1	Genomic and chemical evidence for local adaptation in resistance to different herbivores in
2	Datura stramonium
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4	Running title: Local adaptation of plant resistance to herbivory
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24 Author contributions

25 Conceived and designed the experiments: JNF, IMDC. Performed the experiments: IMDC, JNF,

26 PLV. Analyzed the data: IMDC, JM, JNF. Contributed reagents/materials/analysis tools: IMDC,

27 JNF, CMFO, PLV, JM. Wrote the paper: IMDC, JNF, JM.

28

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47	Data availability
48	Scripts, commands and the entire workflow to obtain the Identity by Descent (IBD) can be
49	consulted in https://github.com/icruz1989/IBDcalculation. ddRad-seq data can be found in
50	DDBJ/ENA/GenBank under the BioProject PRJNA663170. Phenotype data can be found in
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55	
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71 Abstract

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73 Since most species are collections of genetically variable populations distributed to habitats 74 differing in their abiotic/biotic environmental factors and community composition, the pattern 75 and strength of natural selection imposed by species on each others' traits are also expected to be 76 highly spatially variable. Here, we used genomic and quantitative genetic approaches to 77 understand how spatially variable selection operates on the genetic basis of plant defenses to 78 herbivores. To this end, an F₂ progeny was generated by crossing *Datura stramonium* 79 (Solanaceae) parents from two populations differing in their level of chemical defense. This F₂ 80 progeny was reciprocally transplanted into the parental plants' habitats and by measuring the 81 Identity by Descent (IBD) relationship of each F₂ plant to each parent, we were able to elucidate 82 how spatially variable selection imposed by herbivores operated on the genetic background 83 (IBD) of resistance to herbivory, promoting local adaptation. The results highlight that plants 84 possessing the highest total alkaloid concentrations (sum of all alkaloid classes) were not the 85 most well-defended or fit. Instead, specific alkaloids and their linked loci/alleles were favored by 86 selection imposed by different herbivores. This has led to population differentiation in plant 87 defenses and thus, to local adaptation driven by plant-herbivore interactions.

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Key words. *Datura stramonium*, identity by descent, local adaptation, plant-herbivore
interactions, phenotypic selection, resistance.

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Introduction

96 Coevolution between plants and insects that feed on them is thought to be fueled by reciprocal 97 selection imposed by traits (or trait states) that mediate the interaction, potentially given rise to 98 arms races (Ehrlich and Raven 1964; Dawkins and Krebs 1979; Thompson 2005; Janz 2011). At 99 the microevolutionary scale, spatial environmental variation may result in a selection mosaic that 100 favors different traits (or trait states), hence promoting phenotypic and genetic/genomic 101 divergence among populations, and thereby local adaptation (Gomulkiewicz et al. 2002; 102 Thompson and Cunningham 2002; Thompson 2005; Briscoe Runquist et al. 2020). For instance, 103 plant populations are likely to encounter different communities of herbivores both in space and 104 time (Stam *et al.* 2014), making it highly improbable that selection pressures on plant defense 105 traits (e. g., chemical secondary compounds) would be homogenous across populations 106 (Berenbaum et al. 1986; Charlesworth 1998; Züst et al. 2012). Thus, it is expected that natural 107 selection on plant-herbivore interactions between environments can lead to population 108 differentiation of plant defense traits and ultimately to local adaptation. However, evidence of 109 how varying herbivore communities impose selection of phenotypic defense variation, and their 110 role in shaping the genomic constitution of populations is still scarce (Briscoe Runquist et al. 111 2020).

Local adaptation of plant defense against insect herbivores has been primarily studied using traditional quantitative genetic approaches such as common garden and reciprocal transplant experiments (Kawecki and Ebert 2004; de Villemereuil *et al.* 2016). These traditional approaches along with recent advances in genomics and mass spectrometry have made it possible to conduct detailed analyses of the genetic basis of chemical-based plant defense (Savolainen *et al.* 2013). For example, Identity by Descent analyses (IBD), genome-wide

118 association analyses (GWAS), quantitative trait loci mapping (QTL), or Fst vs. Ost comparisons, 119 provide methodologies to conduct in-depth studies on how plant chemical defense have evolved 120 in response to spatial variation in plant-insect interactions (Browning and Browning 2012; 121 Savolainen et al. 2013; Anderson et al. 2014; Flood and Hancock 2017). 122 In particular, IBD analysis estimates to what extent two or more individuals inherit a 123 similar nucleotide sequence from a common ancestor (Thompson 2013) and describes the degree 124 of genetic/familial similarity among a group of individuals (e. g., parents-offspring; Albrechtsen 125 et al. 2010; Thompson 2013). Thus, IBD can be used to evaluate whether the genetic background 126 of a plant is associated with its ability to face its herbivores. Furthermore, it also allows to detect 127 patterns of very recent or ongoing selection in the genome (Albrechtsen et al. 2010). For 128 instance, if insect herbivores are reducing the fitness of individual plants, one might suppose that 129 more resistant plants to herbivory will produce more progeny than less resistant plants (Núñez-130 Farfán et al. 2007). If so, then ongoing natural selection will increase, across generations, the 131 amount of IBD sharing in a population in the region surrounding the allele(s) that confer(s) 132 resistance to herbivory (Browning and Browning 2012). The reasoning behind this is that as a 133 positively selected allele increases in frequency, the region containing the resistance allele will 134 increase in homozygosity and experience less intra-allelic recombination at the population level 135 (Albrechtsen et al. 2010). While IBD analysis has been used to identify how recent or ongoing 136 selection operates on human diseases caused by pathogens (Albrechtsen et al. 2009; 2010; 137 Daniels et al. 2015; Wong et al. 2017; Henden et al. 2018), to best of our knowledge, no studies 138 have used this approach to evaluate how the genetic background of plant resistance to herbivory 139 is driven by natural selection.

