

Precise regulation of monomer reactive sites enhances water permeance and membrane selectivity of polyamide nanofiltration membranes

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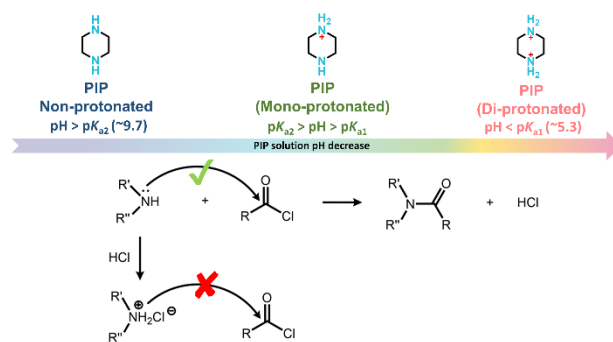
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ABSTRACT

Polyamide structure and chemistry play a critical role in the separation performance of thin film composite (TFC) nanofiltration (NF) membranes. Typically, the polyamide formation is based on the reactive sites on monomers (e.g., amino groups on piperazine (PIP)) during interfacial polymerization (IP). To precisely tailor polyamide properties (e.g., crosslinking degree) and membrane performance, we regulated PIP reactive sites by pH adjustment to vary the dominant species in forms of non-, mono-, and di-protonated PIP. Specifically, at pH values between pK_{a1} (i.e., 5.3) and pK_{a2} (i.e., 9.7), the dominant mono-protonated PIP with relatively fewer non-protonated PIP resulted in reduced crosslinking degree of polyamide. Such reduced crosslinked polyamide exhibited simultaneously improved water permeance and better solute-to-solute selectivity (e.g., $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ and $\text{CaCl}_2/\text{PFOS}$ selectivity), thanks to their looser structure and more negative charge. For example, the membrane NF-pH9, prepared at pH 9, exhibited simultaneously improved water permeance ($20.2 \text{ L m}^{-2} \text{ h}^{-1} \text{ bar}^{-1}$) and higher $\text{CaCl}_2/\text{PFOS}$ selectivity (12.6). The dominant di-protonated PIP with few non-protonation at $\text{pH} < pK_{a1}$ resulted in ineffective crosslinked polyamide with low salt rejection (e.g., $0.9 \pm 0.3\%$ of Na_2SO_4). This study investigated a facile strategy to tailor membrane permeance and selectivity by regulating monomer reactive sites, which provides new insights into developing high-performance NF membranes.

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1. INTRODUCTION

Nanofiltration (NF) technology has been extensively applied in a wide range of water treatment scenarios.^{1,2} This technology is generally based on thin film composite (TFC) membranes featuring a semi-aromatic polyamide layer on a porous substrate.^{3, 4} The polyamide layer is typically prepared by the interfacial polymerization (IP) between piperazine (PIP) in an aqueous solution and trimesoyl chloride (TMC) in an organic solvent.^{5,6} One essential indicator for NF membrane performance is the solute-to-solute selectivity, which enables selective retention/passage of small molecules and ions. The membrane selectivity is greatly influenced by the structure and chemistry (e.g., pore size and surface charge) of its polyamide layer.^{7, 8}

To date, several strategies have been applied to tailor IP for regulating the structure and chemistry of polyamide nanofilms toward better membrane selectivity,⁹⁻¹¹ such as co-solvent addition^{12, 13} and nanofiller incorporation^{14, 15}. For example, some studies applied co-solvents (e.g., acetone^{16, 17} and ethyl acetate^{18, 19}) to regulate the IP reaction to shape polyamide morphology²⁰ and/or tailor the crosslinking degree²¹. Some works incorporated porous metal-organic frameworks²² and/or non-porous nanoparticles²³ during IP reaction to create selective channels toward high membrane performance. However, these strategies can suffer from some uncontrollable factors. For example, the volatility of co-solvents²⁰ and aggregation of nanofillers²³ can be inevitable, leading to their uncontrolled participation in IP. Besides, co-solvent toxicity and nanoparticle leaching can also harm the environment. Therefore, a facile and sustainable strategy to precisely control the IP reaction is desirable to effectively tailor polyamide properties toward better membrane selectivity.

Notably, polyamide polymerization is based on the reactive sites on monomers²⁴ (i.e., amino groups on PIP and acyl chloride groups on TMC). PIP shows strong nucleophilicity due to the lone pair electrons on the nitrogen atom of its amino groups. This can lead to the nucleophilic attack of PIP to the electron-deficient sites of acyl chlorides on TMC.²⁵ Highly reactive PIP molecules can continuously attack acyl chlorides during IP to form crosslinked polyamide.²⁴ According to the Henderson-Hasselbalch equation, PIP as a diacidic base with two pK_a values ($pK_{a1} = 5.3$ and $pK_{a2} = 9.7$)²⁶ can be protonated by H^+ to form mono- and di-protonated PIP at different pH.²⁷ Presumably, adjusting pH can effectively control the dominant monomer species in forms of non-, mono-, and di-protonated PIP, thereby influencing the reactivity of amine solution with TMC for the polyamide formation. Specifically, at a $pH > pK_{a2}$, the dominant non-protonated PIP with high reactivity can form the highly crosslinking degree of polyamide (**Figure 1a**). While at a lower pH, reduced non-protonated PIP with more protonated species can lead to less or ineffective crosslinked polyamide formation (**Figure 1b and c**). Indeed, several studies performed IP at different pH to tailor membrane performance.²⁸⁻³¹ However, the critical role of the pH-regulated reactive sites on PIP monomers in polyamide properties and membrane performance has not yet been systematically investigated.

Herein, we prepared polyamide NF membranes with varied pH values of amine solutions during IP. We hypothesize that the reactive sites on PIP could be effectively regulated by H^+ for polyamide formation with tailored properties. Due to the precise regulation, higher water permeance and better solute-solute selectivity were simultaneously achieved. Expectedly, this work using a facile paradigm provides insights into the role of the monomer reactive sites on membrane performance, paving

a novel direction for developing selective NF membranes toward diverse water treatment scenarios.

