spores (Choi et al 1999). To stimulate sporulation, 1 cm² blocks of colonized agar were placed in water in a bubble chamber overnight.

TAXONOMY

Acanthostigma De Not. and Tubeufia Penz. & Sacc. Acanthostigma currently includes six species and recently has been reviewed by Réblová and Barr (2000). The genus is characterized by vinaceous, reddish-brown or dark brown ascomata that are covered with dark brownish-black, often opaque, obtuse or acute setae. The ascospores are hyaline, multisepate and cylindrical-fusiform to elongate fusiform. The anamorphs are in Helicosporium and Helicomycus. In species of Tubeufia the ascomata are hyaline, whitish or yellowish to pinkish, but may become dark at maturity. They are smooth, or are covered with protruding cells, thick-walled hyphal appendages, or short dark setae (Réblová and Barr 2000).

Acanthostigma scopulum (Cooke & Peck) Peck, Bull. New York State Mus. 1:22 (1887) (Figs. 1–6) Ascomata 145–250 µm diam, superficial, globose, solitary or gregarious, dark brown to black, membranous, ostiolate, with setae; setae (50–)80–90–(99) µm wide at base, 1.5–2 µm wide at apex, dark brown, 1–2-septate, thick-walled, straight. Asc (64–)90–120–(130) × 8–10 µm

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(\( \bar{x} = 103 \times 9.2 \) μm, n = 20), 8-spored, cylindric-clavate, bitunicate, short stalked, pseudoparaphyses. Ascospores (66–70–85–90) × 3–3.5–(4) μm (\( \bar{x} = 78 \times 3.4 \) μm, n = 30), fasciculate, long-fusiform to cylindrical-fusiform, hyaline, straight or slightly curved, (5–)9–12-septate, smooth-walled, guttulate, lacking appendages and sheaths.


Colonies on potato-dextrose agar olive-colored, reaching 1 cm diam in 10 d at room temperature (\( \approx 28 \) C), no pigment diffusing into agar, not sporulating even after submergence overnight in a bubble chamber.

Substratum. Wood submerged in streams, decaying wood (*Fagus sylvatica*, *Pinus* sp. and *Tsuga* sp.)

Known distribution. Australia, Europe, Hong Kong and U.S.A.

Specimen examined. HONG KONG. Lantau Island, Trappist Monastery, small stream in forest, on wood partially submerged, 21 Apr 2002, K.D. Hyde (HKU[M] 17121; living culture HKUCC 9117).

Notes. A key to species of *Acanthostigma* was provided by Reblova and Barr (2000). This collection differs from the description for A. *scopulum* provided by Reblova and Barr (2000) in that it has septate setae and wider ascospores 3–3.5(–4) μm versus (2–)2.5–3(–3.5) μm but in other aspects it is similar.

**Tubefia claspisphaeria** Kodsiueb, sp. nov.

(Figs. 7–16) Ascomata 185–330(–350) μm diametro, superficialia, globosa, solitaria vel aggregata, hyalina vel pallide brunnea ubi immatura, atrobunnea vel atris ubi maturus, membranea, ostiolata, setosa; setae usque 47 μm longa, 10 μm lata, (0–)1(–2)-septata, atrobunnea, hamatus. Peries ascomati usque 60 μm lata crassus, vinacea-brunnea, e cellulis pseudoparenchymatis compositum, 4–5 stratosus, textura globulosa. Asci (93–)105–120(–127) × (12–)14–16(–18) μm, octospori, cylindrico-clavati, bitunicati, pedicellati. Ascosporae (34–)45–60(–64) × 5–7(–8) μm, fasciculatae, elongatae-fusiformis, (3–)5–6(–8)-septatae, hyalinae, curvatae, guttulatae.

Etymology. In reference to the clasp-like setae on the ascomata. Ascomata 185–330(–350) μm diam, superficial, globose, solitary or grouped, hyaline to pale brown when immature, dark brown becoming black when mature, membranous, ostiolate, with setae that form around ostiole. Setae from surface of ascomata, up to 47 μm long and 10 μm wide, (0–)1(–2)-septate, dark brown, thick-walled, hook-shaped. Peridium up to 60 μm wide, vinaceous brown, comprising 4–5 lay-

Table I. *Tubefusia* species and their present synonyms (accepted name in **bold**)