140 The main aim of this study was to assess the extent to which the evolution of plant 141 defenses to insect herbivores has been driven by natural selection. To this end, we generated an 142 F_2 progeny derived from the cross between two populations of the annual herb *Datura* 143 stramonium (Asteridae; Solanaceae), known to differ in their level of chemical defense and 144 herbivore community (De-la-Cruz et al. 2020). The F₂ plants were reciprocally transplanted to 145 the natural environments (populations) of the grandparents. In this way, we were able to 1) 146 determine the level of infestation and damage exerted by different herbivores on plants sowed in 147 each locality, 2) to determine whether the seven most abundant constitutive alkaloids of D. 148 stramonium are linked to the level of herbivore infestation. 3) by estimating the Identity by 149 Descent (IBD) relationship of each F₂ plant to each grandparent, we were able to evaluate 150 whether genomic similarity to either of the grandparents predicts survival/fitness and resistance 151 to herbivores in each experimental site. Finally, 4) by quantifying the strength of natural 152 selection on plant defense traits in the two experimental sites, we assessed whether natural 153 selection favors an increase in plant resistance against herbivores in each of the two study sites. 154 155 **Materials and Methods** 156 The study sites 157 The two study sites, Teotihuacán (State of Mexico, 19°41'6.96"N, 98°52'19.63"W) and Ticumán 158 (State of Morelos, 18°45'39.90"N, 99° 7'13.86"W), were selected for four main reasons. First, 159 the two populations occur in different habitats with distinct climatic characteristics (xerophytic 160 shrub and tropical dry forest, respectively; Valverde et al. 2001). Second, species of herbivores 161 that infest upon D. stramonium differ between the sites (see also Results); in Ticuman, D. 162 stramonium is attacked mainly by the specialist flea beetle *Epitrix* sp. (Valverde *et al.* 2001;

163	Fornoni et al. 2004), whereas in Teotihuacán it is consumed mainly by the specialists beetles
164	Lema daturaphila, Epitrix parvula and the specialist seed predator Trichobaris soror (Bello-
165	Bedoy and Núñez-Farfán 2010; Miranda-Pérez et al. 2016). In fact, L. daturaphila is absent in
166	the Ticumán population. Third, low degree of genetic differentiation at neutral loci has been
167	detected between the two populations ($F_{ST} = 0.06$; De-la-Cruz <i>et al.</i> 2020). Finally, the two
168	populations are highly differentiated in their level of tropane alkaloid concentrations (De-la-Cruz
169	et al. 2020). Previous evidence indicates that tropane alkaloids are central for resistance against
170	herbivores in this species (Shonle and Bergelson 2000; Miranda-Pérez et al. 2016). Indeed, at
171	least the tropane alkaloid scopolamine has been implicated as a defense against herbivores in one
172	population (Castillo et al. 2014).

173

174 Experimental design

175 To produce the F₂ generation progeny for this study, we randomly collected fruits from 45 and 176 47 different plants from the Teotihuacán and Ticumán sites, respectively. Ten seeds from each of 177 the 92 plants were soaked in water containers and maintained in an environmental chamber at a 178 photoperiod of 12:12 L:D, and at a temperature of 30°C during the day and 25°C at night, at 179 constant humidity of 85%. Seeds were scarified to facilitate germination (Fornoni et al. 2000). 180 Germinated seeds were transferred to plastic pots (237 ml) and randomly allocated to positions 181 on benches in the greenhouse. When the first true leaves developed, each plant was transplanted 182 into 10 L plastic pots filled with a 1:1 mix of sand and vermiculite, and again, the pots were 183 placed randomly on the benches. Each plant received the same daily quantity of water (500 ml) 184 during the entire experiment. When the plants reached the flowering stage, flowers were hand 185 pollinated. Plants from Teotihuacán were used as pollen receptors and plants from Ticumán were

used as pollen donators. Prior to manual pollination, flowers from Teotihuacán were emasculated
before dehiscence and covered with bags to avoid pollen contamination from other plants.

188 Cross-pollination was achieved by rubbing anthers of pollen donors onto the stigma of a 189 flower. Mating pairs were set at random. After pollination, flowers were tagged and bagged. 190 Because a plant can produce several flowers, each flower could be pollinated by different pollen 191 donors. Thus, we produced ca. 200 crosses. Fruits of each cross (F₁ generation progeny) were 192 tagged and collected in paper bags and stored at room temperature. When plants reached the 193 flowering stage and second bifurcation (~30 days after planting in pots), 6-8 leaves from each 194 plant were harvested to quantify the diversity and concentration of tropane alkaloids. There is 195 evidence that the highest concentration of tropane alkaloids in D. stramonium occurs at the 196 flowering stage, which is related to the timing of infestation by the main herbivores of D. 197 stramonium (Kariñho-Betancourt et al. 2015). A total of 21 tropane alkaloids were identified and 198 analyzed for all parental plants using methods described in De-la-Cruz et al. (2020). 199 Once the total tropane alkaloid concentration of the parental plants was completed, we 200 selected the individual plant with the lowest (Teotihuacan) and the highest (Ticumán) 201 concentration of tropane alkaloids (grandparents Teotihuacán 1 and Ticumán 23) (S1). These 202 plants differed 58-fold in their total alkaloid concentration (1,013 vs. 59,000 ng/g of leaf, 203 respectively) (S1). F_1 seeds derived from the cross between these two parental plants were 204 sowed, following the procedure described above, to produce the F₂ progeny (single family: S1). 205 To this end, we used seeds from three fruits of the same crossing. From the germinated F_1 plants 206 (n = 8), we randomly chose one plant whose flowers were bagged to avoid pollen contamination 207 from other plants (although plants were grown in a glasshouse; S1). We allowed this F1 208 individual to self-pollinate to produce the F₂ generation progeny (S1).