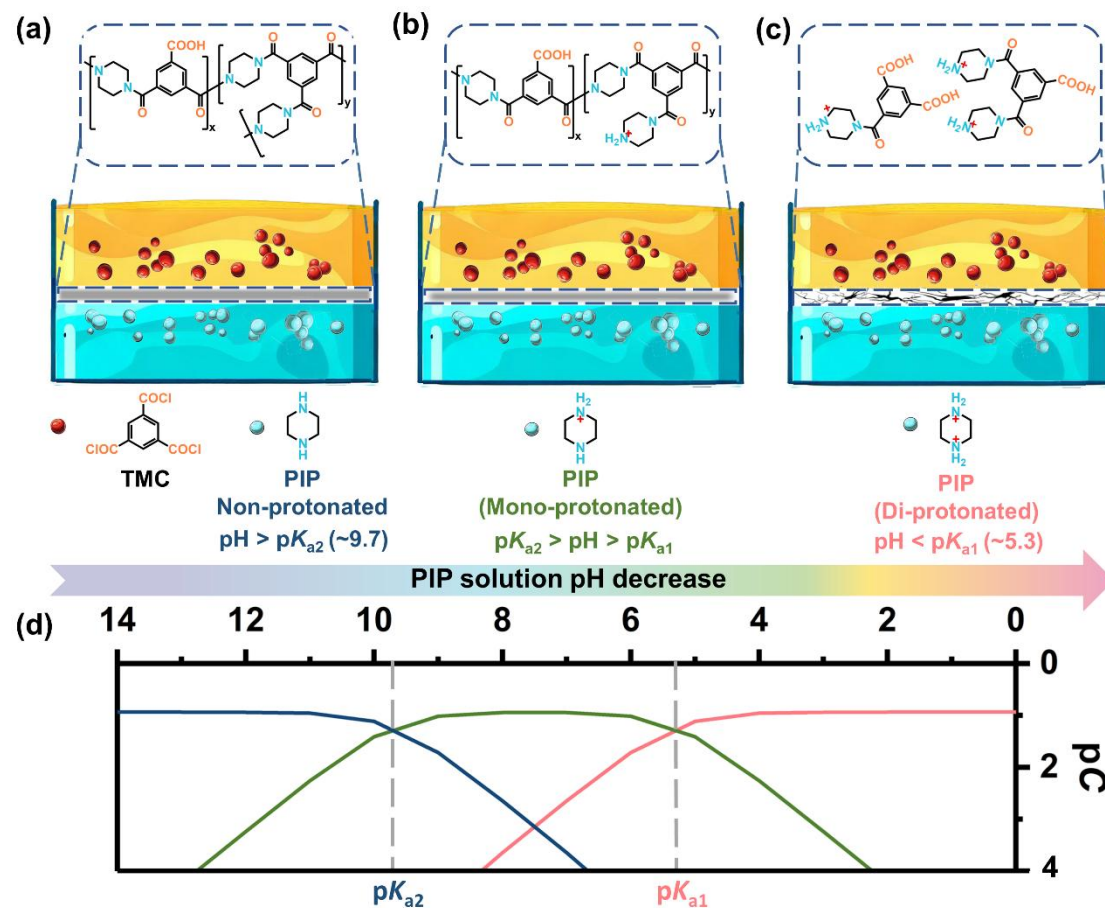


Figure 1. Schematic illustration of the dominant PIP species at different pH and corresponding interfacial polymerization. The formation of (a) highly crosslinked polyamide, (b) less crosslinked polyamide, and (c) ineffective crosslinked polyamide, at $\text{pH} > \text{p}K_{\text{a}2}$ (non-protonated PIP is dominant), $\text{p}K_{\text{a}1} < \text{pH} < \text{p}K_{\text{a}2}$ (mono-protonated PIP is dominant), and $\text{pH} < \text{p}K_{\text{a}1}$ (di-protonated PIP is dominant), respectively. (d) The effect of pH on the different PIP species in amine solution. The total PIP concentration is 1.0 wt%. The $\text{p}K_{\text{a}1}$ and $\text{p}K_{\text{a}2}$ of PIP are 5.3 and 9.7, respectively (values obtained from reference²⁶).

2. EXPERIMENTAL SECTION

2.1 Materials

Monomers piperazine (PIP, 99%, Sigma-Aldrich) and trimesoyl chloride (TMC, 98%, Sigma-Aldrich) were used for the IP reaction to form polyamide. n-Hexane (95%, Duksan) was used to dissolve TMC. Hydrochloride acid (HCl, VWR International) was applied to adjust the pH values of PIP solutions to fabricate pH-regulated NF membranes. The polyethersulfone (PES) ultrafiltration membrane (UP150) with a molecular weight cut-off (MWCO) of 150,000 Da, purchased from MICRODYN-NADIR, was used as the substrate in this study. The PES substrate was pre-treated with 25% (v/v) isopropanol/water solution for 1 h and then stored in deionized (DI) water before use. Sodium sulfate (Na_2SO_4 , Dieckmann), sodium chloride (NaCl, Dieckmann), calcium chloride (CaCl_2 , Dieckmann), and magnesium chloride (MgCl_2 , Uni-Chem) were used to test the salt rejection of NF membranes. Glycerol (M.W. 93.12, Dieckmann), glucose (M.W. 180.16, Dieckmann), raffinose (M.W. 504.44, Sigma), and dextran (M.W. 1000, D-chem) were used to measure the MWCO of NF membranes. Anionic perfluoroalkyl substances, including perfluoro (2-methyl-3-oxahexanoic) acid (GenX, Macklin), potassium perfluorooctanesulfonate (PFOS, Sigma-Aldrich), perfluorobutyric acid (PFBA, Alfa Aesar), sodium perfluorooctanoate (PFOA, Alfa Aesar), and potassium nonafluoro-1-butanesulfonate (PFBS, TCI) were used for testing

micropollutant removal of NF membranes. Unless specified otherwise, the chemicals were dissolved in DI water for experiments.