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Synonym</th>
<th>Notes</th>
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<tbody>
<tr>
<td><em>T. acaciae</em> Tilak &amp; S. B. Kale</td>
<td>None</td>
<td>Although not examined this taxon may not be a <em>Tubefusia</em> with its violet to red ascomata and multiseptate ascospores</td>
</tr>
<tr>
<td><em>T. aciculospora</em> Katum. &amp; Y. Harada</td>
<td>None</td>
<td>This species is unusual as it has ascospores with pointed ends Katumoto and Harada (1979)</td>
</tr>
<tr>
<td><em>T. alpina</em> L. Holm &amp; Nograsek</td>
<td><em>Acanthostigmina longisporum</em></td>
<td>This was considered to be synonymous with <em>A. longisporum</em> by Réblova and Barr (2000)</td>
</tr>
<tr>
<td><em>T. aureolata</em> Bat. &amp; Garnier</td>
<td><em>Saccardomyces socius</em> Hennebert</td>
<td>Rossman (1979)</td>
</tr>
<tr>
<td><em>T. aurantiella</em> (Penz. &amp; Sacc.) Rossman</td>
<td><em>Calonectria aurantiella</em></td>
<td>Mentioned in Crane et al (1998), transferred from <em>Calonectria</em> (Rossman 1979)</td>
</tr>
<tr>
<td><em>T. brevispina</em> (M. E. Barr &amp; Rogers) J. L. Crane, Shearer &amp; M. E. Barr</td>
<td>(=<em>Acanthostigmella brevispina</em>)</td>
<td>Crane et al (1998)</td>
</tr>
<tr>
<td><em>T. clintonii</em> (Peck) M. E. Barr</td>
<td><em>Acanthostigma perpusillum</em></td>
<td>This was considered to be synonymous with <em>A. perpusillum</em> by Réblova and Barr (2000)</td>
</tr>
<tr>
<td><em>T. coccicola</em> (Ellis &amp; Everh.) Höhn.</td>
<td><em>Podonectria coccicola</em> Petch</td>
<td>Roszman (1987)</td>
</tr>
<tr>
<td><em>T. corynespora</em> Munk</td>
<td><em>Tubefusia paludosa</em></td>
<td>Synonymized by Rossman (1977)</td>
</tr>
<tr>
<td><em>T. dactylariae</em> Chang</td>
<td>None</td>
<td>Etayo (2002)—possibly <em>Chaetosphaerulina</em></td>
</tr>
<tr>
<td><em>T. eriodermae</em> Etayo</td>
<td><em>Acanthostigmella genuflexa</em></td>
<td>Réblova and Barr (2000)</td>
</tr>
<tr>
<td><em>T. minuta</em> Munk</td>
<td><em>Herpotrichiella</em></td>
<td>Barr (1980)</td>
</tr>
<tr>
<td><em>T. nigrotuberculata</em> T. Hino &amp; Katsum.</td>
<td><em>Herpotrichia nigrotuberculata</em></td>
<td>Pirózynski (1972)</td>
</tr>
<tr>
<td><em>T. pacchythrix</em> (Rehm) Rossman</td>
<td>None</td>
<td>For discussion see Rossman (1979)</td>
</tr>
<tr>
<td><em>T. paniculata</em> (Peck) Dennis</td>
<td>None</td>
<td>Etayo (2002)—possibly <em>Chaetosphaerulina</em></td>
</tr>
<tr>
<td><em>T. parvula</em> Dennis</td>
<td>None</td>
<td>Drawn in Ellis and Ellis (1985)</td>
</tr>
<tr>
<td><em>T. pezizula</em> (Berk. &amp; M. A. Curtis) M. E. Barr</td>
<td><em>Thaxteriella pezizula</em></td>
<td>Type species of <em>Thaxteriella</em> (Sivanesan 1984, Barr 1980)</td>
</tr>
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</table>
| *T. roa**
### Table I. Continued

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Synonym</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>T. rugosa</em> C. Booth</td>
<td><em>Tubeufia helicoma</em></td>
<td>Barr (1980)</td>
</tr>
<tr>
<td><em>T. scopula</em> (Cooke &amp; Peck), M. E. Barr</td>
<td><em>Acanthostigma scopulum</em></td>
<td>Barr (1980), Rélová and Barr (2000)</td>
</tr>
<tr>
<td><em>T. stromaticola</em> (Henn.) Rossman</td>
<td>Transferred from <em>Calonectria</em></td>
<td>Rossman (1979)</td>
</tr>
<tr>
<td><em>Ophionectria trichospora</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>T. trichospora</em> (Berk. &amp; Broome) Petch</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>T. vermicularispora</em> (T. Hino &amp; Katum.) Sivan.</td>
<td><em>Chaetosphaerulina vermicularispora</em></td>
<td>This has black carbonaceous ascomata and is probably not a <em>Tubeufia</em>. It was described as a <em>Chaetosphaerulina</em> (Hino and Katumoto 1954)</td>
</tr>
</tbody>
</table>

ers of thick-walled textura globulosa, outer cells brown-walled, inner cells with hyaline walls. Pseudo-paraphyses 1.5–2.5 μm wide, hypha-like, numerous, cellular, unbranched. Asci (93–)105–120(–127) × (12–)14–16(–18) μm (x = 109 × 15.1 μm, n = 15), 8-spored, cylindric-clavate, bitunicate, with a small ocular chamber, persistently pedicellate. Ascospores (34–)45–60(–64) × 5–7(–8) μm (x = 49 × 5.9 μm, n = 35), fasciculate, elongate-fusiform, (3–)5–6(–8)–septate, hyaline, sometimes slightly curved, smooth-walled, guttulate, lacking appendages or sheaths.

**Anamorph.** Unknown.

Colonies on potato-dextrose agar dark green to black, 1 cm diam in 1 wk at room temperature (~28 C). Mycelium mostly immersed, aerial mycelium velvety to fluffy, no pigment diffusing into agar, not sporulating even after submergence overnight in a bubble chamber. Mycelium less dense at the outer edge than in center, with branching mycelial strands extending from edge of colony.