209

210 Transplant experiment in the two sites

211 Experiment. F2 seeds, taken randomly, were germinated and grown in the greenhouse as 212 described above. When the two true leaves appeared, F_2 seedlings (n = 430) were transplanted to 213 experimental plots in Teotihuacán (n = 230) and Ticumán (n = 200) in order to expose the F_2 214 plants to the local herbivores and natural environmental conditions of their grandparents (S1). 215 During the first days after transplanting, high seedling mortality occurred at the tropical site 216 (Ticumán), reducing sample size to 103 plants. In each site, seedlings were planted in the 217 experimental plot according to a complete randomized design. Plants were spaced 1 m apart in a 218 regular grid. Experimental plots were regularly weeded to prevent interference and competition 219 by other species.

220 Damage by herbivores. Leaf damage to plants by herbivores was measured with the mobile

221 application BioLeaf (Machado et al. 2016) on four sampling periods (15, 30, 45, 60 days after 222 planting). On each sampling date, we took photographs of eight randomly chosen full expanded 223 leaves per plant using a mobile phone (Samsung Galaxy S6 edge). The app automatically 224 calculates the injured leaf regions caused by insect herbivory and then estimates the damage (in 225 percentage) relative to the total leaf area (Machado et al. 2016). Thus, we were able to quantify 226 the damage inflicted by herbivores to the plants during the experiment. Likewise, the average 227 proportion of leaf damage by herbivores per plant was obtained. However, it is important to 228 highlight that in the Teotihuacán site, most leaf tissue was completely eaten by herbivores in 229 many plants. In these cases, we assigned 100% of the damage to these plants. 230 Herbivore infestation. At the Teotihuacán site, three species of herbivores were recorded during

three sampling periods (15, 30, 45 days after planting). In each plant, we counted the number of

232 1) adults *Epitrix parvula*, 2) adults *Lema daturaphila*, 3) larvae of *Lema daturaphila*, and 4) 233 adults Trichobaris soror. Since larval development and pupation of E. parvula occur in the soil, 234 we were unable to record these stages. Therefore, only the number of adults on plants was 235 obtained for this insect species, as well as for T. soror. To minimize bias in insect counting, only 236 one person counted the herbivores on each plant in all the sampling periods. In the Ticumán site, 237 we recorded the infestation accounted only by *Epitrix* sp., since L. daturaphila is absent and T. 238 soror is very rare (only 3 individuals registered at this site). At the end of the experiments, we 239 had a measurement of total infestation that each plant experienced by each herbivore in both 240 sites.

Leaf tissue sampling. In order to determine alkaloid concentration, we collected one leaf (10 cm in length) per plant when plants reached their second bifurcation and were flowering (~25 days after sowing). The leaf sampled was packed in aluminum foil, labeled and immediately frozen in liquid nitrogen. All samples were transported and stored in a freezer at -80°C. In order to obtain DNA from each F_2 plant for genetic analyses, one additional leaf was collected, frozen and stored as described above.

Plant survival and reproduction. In the Teotihuacán site, plant mortality was caused by heavy
damage exerted by insects (n = 66). We recorded plant survival as a nominal variable
(dead/alive). At the Ticumán site, however, there was no record of single plant mortality due to
damage exerted by herbivores.

At the end of the experiment (two months after sowing), we collected all fruits produced by each plant in each experimental site. Fruits were bagged individually and labelled. In the lab, seed set per fruit was counted and total number of seeds per plant was used as an estimator of

254 maternal plant fitness (*see* statistical analyses section; Motten and Antonovics 1992; Núñez255 Farfán *et al.* 1996; Mauricio and Rausher 1997).

256

257 Alkaloid extraction of F₂ plants

258 In order to extract tropane alkaloids from each plant, frozen leaf tissue was first transferred to 2 259 mL Eppendorf tubes, grinding it with a plastic pestle while keeping it frozen by adding liquid 260 nitrogen. Second, we weighted the pulverized frozen leaf tissue in Eppendorf tubes. Third, we 261 added two steel balls to each Eppendorf tube along with 1.5 mL of extraction buffer (80% 262 methanol; MeOH and 1% formic acid); the tubes were then shaken for 60 s at 30 Hz in a 263 TissueLyser II (QIAGEN). Finally, the samples were centrifuged for 20 min at 14,000 rpm; 700 264 µL of supernatant was collected and stored in glass vials (1.5 mL) and maintained at -4°C until 265 quantified in the Liquid Chromatography/Time-of-Flight/Mass Spectra (HPLC-TOF-MS).

266

267 Liquid Chromatography/Time-of-Flight/Mass Spectra

268 Before analysis, 300 µL of MeOH was added to each sample (stored in a glass vial; *see* above)

and then injected into an Agilent 1260 Infinity, coupled to an Accurate-Mass Time-of-Flight

270 (TOF) LC/MS-6230, with an auto-sampler Agilent Technology 1200 Infinity. The

271 chromatographic separations were performed in a HPLC Agilent ZORBAX column. Before

samples were injected into the column, it was cleaned with 15 mL of MeOH. For this, a gradient

273 of mobile phase A (1% (v/v formic acid in water) and mobile phase B (1% (v/v formic acid in

274 methanol) were used. The gradient profile was set to 0.00 min 90% A eluent, 10 min 10% A

eluent, 17 min 90% A eluent, 17.10 min 90% A eluent. Conditions of this last step were

maintained for 5 min to balance the column. The flow rate was 0.200 μ L 1 min⁻¹ each 5 min, so

each sample was analyzed for 23 min, and the column temperature was 50°C. The injection volume was 1 μ l for all samples. The electrospray source (ESI) was operated in the positive mode, and the interface conditions were as follows: the fragmentor of 200 V; Skimmer 65 V; oct 1 RF Vpp 750 V; gas temperature of 350°C; drying gas flow rate of 6 L min⁻¹; the nebulizer

worked at 50 psig. The ions of the compounds and their retention times are given in S2.