2.2 Synthesis of NF Membranes

The NF membrane was fabricated via an IP reaction on a UP150 substrate (**Figure S1**).³²⁻³⁴ In brief, a 1.0 wt% PIP solution with different regulated pH values (i.e., 10, 9, 8, and 7) was poured onto a UP150 substrate with an immersing time of 2 min. Then, the excessive PIP solution was removed with a rubber roller. Afterward, a 0.1 wt% TMC hexane solution was poured onto the PIP-impregnated substrate with a time of 1 min to produce the polyamide NF membrane. The fabricated membrane was rinsed by fresh n-hexane to clean off excess TMC and then stored in DI water before further use. These pH-regulated membranes were denoted as NF-pHx, where x represented the corresponding pH values of PIP solutions. The control membrane (denoted as NF-control) was formed by a 1.0 wt% PIP (pH 11.6, without pH regulation). We further adjusted the pH to 4 for the fabrication of ineffective crosslinked polyamide (denoted as NF-pH4)

2.3 Calculation of distribution coefficients for PIP species

The distribution coefficients of di-, mono-, and non-protonated PIP species in amine

solution (i.e., α_0 , α_1 , and α_2 , respectively, see **Figure 1d**) can be specified as the fractions of corresponding PIP species over the total dissolved PIP molecules, which could be calculated by:

$$\alpha_0 = [H^+]^2 / (K_{a1}K_{a2} + K_{a1}[H^+] + [H^+]^2) \quad (1)$$

$$\alpha_1 = K_{a1}[H^+] / (K_{a1}K_{a2} + K_{a1}[H^+] + [H^+]^2) \quad (2)$$

$$\alpha_2 = K_{a1}K_{a2} / (K_{a1}K_{a2} + K_{a1}[H^+] + [H^+]^2) \quad (3)$$

where K_{a1} and K_{a2} are the equilibrium constants of protonated PIP species. $[H^+]$ is the concentration of hydrogen ions in PIP solution, given as a pH value.

2.4 Characterization Methods

2.4.1 Morphology observation

Unless specified otherwise, membrane samples for all characterizations were dried in an oven with a temperature of 60 °C for 30 min. Field-emission scanning electron microscopy (FE-SEM, S-4800, Hitachi) was used for the surface morphology characterization of NF membranes. Before SEM characterization, a thin gold layer was sputter-coated onto the dried membrane samples to avoid charge accumulation.³⁵

Transmission electron microscopy (TEM, G2 20 Scanning, FEI Tecnai) was used for cross-section structure characterization of membrane samples.³⁶ Atomic force microscopy (AFM, Dimension Icon, Bruker) was used for the measurement of

membrane surface roughness with a $5.0 \times 5.0 \mu\text{m}^2$ scanning area.³⁷

2.4.2 Chemical properties of NF membranes

Attenuated total reflection Fourier transform infrared spectroscopy (ATR FTIR, Nicolet iS5, Thermo Fisher Scientific) was conducted to analyze the polyamide formation.¹⁶ A goniometer installed with a video capture device (OCA20, Dataphysics) was used to measure the water contact angles of dried membrane samples.³⁸ Streaming zeta potential (ζ) measurement tested by an electrokinetic analyzer (SurPASS 3, Anton Paar, Austria) was employed to analyze membrane surface charge in a pH range of 3.0-10.0 with 1.0 mM potassium chloride as the background solution.¹⁶ X-ray photoelectron spectroscopy (XPS, Thermo Fisher Scientific, USA) with an X-ray source of Al- $K\alpha$ gun was conducted to evaluate elemental compositions of membrane surface. The crosslinking degree of a typical PIP-based polyamide network could be calculated by the oxygen-to-nitrogen ratio (O/N) using the atomic percent of O and N:³⁵

$$n = \frac{4-2r_{O/N}}{1+r_{O/N}} \quad (4)$$

where n is the crosslinking degree of polyamide and $r_{O/N}$ is the oxygen-to-nitrogen ratio obtained from XPS results.

2.5 Membrane Separation Performance

2.5.1 Water permeance and salt rejections

A cross-flow instrument was employed for membrane filtration experiments. The active filtration area of a membrane sample was $1 \times 2 \text{ cm}^2$ and the operation temperature was $25 \pm 1 \text{ }^\circ\text{C}$. Before the sample collection for the tests of separation performance, the membrane coupon was stabilized under 5 bar for 1h. Then, the applied pressure was kept at 5 bar for the evaluation of water permeance and solute rejection. The water flux was calculated according to the following equation:

$$J_w = \frac{\Delta V}{\Delta t \times S} \quad (5)$$

where J_w ($\text{L m}^{-2} \text{ h}^{-1}$) is the water flux, ΔV (L) is the permeate volume, Δt (h) is the sample collecting time, and S (m^2) is the effective membrane area. The water permeance was further calculated by the following equation:

$$A = \frac{J_w}{\Delta P - \Delta \pi} \quad (6)$$

where A ($\text{L m}^{-2} \text{ h}^{-1} \text{ bar}^{-1}$) is the water permeance, ΔP (bar) is the applied hydraulic pressure, and $\Delta \pi$ (bar) is the transmembrane pressure difference.

The salt rejection was measured using the feed solution with a single type of salt dissolved (1000 ppm Na_2SO_4 , MgCl_2 , CaCl_2 , and NaCl), respectively. The MWCO experiment was performed by conducting the rejection tests of different neutral solutes

(200 ppm glycerol, glucose, raffinose, and dextran). The solute rejection was calculated according to the following equation:

$$R = \frac{C_f - C_p}{C_f} \quad (7)$$

where R is the solute rejection, C_f and C_p are the solute concentration of feed and permeate solution, respectively. A conductivity meter (Ultrameter II, Myron L Company, Carlsbad, CA) was applied to measure salt concentration. The concentration of neutral solutes was measured by a total organic carbon analyzer (TOC-L CPH, Shimadzu, Japan).