**Substratum.** Wood submerged in streams.

**Known distribution.** Hong Kong.


**Tubeufia paludosa** (P. Crouan & H. Crouan) Rossman, Mycologia 69:383 (1977) *(Figs. 17–23)*

Ascomata 185–350 μm diam, superficial, globose, brown to dark-brown, solitary, sparse, surface roughened, membranous. Asci (162–)170–180 × (14–)15–18(–19) μm (x = 170 × 16.1 μm; n = 17), numerous, 8-spored, cylindric-oblong, bitunicate, hyaline, with a subapical ring, rounded at apex, persistently pedicellate, pseudoparaphyses. Ascospores (77–)120–135(–155) × 4–5(–6) μm (x = 127 × 4.3 μm, n = 30), fasciculate, narrowly elongate, cylindric or filiform, hyaline, (18–)20–25-septate, straight or slightly curved, smooth-walled, guttulate, with small mucilaginous pads at each end.

**Anamorph.** Helicosporium sp.

Colonies on potato-dextrose agar 3.5 cm diam in 3 wk. Mycelium mostly immersed, no pigment diffusing into agar, hyphae less dense in outer zone, sporulating, forming a brownish-gray turf of upright setiform conidiophores, becoming ochraceous or brownish with age; conidiogenous cells produced laterally as thin-walled pegs. Conidia coiled two or three times, multiseptate, coils 37–86.4 μm diam, cells 4.6–5.4 μm wide.

**Substratum.** Wood submerged in streams, decaying woody fruit, palm fruit peduncle and bamboo.

**Known distribution.** Bermuda, Brazil, Columbia, Europe, Hong Kong, India, Indonesia, Panama, Trinidad, U.S.A. and Venezuela.


The genus *Tubeufia* has included at least 40 epi-thets as listed in IndexFungorum (http://www.indexfungorum.org/Names/Names.asp). Many of these taxa have been transferred to other genera (Barr 1980, Crane et al 1998) as summarized in Table I. Our two species are typical of *Tubeufia* because the ascomata are initially pale and have either...
DICHTOMOUS KEY TO THE SPECIES OF TUBEUFIA

1. Lichenicolous fungi ................................................................. 2
1. Not lichenicolous fungi .......................................................... 3

2. Ascomata orange-yellow to grayish-yellow, ascospores 40–50 × (3.5–4.5) µm, fusiform, 5–8-septate ........ T. panuariae

3. Ascospores < 10 septa .............................................................. 4
3. Ascospores with > 10 septa .................................................... 14
   4. Ascospores shorter than 21 µm ........................................... 5
   4. Ascospores longer than 21 µm ........................................... 6

5. Ascomata reddish-brown, ascospores 14–15 × 3.5–4 µm, oblong-elliptical, 3-septate .................. T. parvula
5. Ascomata hyaline to pale yellow, ascospores (13–)14–18 × (3.5–)4–5(–7) µm, fusoid, 3-septate ........ T. brevispina
6. Ascospores up to 7 septa ..................................................... 7
6. Ascospores > 7 septa ............................................................ 11
7. Ascospore 48–56 × 6–7 µm, cylindrical or worm-like, often curved, pointed at both ends, 5–7-septate .... T. acaciae
7. Ascospores narrower than 6 µm .............................................. 8
8. Ascospores less than 3.5 µm wide ........................................ 9
8. Ascospores more than 3.5 µm wide ..................................... 10
9. Ascospores 65–75 × 2.5–3.5 µm, acicular, 4–7-septate .................. T. aciculospora
10. Ascospores 40–57 × 4–5 µm, long-fusiform, multisepaete (4-celled according to Rehm 1907) ........ T. pachythrix
10. Ascospores 50–70 × 4–5 µm, long-fusoid, 5–7-septate ........ T. stromaticola
11. Ascomata with a row of hook-like setae, form radially around the ostiole, ascospores (34–)45–60(–64) × 5–7(–8) µm, elongate-fusiform, (3–)5–6(–7)-septate .................. T. claspisphaeria
11. Ascomata not as above .......................................................... 12
12. Ascospores 32–36 × 6–7 µm, fusiform, 7–8-septate ................ T. dactylariae
12. Ascospores narrower than 6 µm .......................................... 13
13. Ascospores 45–66 × 3.5–5 µm, narrowly fusiform to cylindric, 7–9-septate ......................... T. aurantiella
14. Ascospores shorter than 70 µm ........................................... 15
14. Ascospores (70–)100–200(–230) × (2–)3.5–7(–8) µm, fusiform with acute ends, up to 35 septa ...... T. paludosa
15. Ascospores 40–55(–65) × (2.5–)3–5 µm, elongate clavate or fusoid, (5–)7–9(–13)-septate ........ T. cylindrothecia
15. Ascospores (27–)30–52 × 2.5–3.5(–4.5) µm, elongate fusoid, often curved, (5–)7–10(–13)-septate ........ T. cerea

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