To standardize the method and optimize the detection of alkaloids in the HPLC-TOF-MS system, we prepared standard solutions (1:1000; mg/ml) of Atropine and Scopolamine (Sigma-Aldrich, St. Louis, MO, USA) of MeOH and injected these at volumes of 2, 4 and 8 µl. Since atropine showed a better calibration curve, we used this curve to calculate the concentration for each identified alkaloid per plant.

287

288 Identification and quantification of alkaloids in *D. stramonium* leaves

First, we identified the seven most abundant constitutive alkaloids in *D. stramonium*: four

290 tropane alkaloids (atropine, scopolamine, 3-hydroxy-6-tigloyloxytropane and anisodamine; De-

291 la-Cruz et al. 2020), one alkaloid derived from the phenylalanine biosynthesis

292 (phenylacetaldehyde), one pyrrolizidine alkaloid (pyrroline), and one triterpenoid of unknown

293 name but of similar structure and molecular weight to azadirone triterpenoid (Álvarez-Caballero

and Coy-Barrera 2019) (S2). Each alkaloid was searched and integrated (peak integration)

295 individually in each chromatogram of each plant. The MassHunter Workstation software (v. B.

296 06.00; Agilent Technologies) was used to identify the alkaloids using data of mass spectra,

retention time, and molecular formula obtained in the chromatograms (S2). The total

298 concentration for each alkaloid per plant was obtained using the slope and the intersect from the

299 regression equation of the calibration curve (curve from atropine standard):

300
$$\left(\left(\frac{(a + bX) x \ 1000}{d}\right) x \ 1000\right)$$

301 where *a*, is the intercept obtained from the regression of the calibration curve; *b*, is the slope 302 obtained from the regression of the calibration curve, *X* is the concentration of given alkaloid in 303 each plant and *d* is the dry weight of the sample. Alkaloid concentration was expressed in $\mu g/g$ 304 units of leaf weight. Total alkaloid concentration was obtained as the sum of the seven alkaloids 305 per plant (Kariñho-Betancourt *et al.* 2015).

306

307 DNA extraction, library preparation for ddRad- sequencing

308 Genomic DNA (gDNA) was extracted from 163 individuals planted in Teotihuacán and 51 309 individuals planted in Ticumán. Since we had high mortality of seedlings at the beginning of the 310 experiment in Ticumán, we extracted DNA from more individuals sowed in Teotihuacán. gDNA 311 was isolated from fresh leaves with a modified CTAB mini-prep protocol for ddRad-seq (Doyle and Doyle 1987). The total amount of gDNA was measured using Qubit dsDNA HS Assay Kit 312 313 (Invitrogen, Thermo Fisher Scientific, Waltham, USA). A total of 200 ng of gDNA was used for 314 library preparation. The qualified DNA samples were digested with EcoRI and Hin1II (NlaIII) 315 restriction enzymes (Takara, Osaka, Japan) and subjected to adapter ligation. The digestion and 316 ligation were performed at 37°C for 16 hrs. The ligation products barcoded with unique P1 317 adapter were pooled and purified by size selection using E-Gel SizeSelect 2% agarose (Life 318 Technologies, Carlsbad, CA, USA). Approximately 400-600 bp fragments were retrieved. The 319 selected size and adaptor-ligated DNA was subsequently amplified by PCR. The PCR products 320 were purified using AMpure XP beads (Beckman Coulter, Brea, CA, USA). The purified library 321 was sequenced using Illumina Hiseq X Ten platform (Illumina, San Diego, CA, USA). Library 322 preparation and sequencing were carried out by CD Genomics company (Shirley, NY, USA). For 323 the two grandparents, gDNA was isolated and measured as above. However, whole genome 324 sequencing was carried out for both, rather than ddRad-seq. Libraries were sheared on the 325 Covaris and then prepped for 150PE (paired-end) Illumina HiSeq 4000 sequencing using the 326 Kapa Hyper prep Illumina library prep kits. Final libraries were visualized on the Agilent 327 Fragment Analyzer, then quantified and pooled at equimolar amounts with Kapa qPCR Illumina 328 library quant Universal Kits. The sequencing and library preparations for the grandparents were 329 carried out in the QB3 Functional Genomics and Vincent J. Coates Sequencing Laboratories at 330 the University of California, Berkeley.

331

332 Identity by Descent (IBD)

333 Two haplotypes are identical by descent (IBD) if they share the same alleles inherited from a 334 common ancestor (Thompson 2013). Thus, closely related individuals have a high proportion of 335 IBD (Thompson 2013). We estimated IBD between each individual F₂ plant and each of the two 336 grandparents (214 F_2 plants vs. grandparent from Teotihuacán/grandparent from Ticumán). This 337 information was used to evaluate whether F₂ plants more related to a given grandparent (*i. e.*, 338 grandparent from Ticumán or grandparent from Teotihuacán) were more or less resistant to 339 herbivory or had higher or lower fitness/survival in the experimental sites (see below). 340 For IBD estimation, demultiplexing was performed with the Illumina bcl2fastq v2.19

software, which returned sequence data in fastq format for each individual. Barcodes and indexes had been removed previously by CD Genomics and QB3 services. Illumina reads were trimmed using a Phred quality score > 20 in TRIMMOMATIC v0.32 (Bolger *et al.* 2014). We visually verified the quality of the grandparents and some individuals (~80) before and after trimming with FastQC (Andrews 2010). This allowed us to keep only high-quality reads for IBD analyses.