2.5.2 The analysis of polyamide pore size

According to previous literature,^{32,39} the rejection of these neutral solutes could be used to determine the pore size of polyamide. The distribution curve of polyamide pore size is presented as a log-normal distribution based on following assumptions: (1) the mean pore radius of polyamide membranes is equal to the radius of neutral solute molecular at 50% rejection; (2) the geometric standard deviation is defined as the ratio of solute radius at 84.13% rejection to that at 50% rejection; (3) there is no hydrodynamic or steric interaction between neutral solutes and polyamide pores.^{11, 40} The mathematical expression for the distribution curve is given by:

$$\frac{dR(r_p)}{dr_p} = \frac{1}{r_p \ln \sigma_p \sqrt{2\pi}} \exp \left[-\frac{(\ln r_p - \ln \mu_p)^2}{2(\ln \sigma_p)^2} \right] \quad (8)$$

where r_p is the polyamide pore size, μ_p is the mean pore size, and σ_p is the geometric standard deviation of probability density function curve.

2.5.3 The rejection tests of contaminants by NF membranes

Anionic perfluoroalkyl substances, including PFOS, PFOA, GenX, PFBS, and PFBA, were applied to measure the rejection of contaminants by NF membranes. **Table S1** shows the physicochemical properties (e.g., structure formula and molecular weight) of these contaminants used in this work. Generally, these contaminants exist as anionic compounds under a circumneutral pH (~6.5) used for filtration tests. Before water sample collection, the testing membranes were compacted for 2 h with a feed solution consisting of PFOS, PFOA, GenX, PFBS, and PFBA (each with a concentration of 200 ppb) at an electrolyte background of 10 mM NaCl. The micropollutant concentration of feed and permeate solutions was tested by liquid chromatography with tandem mass spectrometry (LC-MS/MS, 1290 Infinity, Agilent; 3200 QTRAP, AB SCIEX, Singapore). The reversed-phase column (ZORBAX Eclipse Plus C18, Agilent) was 2.1×50 mm in dimension with the particle size of 1.8 μm. Details of contaminants analytical method can be referred in our previous work.⁴¹

2.5.4 The analysis of solute-to-solute selectivity

The solute-to-solute selectivity of NF membranes was evaluated according to the following equations:^{42, 43}

$$S_{i/j} = \frac{1-R_i}{1-R_j} \quad (9)$$

where $S_{i/j}$ is the solute/solute selectivity of solute i over j (e.g., S_{CaCl_2/Na_2SO_4} represents the membrane selectivity between $CaCl_2$ and Na_2SO_4).

3. RESULTS AND DISCUSSIONS

3.1 Effectively Tailored Structure and Chemistry of NF Membranes

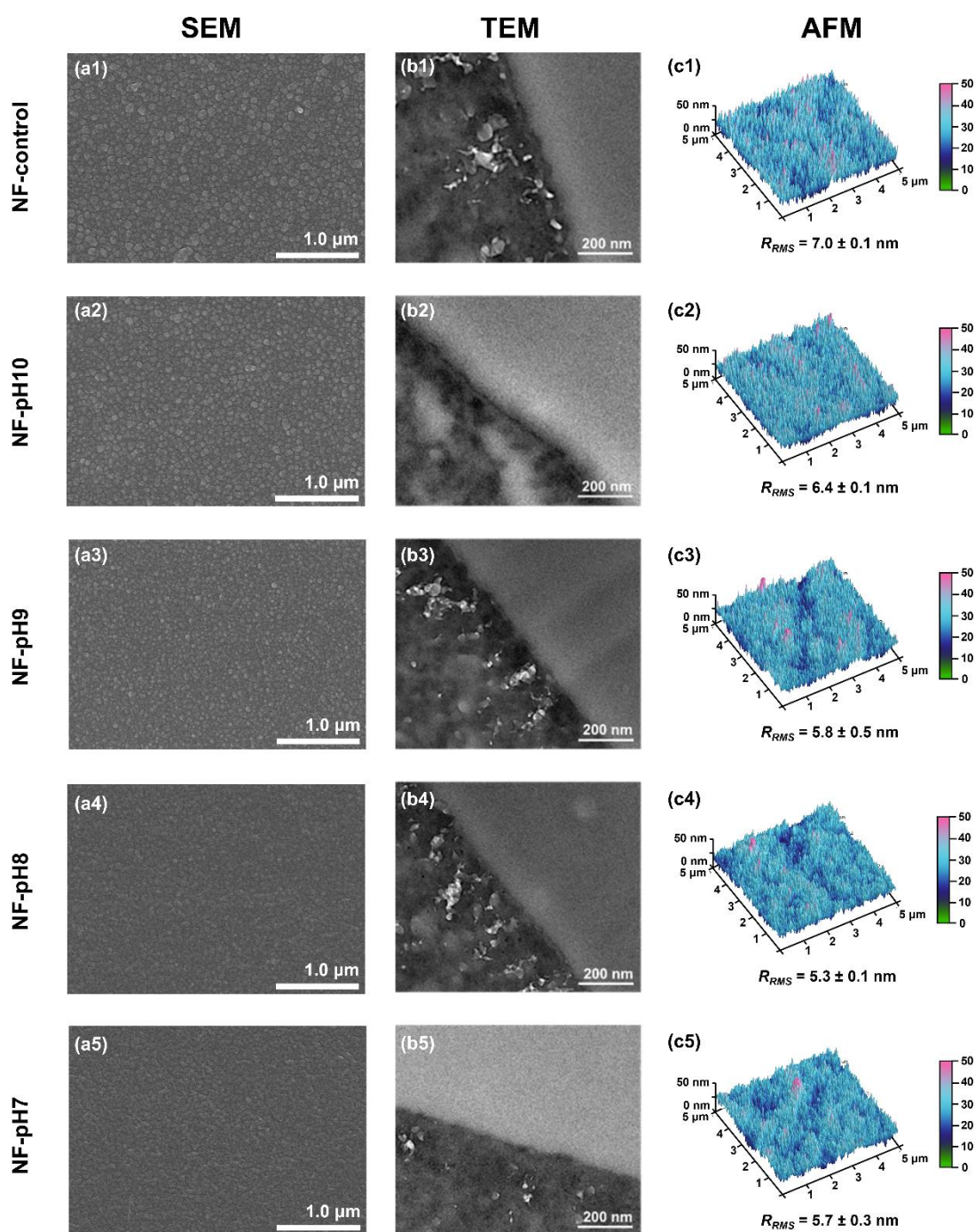


Figure 2. Micrographs of the polyamide surface morphology for NF membranes. (a1-a5) SEM top-viewed images. (b1-b5) TEM images. (c1-c5) AFM surface micrographs, where R_{RMS} is the root-mean-square surface roughness based on three independent tests.

Figure 2a shows the surface morphology of fabricated NF membranes (The SEM

image of UP150 substrate is shown in **Figure S3**). NF-control membrane, formed by PIP solution without pH adjustment, exhibited typical nodule-like surface nanostructures⁴⁴ (**Figure 2a1**). The adjustment of PIP pH for IP reactions resulted in relatively smaller nodule-like nanostructures on the membrane surface (**Figure 2a2-a5**). In addition, general trends of lower surface roughness (i.e., R_{RMS} obtained from AFM micrographs, **Figure 2c**) and reduced polyamide thickness (obtained from TEM micrographs, **Figure 2b**) were observed for NF membranes formed at lower pHs. These observations can be attributed to the varied dominant PIP species from the non-protonated ($\text{pH} > \text{p}K_{a2}$ of 9.7) to the protonated ($\text{pH} < \text{p}K_{a2}$) (**Figure 1d**). Specifically, the non-protonated PIP is highly reactive, which could trigger a relatively violent IP reaction for the polyamide formation^{25, 45} of NF-control (i.e., pH 11.6) and NF-pH10 membranes. Decreasing the pH values below $\text{p}K_{a2}$ resulted in a reduced fraction of highly reactive non-protonated PIP while an increased fraction of less reactive protonated species. This can lead to the lower reactivity of amine solution, which can slow down the IP reaction rate^{25, 45} and thus form the relatively smaller surface nanostructures and smoother surface morphology.⁴⁶

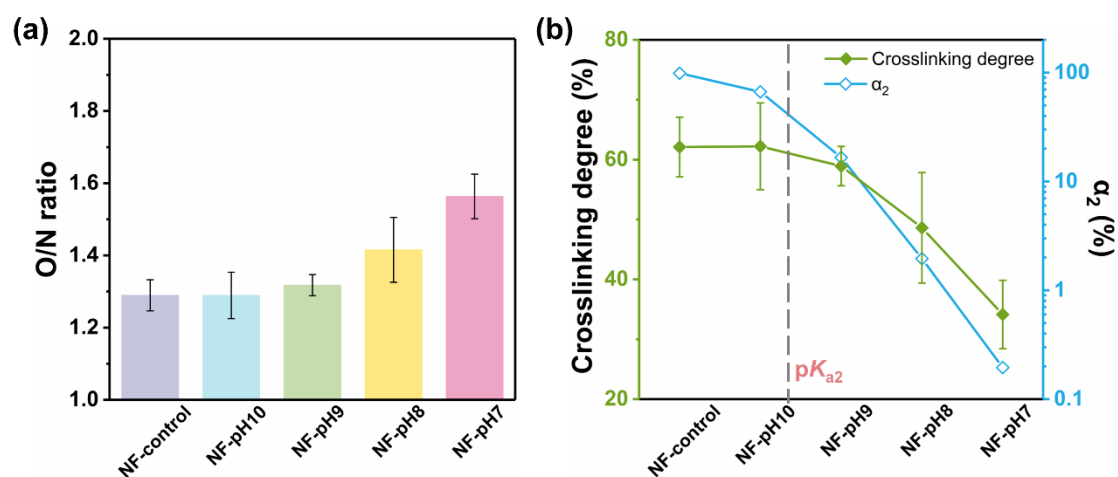


Figure 3. Polyamide physicochemical properties of NF membranes. (a) The ratio of oxygen to nitrogen (O/N ratio). The O/N ratio is calculated based on XPS results from at least three independent experiments. (b) Crosslinking degree of polyamide membranes based on at least three parallel measurements. α_2 is the distribution coefficient of non-protonated PIP (i.e., the fraction of non-protonated PIP over total dissolved PIP) in aqueous solution calculated based on Eq. 3.

Generally, the O/N ratio based on XPS surface analysis increased from 1.29 for NF-control to 1.56 for NF-pH7 membrane (**Figure 3a**), corresponding to a decreased crosslinking degree from 62% to 34% (**Figure 3b**). This trend of crosslinking degree was consistent with the reduced distribution coefficient (α_2) of non-protonated PIP (i.e., the fraction of non-protonated PIP) (**Figure 3b**), implying the critical role of the fraction of highly reactive PIP in polyamide formation. At $\text{pH} > \text{p}K_{a2}$ (i.e., NF-control and NF-pH10 membranes), non-protonated PIP is the dominant species in the amine solution (**Figure 1d**). These PIP molecules with high reactivity could lead to highly crosslinked polyamide (**Figure 1a**). In contrast, decreasing the amine pH below $\text{p}K_{a2}$ led to less crosslinked polyamide (**Figure 1b**), which can be due to the reduced α_2 accompanied by more mono-protonated species (**Figure 1d**). Further decreasing amine pH below

pK_{a1} resulted in ineffective crosslinked polyamide, as indicated by XPS results (**Table S3**). This could be attributed to the dominant di-protonated PIP with few non-protonation. Lower crosslinking degree often leads to a relatively looser polyamide structure with a larger pore size.⁴⁷ For example, the NF-pH8 membrane possessed a larger MWCO (around 500) than that (around 180) of the NF-control membrane (**Figure S6**), indicating its larger pore size. The pore size of membranes can be calculated according to the rejection value based on the log-normal distribution assumption. The estimated mean pore radius of NF-control and NF-pH8 was 0.28 nm and 0.38 nm, respectively (**Figure 4a**). In addition, less crosslinked polyamide features more residual acyl chlorides, which can be hydrolyzed to enhance the negative charge of polyamide surface. Consistently, the zeta potential values of the NF-pH8 membrane were generally lower than those of the NF-control membrane (**Figure 4b**). In respect to the other surface properties, these NF membranes possessed similar water contact angles (around 90°, **Figure S5**) and characteristic FTIR peaks (1620 cm^{-1} for amide group, **Figure S2**).