346	Sequences of each individual were aligned to the Datura stramonium reference genome (De-la-
347	Cruz et al. under review; available in NCBI, BioProject PRJNA622882, biosample
348	SAMN14531593, accession JAAWWY000000000) using the BWA v0.7.17 software (Li and
349	Durbin 2009) with default parameters. SAM files from BWA were converted to BAM format
350	and these BAM files were sorted using SAMtools v1.10 (Li et al. 2009).
351	The calculation of IBD values between each individual in relation to the grandparents
352	was based on the genotype posterior probabilities (GPP) of each individual (Li et al. 2009;
353	Rastas 2017). These GPPs were calculated using SAMtools mpileup (Li et al. 2009) and custom
354	scripts provided in the tutorial of the Lep-MAP3 program (Rastas 2017;
355	https://sourceforge.net/p/lep-map3/wiki/LM3%20Home/). These scripts also account for the
356	alignment quality and filtering. The Lep-MAP3 program (Rastas 2017) was then used to
357	calculate the IBD values between each individual and their grandparents.
358	
359	Statistical analysis
360	All statistical analyses were performed using the JMP statistical package (v. 15.0; SAS Institute).
361	Plotting was made using ggplot2 (Wickham 2016) in RStudio version 1.1.463 (R Core Team
362	2020).
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364	
	Damage, herbivore infestation and concentration of alkaloids
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365 366 367	Damage, herbivore infestation and concentration of alkaloidsEstimates of herbivore damage, herbivore infestation and alkaloid concentration were natural logtransformed (log n + 1) to meet normality assumptions. An ANOVA was performed to test fordifferences in the levels of infestation by different herbivore species. A repeated measures

experimental site. To assess the severity of damage as a function of the total infestation rate by
each species of herbivore in each of the two populations, a Pearson correlation analysis (Zar
1999) was performed between the average damage and the total infestation by each herbivore in
each population.

373

374 IBD and survival in Teotihuacán

To analyze if genomic similarity among the F₂ individuals to their Ticumán and Teotihuacán grandparent was associated with their survival probability (alive/dead), two-tailed *t*-tests were used to compare mean IBD of individuals that survived or died. This analysis was only carried out for plants grown at the Teotihuacan site, because there was not plant mortality due to damage exerted by herbivores at the Ticumán site (*see* above).

380

381 Relationships between resistance, IBD and herbivory

382 Prior to analyses, all variables were standardized to a mean of zero and a standard deviation of 383 one (x = 0, SD = 1). Generalized linear models (GLMs) were employed to evaluate the 384 relationships between resistance and IBD and herbivory. The GLMs described hereafter, were 385 selected based on the statistical significance of the model and on the lowest corrected AIC 386 values, *i. e.*, models that best explained the relationship between the variables (Akaike 1974). First, general plant resistance (R_i) of the plant *i* was defined as $R_i = 100$ -def, where def is 387 388 the average proportion of leaf damage experienced by each plant (Núñez-Farfán and Dirzo 389 1994). To evaluate the relationship between resistance and IBD, two GLMs (link = identity, 390 distribution = normal) were constructed; one using the IBD values between F_2 plants and the 391 Teotihuacán grandparent, and the other using the IBD values between F₂ plants and the Ticumán 392 grandparent. In these models, the response variable was resistance, whereas IBD, the 393 experimental site, and their interaction were used as predictors. Adding the interaction between 394 experimental site and the covariate in the models allowed us to assess whether the effect of the 395 IBD to each one of the grandparents (Teotihuacán or Ticumán) differed depending on the site of 396 testing (*cf.* Zar 1999).

397 Since herbivore species differed between the sites, we independently assessed resistance
398 as a function of herbivore species by GLMs (link = identity, distribution = normal). In

399 Teotihuacán, we used the abundance of adults and/or larvae of *L. daturaphila*, *T. soror* and *E.*

400 *parvula* as covariates, whereas in Ticumán, only the abundance of *Epitrix* sp. (the only herbivore
401 detected in this site) was used as a predictor.

402

403 **Relationship between herbivore infestation and alkaloid concentration**

404 To assess the effect of the alkaloids on herbivore infestation, we also carried out GLMs (link = 405 identity, distribution = normal) in which the response variables were *E. parvula*, *Epitrix* sp. 406 adults or larvae of L. daturaphila or T. soror abundances on plants. The predictors in these 407 models were the concentrations of the seven alkaloids. In addition, we performed stepwise 408 GLMs (link = identity, distribution = normal) following a backward selection, which starts with 409 all predictors in the model (seven alkaloids), and iteratively removes the least contributive 410 predictors (Sokal and Rohlf 1994; Zar 1999). This allowed us to detect which alkaloid 411 configuration had a greater positive or negative effect (or both) on the infestation of each 412 herbivore species. The best GLMs were selected based on the statistical significance of the 413 model and on the lowest corrected AIC values (Akaike 1974). An additional GLM (link = 414 identity, distribution = normal) with the total alkaloid concentration as a predictor was carried

415 out to see the impact of total alkaloid concentration on herbivores. Tests for the interaction 416 between experimental site and one particular herbivore were not possible because different 417 species were present in the two sites.