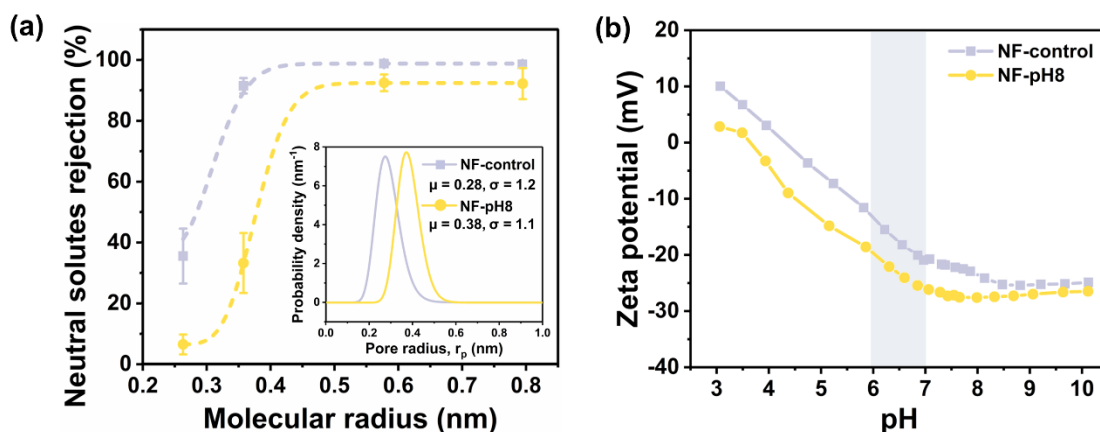


Figure 4. (a) Neutral solute (i.e., glycerol, glucose, raffinose, and dextran) rejections with different molecular weights (93.12, 180.16, 504.44, and 1000 Da) for NF-control and NF-pH8 membranes. The filtration tests of neutral solutes were repeated at least three times. The dash lines are established based on the log-normal distribution of membrane pore size. Inset is the pore size distribution of NF membranes estimated by neutral solutes rejection results. The mean pore radius and geometric standard deviation are also annotated. (b) Zeta potential analysis of NF-control and NF-pH8 membranes. Generally, the higher MWCO represents the larger mean effective pore size of polyamide.⁸

3.2 Membrane Separation Performance

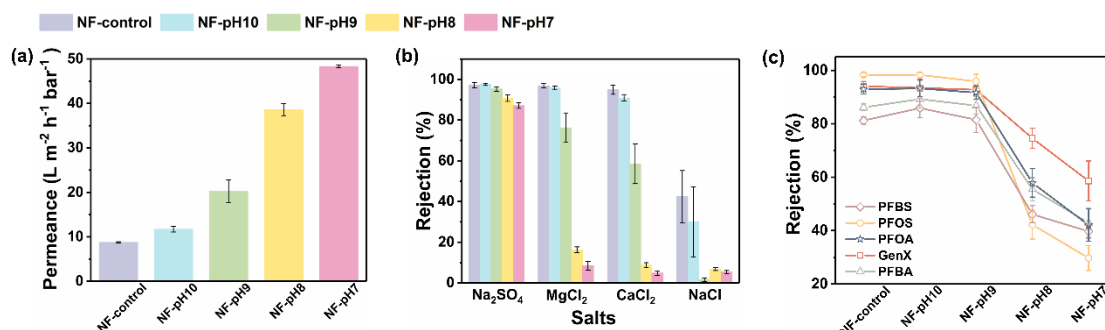


Figure 5. The separation performance of NF membranes. (a) Pure water permeance, (b) rejection of four salts, and (c) rejection of micropollutants. Filtration experiment conditions: applied hydraulic pressure is 5 bar and temperature is 25 °C. The membrane rejection of four salts was analyzed by the feed solution with a single type of salt dissolved (1000 ppm Na₂SO₄, MgCl₂, CaCl₂, and NaCl), respectively. The rejection test of micropollutants was conducted using a mixed solution containing 5 anionic perfluoroalkyl substances with a concentration of 200 ppb for each. All the results were based on at least three parallel measurements.

Generally, decreasing PIP pH for polyamide formation resulted in improved water

permeance of NF membranes (**Figure 5a**). For the control and NF-pH10 membranes, their water permeance was relatively lower. For other membranes formed at $\text{pH} < \text{p}K_{\text{a}2}$, the membrane permeance was significantly improved, up to $48.3 \pm 0.3 \text{ L m}^{-2} \text{ h}^{-1} \text{ bar}^{-1}$ of the NF-pH7 membrane. This significant improvement could be attributed to the obvious reduction of polyamide crosslinking degree at $\text{pH} < \text{p}K_{\text{a}2}$ (**Figure 3b**) and thinner polyamide layer (**Figure 2b**), which could greatly reduce the water transport resistance across the membrane.