418

419 Natural selection on alkaloids, resistance, herbivore infestation and IBD

420 To quantify the magnitude and direction of natural selection acting on the seven alkaloids, we 421 used the number of seeds produced by each plant as a fitness proxy to perform phenotypic-422 selection analyses (Lande 1979; Lande and Arnold 1983). For this purpose, standardized 423 individual fitness (relative fitness) was calculated as $w_i = x_i/x$, where x_i is the total number of 424 seeds produced per plant, and x is the average number of seeds per plant in the population in 425 each site. In all analyses, w_i was used as a response variable. Thus, one GLM (link = identity, 426 distribution = normal) was constructed using the concentrations of seven alkaloids, the 427 experimental site, and their interactions. An additional two separate GLMs were constructed 428 using resistance and total alkaloid concentration as predictors, as well as experimental site as a 429 factor. Interactions between site and predictors allowed us to test if the effects of predictors on 430 fitness differed between the two sites.

Two separate models, one for each experimental site, were carried out to assess selection on the infestation by each herbivore (independent variables). As pointed out earlier, we could not evaluate the effect of the experimental site and its interaction with predictors, since different species of herbivores were present in the two populations.

Finally, to evaluate the effects of identity by descent (IBD) on fitness (seed production),
two GLMs were constructed (one using the IBD values between F₂ plants and the Teotihuacán
grandparent, and the other using the IBD values between F₂ plants and the Ticumán

438 grandparent). In these models, the response variable was relative fitness, whereas IBD,

439 experimental site and its interaction were predictors.

440 The generalized linear coefficients (*i. e.*, the selection gradients; β_i , Lande and Arnold 441 1983) obtained from the selection analyses represent the strength and direction of selection 442 acting directly on each alkaloid, resistance, infestation by each herbivore and IBD in comparable 443 units (standard deviations; Wise and Rausher 2013).

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Results

446 Damage, herbivore infestation and alkaloid concentrations in the two experimental sites

447 Damage by herbivores varied between sampling dates in each site (Teotihuacán: $F_{710} = 110.98$,

448 $R^2 = 0.41, p = 0.0001$; Ticumán: $F_{262} = 27.16, R^2 = 0.27, p = 0.0001$; S3 a, b, S4). There were

449 clear differences in level of infestation by the different species of herbivores ($F_{545} = 215.32$, $R^2 =$

450 0.61, p = 0.0001; S3 c, S4). Correlation analyses indicated that plant damage in Teotihuacán site

451 was mainly imposed by larvae of L. daturaphila, whereas that in the Ticumán site mainly by

452 *Epitrix* sp. (S5).

453

454 Effect of the Identity by Descent (IBD) on fitness/survival and on resistance

455 The GLM between fitness and the IBD to the Ticumán grandparent as measured by genome wide

456 IBD was significant, revealing a positive effect of increasing IBD on fitness (L-R chi-square₃ =

457 12.91, AICc = 273.68, p = 0.0048, Table 1, Fig. 1 a, b). The model between fitness and the IBD

458 with the Teotihuacán grandparent was not significant (L-R chi-square₃ = 2.68, AICc = 283.90, p

459 = 0.4424, Table 1, Fig. 1 c, d). Our results also showed that plants more related to the Ticumán

460 grandparent had higher survival than F2 plants less related to the Ticumán grandparent in the

Teotihuacán site ($F_{129} = 17.52$, $R^2 = 0.12$, p = 0.0001; Fig. 1 e). In contrast, plant survival was not 461 significantly associated with IBD to the Teotihuacán grandparent in Teotihuacán ($F_{129} = 1.92, R^2$ 462 = 0.014, p = 0.1682; Fig. 1 f). The effect of IBD on plant survival in the Ticumán site was not 463 464 evaluated because there was not plant mortality due to damage exerted by herbivores in this site 465 (see above). The mean F₂ full-sibs relatedness (IBD) was 0.47 (range 0.006-0.803, standard error 466 = 0.0007). Identity by descent between the F₂ plants to each grandparent range between 0.006-467 0.5 (Teotihuacán grandparent) and 0.031-0.5 (Ticumán grandparent). Relatedness between the 468 two grandparents was zero (Fig. 1 g). 469 The GLM between resistance and the IBD with the Ticumán grandparent was significant 470 $(L-R \text{ chi-square}_3 = 54.21, AICc = 441.47, p = 0.0001)$. Significant effects included population, 471 and the interaction between population and IBD to the Ticumán grandparent (positive 472 relationship in Teotihuacán site, while an opposite effect was observed in the Ticumán site; 473 Table 2, Fig. 2 a, b). The GLM between resistance and the IBD with the Teotihuacán 474 grandparent was significant (*L-R* chi-square₃ = 32.50, AICc = 463.18, p = 0.0001). However, 475 only the population effect was significant (Table 2, Fig. 2 c, d). 476 477 Resistance against herbivore infestation levels in the two experimental sites 478 Resistance to herbivory was significantly related to herbivore infestation levels in Teotihuacán 479 (*L-R* chi-square₄ = 111.09, AICc = 426.37, p = 0.0001). Resistance was only positively related to 480 levels of E. parvula infestation and negatively related to infestation by larvae of L. daturaphila 481 in Teotihuacán (Table 2, Fig. 2 e, f). However, resistance and *Epitrix* sp. infestation levels were 482 negatively related in Ticumán (*L-R* chi-square₁ = 14.96, AICc = 223.89, p = 0.0001; Table 2, Fig. 483 2 g).