All these NF membranes exhibited relatively high rejections of Na_2SO_4 (over 87%) while their rejections of NaCl were generally lower than 40% (**Figure 5b**). This can be attributed to the combined effects of size exclusion and charge repulsion.^{2, 47} NF membranes possessed stronger size exclusion for SO_4^{2-} than Cl^- due to the larger hydraulic radius of SO_4^{2-} . Besides, these membranes also gave enhanced electrostatic repulsion to divalent SO_4^{2-} than monovalent Cl^- by their negatively charged surfaces. Interestingly, the significantly reduced rejection of CaCl_2 was observed from 95% of NF-control to 5% of NF-pH7. A similar trend for the rejection of MgCl_2 was also observed (**Figure 5b**). These greatly reduced rejections of divalent cations could be attributed to the reduced crosslinking degree of polyamide formed at $\text{pH} < \text{p}K_{\text{a}2}$. The less crosslinked polyamide generally features a looser structure for weakened size

exclusion and more negative charge for enhanced charge interaction.⁴⁷ Both facilitate the passage of CaCl_2 and MgCl_2 . In addition, the membrane formed at $\text{pH} < \text{p}K_{\text{a}1}$ (i.e., NF-pH4) gave negligible salt rejection (e.g., 0.9 ± 0.3 % for Na_2SO_4 and 0.7 ± 0.3 % for CaCl_2) (Table S3) due to its ineffective-crosslinked polyamide. Furthermore, the rejection of micropollutants was reduced at lower pH (Figure 5c), which could be explained by the weakened size exclusion⁴⁸ resulting from the reduced crosslinking degree of polyamide (Figure 3b).

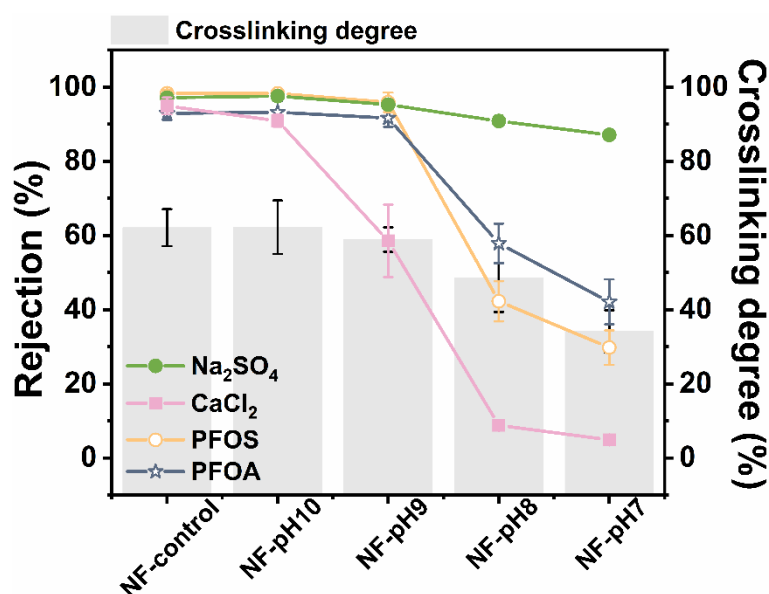


Figure 6. The rejection performance of Na_2SO_4 , CaCl_2 , PFOS, and PFOA by NF membranes. Na_2SO_4 and perfluoroalkyl substances (i.e., PFOS and PFOA) represent the divalent and monovalent anions, respectively. CaCl_2 represents the divalent cations. The rejection of Na_2SO_4 and CaCl_2 was measured using the feed solution with a single type of salt dissolved. The rejection of anionic perfluoroalkyl substances was measured with a feed solution containing a cocktail of PFOS, PFOA, GenX, PFBS, and PFBA in a NaCl background solution.

Generally, the solute rejection is influenced by the combined effects of size exclusion

and charge interaction.^{2, 8} To better elucidate the underlying rejection mechanisms, we compared the rejection performance of several solutes and polyamide crosslinking degree for these NF membranes (**Figure 6**). For the rejection of anions (i.e., Na₂SO₄, PFOS, and PFOA), disparate trends were observed. Specifically, the rejection of Na₂SO₄ remained high (87-97%). In contrast, the rejection of monovalent anionic PFOS and PFOA remained around 90% at pH \geq 9 and then decreased greatly at lower pH. This different rejection performance could be attributed to the competing effects of size exclusion and charge repulsion. As shown in **Figure 3** and **4**, decreasing the pH of the PIP solution weakens the size exclusion effect (due to increased membrane pore size) but enhances the charge interaction (due to the more negative membrane surface charge). In addition, the charge repulsion between divalent SO₄²⁻ and negative polyamide was stronger compared to that between monovalent anions (PFOA and PFOS) and polyamide.^{47, 48} For membranes formed at lower pH, the enhanced charge repulsion of Na₂SO₄ could play a dominant role to offset the weakened size exclusion effect, thereby maintaining high rejection. In contrast, the enhancement effect for charge repulsion is less pronounced for the monovalent anions PFOA and PFOS, which is less effective in offsetting the reduced size exclusion, leading to decreased rejection of these compounds at lower pH. For the divalent cations (CaCl₂), the rejection dramatically decreased from 95% to 5%. This sharp reduction can be explained by the

synergic effects of looser structure and more negative charge on the passage of divalent Ca^{2+} through the membrane.