485	Effect of the alkaloid concentration on herbivore infestation levels
486	The effect of the seven alkaloid concentrations on <i>E. parvula</i> infestation level was significant (<i>L</i> -
487	<i>R</i> chi-square ₇ = 15.67, $AICc = 444.78$; $p = 0.0282$). However, the effects of individual alkaloids
488	were different: triterpenoid had a positive significant effect, whereas negative significant effects
489	were detected in case of phenylacetaldehyde and pyrroline (S6). The effect of the total alkaloid
490	concentration on <i>E. parvula</i> infestation was negative and significant (<i>L-R</i> chi-square ₁ = 3.91 ,
491	<i>AIC</i> c = 462.23; <i>p</i> = 0.0479; S6).
492	The effect of alkaloid concentrations on infestation levels by larvae of L. daturaphila was
493	significant (<i>L-R</i> chi-square ₅ = 12.53, $AICc = 460.49$, $p = 0.0281$). Significant effects included
494	phenylacetaldehyde (positively) and the triterpenoid (negative effect; S6). The GLM of
495	infestation levels by larvae of L. daturaphila against the total alkaloid concentration was not
496	significant (<i>L-R</i> chi-square ₁ = 1.45, <i>AIC</i> c = 473.76; <i>p</i> = 0.2272; S6).
497	The effect of alkaloid concentration on infestation levels by adults of L. daturaphila was
498	significant (<i>L-R</i> chi-square ₃ = 9.49, $AICc = 453.93$; $p = 0.023$). In this model we observed that 3-
499	hydroxy-6-tigloyloxytropane was significantly and negatively related with L. daturaphila adults,
500	whereas atropine showed a significant positive relationship (S6). The GLM of infestation levels
501	by adults of <i>L. daturaphila</i> against the total alkaloid concentration was also not significant (<i>L-R</i>
502	chi-square ₁ = 0.49, <i>AIC</i> c = 469.65; <i>p</i> = 0.4797; S6).
503	The GLM testing for the effects of alkaloid concentration on <i>T. soror</i> infestation rate was
504	significant (<i>L-R</i> chi-square ₄ = 10.95, $AICc = 467.18$, $p = 0.027$). In this model, <i>T. soror</i>
505	infestation level was negatively related to scopolamine concentration, but positively related to
506	the concentration level of the triterpenoid (S6). The effect of the total alkaloid concentration on

T. soror infestation was also not significant (*L-R* chi-square₁ = 0.11, *AIC*c = 478.92; *p* = 0.7367;
508 S6).

509	The GLM between <i>Epitrix</i> sp. infestation levels and the alkaloid concentrations in
510	Ticumán was significant (<i>L-R</i> chi-square ₇ = 17.89, $AICc = 134.49$, $p = 0.0125$; S6). Significant
511	effects included 3-hydroxy-6-tigloyloxytropane (negative effect) and pyrroline (positive effect;
512	S6). The effect of the total alkaloid concentration on <i>Epitrix</i> sp. infestation was not significant
513	$(L-R \text{ chi-square}_1 = 0.70, AICc = 141.09; p = 0.4000; S6).$
514	
515	Natural selection on resistance, alkaloids and herbivore infestation in the two experimental
516	sites
517	The GLM of relative fitness against resistance to herbivory was significant (L-R chi-square ₃ =
518	20.77, AICc = 487.04; $p = 0.0001$). While there was no main effect of the population, the
519	population \times resistance interaction was significant, revealing that the relative fitness was
520	positively related to resistance in Teotihuacán, but negatively in Ticumán (Table 1, S7 a, b).
521	The GLM of relative fitness as a function of the concentration of seven alkaloids was
522	significant (L-R chi-square ₁₅ = 26.14, AICc = 400.98; $p = 0.036$). There was a significant
523	positive main effect of the pyrroline concentration on fitness (Table 1), but the experimental site
524	\times pyrroline interaction was also significant, revealing a negative trend between fitness and
525	pyrroline concentration in Teotihuacán, whereas the opposite was true in Ticumán (Table 1, S8).
526	Similarly, the significant experimental site \times triterpenoid interaction revealed that fitness was
527	positively related to triterpenoid concentration in Teotihuacán, but with a negative trend in
528	Ticumán (Table 1, S8). The GLM of relative fitness against total alkaloid concentration was not

significant in either of the populations (*L-R* chi-square₃ = 2.86, AICc = 418.28; p = 0.41; Table 1).

The GLM of relative fitness against level of herbivore infestation (larvae and adults of *Lema, E. parvula* and *T. soror*) was significant in Teotihuacán (L-R chi-square₄ = 17.29, AICc = 342.00; p = 0.002) (Table 1). Nevertheless, only the negative effect of *L. daturaphila* larvae on fitness was significant (Table 1). Likewise, a significant positive effect of *Epitrix* sp. infestation level on fitness was detected in Ticumán (*L-R* chi-square₁ = 13.06, AICc = 130.18, p = 0.001; Table 1).

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Discussion

539 Our results revealed differentiation in plant-herbivore interactions among the study sites. First, 540 different herbivore species are present in each population, and the infestation levels and the 541 amount of foliar damage exerted by each herbivore on plants differed within and between 542 populations. Second, different chemical compounds were related to infestation by each specific 543 herbivore. Third, variable spatial selection was detected on identity by descent (IBD), resistance, 544 chemical defensive traits and herbivore infestation levels. 545 A number of studies have also documented geographic variation in the level of herbivory 546 and chemical defenses (Castells et al. 2005; Muola et al. 2010; Agrawal et al. 2012; Züst et al.

547 2012; Castillo *et al.* 2014; Verçosa *et al.* 2019; Hanh *et al.* 2019). However, there has been no

548 previous attempts to determine how the plants' genetic background (IBD) is driven by ongoing

549 natural selection-imposed by herbivores. Furthermore, the results provide strong evidence of

550 local adaptation in plant-herbivore interactions in both populations of *D. stramonium*.