3.3 Tailored Solute-to-Solute Selectivity of NF membranes

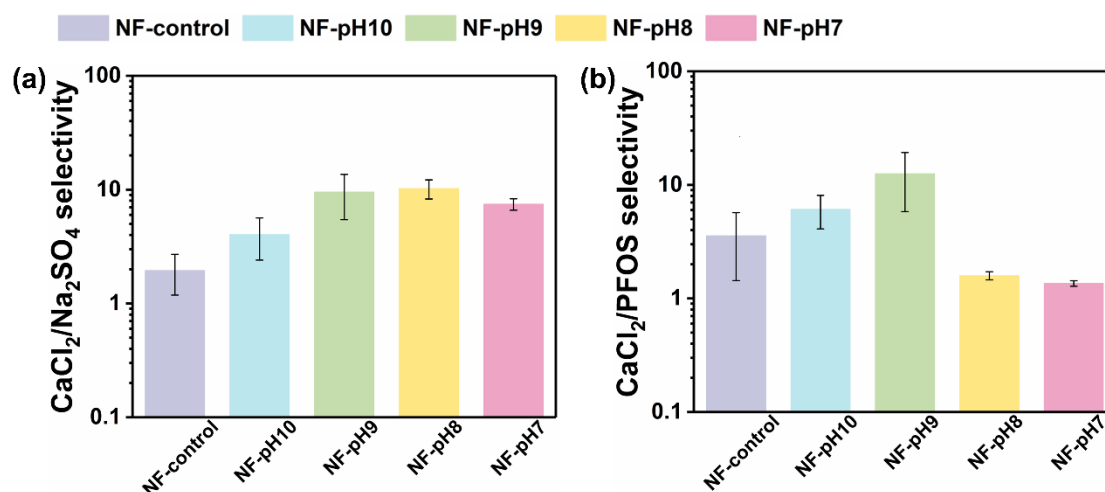


Figure 7. The solute-to-solute selectivity of NF membranes: (a) $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ and (b) $\text{CaCl}_2/\text{PFOS}$. The membrane selectivity is calculated based on the solute rejection.

Our results indicated that tailoring reactive sites on PIP by varying the dominant amine species could effectively regulate polyamide crosslinking degree to influence the rejection to different solutes. **Figure 7** and **Figure S7** further present different solute-to-solute selectivity. Generally, decreasing pH significantly improved $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ selectivity up to 10 (for NF-pH8), which was an order of magnitude higher than that of the NF-control membrane (**Figure 7a**). This significant improvement in membrane selectivity was attributed to the greatly reduced rejection of CaCl_2 and maintained high rejection of Na_2SO_4 (**Figure 6**). To further evaluate the NF membranes in this study, we

also included a benchmark in the form of an upper bound plot of $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ selectivity vs. water permeance based on the performance data of NF membranes reported in literature (**Figure 8**). NF-pH8 and NF-pH9 in this study showed comparable or even better $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ selectivity compared to many other membranes. In addition, the selectivity of $\text{CaCl}_2/\text{PFOS}$ was increased up to 12 (for NF-pH9), which was 3.5 times higher than that of the control membrane (**Figure 7b**). This obvious enhancement was attributed to the different rejection performance between CaCl_2 and PFOS (**Figure 6**).

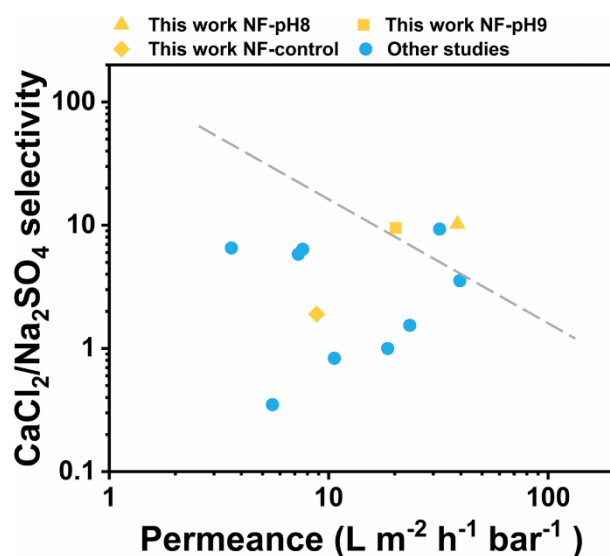


Figure 8. Comparison of $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ selectivity for prepared NF membranes vs. recent NF membranes in literature. Their specific separation performance data are shown in **Table S4**.

In the NF literature, it is commonly believed that the water permeance and solute-to-solute selectivity are key indicators to evaluate the membrane performance for

applications.¹ Here, we demonstrated the potential of tailoring PIP reactive sites to simultaneously enhance water permeance and membrane selectivity. To be noted, the choice of optimal membrane is greatly influenced by the specific application. For example, the NF-pH8 membrane featuring high $\text{CaCl}_2/\text{Na}_2\text{SO}_4$ selectivity (**Figure 7a**) could be applied for the desalination pretreatment to avoid scaling issue.³⁰ For the environmental applications involved in contaminant removal (e.g., PFOS removal from contaminated groundwater), the NF-pH9 membrane could be a better candidate due to its high rejection of contaminants (**Figure 5c**) and enhanced mineral/contaminant selectivity⁴⁸ (**Figure 7b**). For example, in the context of drinking water treatment, Ca^{2+} is crucial for human health. Therefore, enough minerals (i.e., low rejection to Ca^{2+}) are crucial for high-quality drinking water production in addition to high PFOS rejection.³⁴

4. Conclusions

This study clarified that the pH regulation of amine solution for varied PIP reactive sites can effectively control polyamide properties toward enhanced membrane performance.

The less crosslinked polyamide was formed at pH values between the two pK_a values (i.e., 5.3 and 9.7, respectively) of the diacidic base PIP, where mono-protonated PIP is dominant with the reduced fraction of non-protonated species. Such polyamide featured looser structure and more negative charge compared to the highly crosslinked NF-control formed at $pH > pK_{a2}$. The looser structure facilitated water transport through polyamide to significantly improve the water permeance (e.g., over 4 times higher than that of NF-control). Meanwhile, the more negative charge enhanced charge repulsion to anions and charge attraction to cations, leading to significantly higher solute-to-solute selectivity (e.g., an order of magnitude higher $CaCl_2/Na_2SO_4$ selectivity). These results suggested that precise regulation of monomer reactive sites could be a promising strategy to enhance water permeance and membrane selectivity of polyamide NF membranes. Future studies would further investigate if the reactive sites and monomer species could be regulated by other factors. Our work proposed a facile strategy to tailor the polyamide properties and membrane performance, providing novel insights into the future design and development of selective NF membranes toward diverse application scenarios.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at

Section S1, Synthesis of NF membranes; Section S2, Physicochemical properties of anionic perfluoroalkyl substances; Section S3, Characterization results; Section S4, Membrane selectivity.

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Notes

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