551 In the locality of Teotihuacán, F₂ plants more related to the local grandparent (selected as 552 a parent due to its low alkaloid concentration) were less resistant and had higher mortality due to 553 herbivory than F₂ plants more related to the Ticumán grandparent. Furthermore, we did not 554 detect any relationship between fitness and IBD to the Teotihuacán grandparent in this site. This 555 result was anticipated since it is not expected that natural selection would favor poorly defended 556 plants in a habitat where damage by herbivores can be lethal (e.g., plant deaths due to herbivory 557 caused by larvae of Lema daturaphila) (Valverde et al. 2001; 2003; Fornoni et al. 2004). Hence, 558 it is plausible that F₂ plants more related to the Teotihuacán grandparent inherited the loci/alleles 559 that do not confer resistance (Albrechtsen et al. 2010; Browning and Browning 2012). Likewise, 560 alkaloid concentration in F₂ plants more related to the Teotihuacán grandparent remained at low 561 levels after damage by different herbivores in Teotihuacán. This result indicates that the 562 chemical defenses studied here are not induced and have a genetic basis, since a positive 563 significant relationship between plant resistance and IBD to the Teotihuacán grandparent would 564 be expected if plant defenses were induced after herbivore damage (Baldwin 1998; Karban and 565 Baldwin 2007).

566 In marked contrast, we detected strong positive selection on IBD to the Ticumán 567 grandparent in the locality of Teotihuacán. Also, plant resistance to herbivores and IBD to the 568 Ticumán grandparent were positively related in this site. We suggest that F₂ plants more related to the Ticumán grandparent - selected as a parent on the basis of its high alkaloid concentration -569 570 had higher survival in Teotihuacán as they inherited the loci/alleles that confer resistance to 571 herbivores; positive selection of these loci/alleles would be associated with different defensive 572 chemical compounds which are produced in high concentration (Albrechtsen et al. 2010; Lowry 573 et al. 2019). In fact, our findings indicate that the higher resistance of F₂ plants more related to

574 the Ticumán grandparent in the Teotihuacán site was provided by specific alkaloids that are 575 produced in very high concentration to face different herbivore species. Total alkaloid 576 concentration (sum of the concentration of all classes of alkaloids; Moore et al. 2014) only seems 577 to affect negatively the infestation levels of *E. parvula*. Since alkaloid concentrations vary in 578 wild Teotihuacán plants (Castillo et al. 2014; Miranda-Pérez et al. 2016; De-la-Cruz et al. 2020), 579 we think that wild plants from Teotihuacán that produce specific alkaloids in very high 580 concentrations (i. e., plants more related to the Ticumán grandparent) have strong chemical 581 defense against the herbivores in this site. For instance, we observed strong positive selection to 582 increase the concentration of the triterpenoid compound in Teotihuacán, which seems to affect 583 negatively the infestation levels of the most harmful herbivore of D. stramonium, the larvae of L. 584 daturaphila.

585 The defensive role of specific alkaloids in the Teotihuacán site revealed unexpected 586 results, namely, changing the sign of their relationship with the infestation by different 587 herbivores. For instance, while the triterpenoid compound appears to reduce the infestation of 588 Lema larvae (the most dangerous herbivore of D. stramonium), it was also positively associated 589 with infestation levels by *E. parvula* and *T. soror*. Triterpenoids are structurally similar to insect 590 hormones known as ecdysones (Oliveira et al. 2019) known to control metamorphosis as insects 591 pass from larva to pupa to adult (Yamanaka et al. 2013). It has been reported that many 592 triterpenoids function as ecdysone blockers (e. g., azadirone; Ujváry 2010; Oliveira et al. 2019). 593 Therefore, the most parsimonious explanation for our observations is that this triterpenoid of D. 594 stramonium is acting mainly on larvae of Lema (Miller et al. 1989; Ujváry 2010), and since this 595 compound is structurally similar to insect hormones (Ujváry 2010), it may be used by E. parvula 596 and T. soror adults to trace D. stramonium plants (and potential mates on them). Complex

597 interactions where one compound is toxic to insects at one developmental stage (*e. g.*, larvae) or 598 to a particular herbivore species, but functioning as an attractant at other stage (adults) or to other 599 herbivore species have been reported, for instance, in *Nicotiana attenuata* (Zhou *et al.* 2017).

Local adaptation of plant defenses to herbivores depends on (1) the strength of selection as a result of the interaction, and (2) the level of specificity on the interaction (*e. g.*, folivores, seed predators, stem-borers) (Thompson *et al.* 2005; Cogni and Futuyma 2009; Agrawal *et al.* 2012). In the Teotihuacán site, our results suggest that the strong selection pressure exerted by one herbivore (the folivore *L. daturaphila*) on *D. stramonium* plants may affect the interaction between plants and other insects, leading to local adaptation of plant defenses to different herbivore species (Wise 2009, 2010).

607 On the other hand, in the Ticumán site, we detected strong positive selection on pyrroline 608 alkaloid. It has been reported that pyrroline is a defensive compound against many insect species 609 and pathogens (bacteria, virus, fungi) (Qamar et al. 2015; Martins et al. 2015; Tamariz et al. 610 2018). Pyrroline has also been related to different physiological processes such as plant growth 611 (Chen et al. 2018; Tamariz et al. 2018). It is worth mentioning that polyamine oxidase, an 612 enzyme involved in the biosynthesis of the pyrroline, is a growth-regulating enzyme (Chen et al. 613 2018). Nevertheless, an unexpected finding is that we observed a positive association between 614 pyrroline concentration and infestation level by *Epitrix* in Ticumán. It has reported that some 615 herbivore insects can tolerate pyrrolizidine alkaloids and use them for defense against their 616 predators or as precursors of insect hormones (Martins et al. 2015). Thus, our most parsimonious 617 explanation is that *Epitrix* sp. is surpassing the defensive role of the pyrroline alkaloid in 618 Ticumán. This could explain why the F_2 plants more genetically related to the Ticumán 619 grandparent (with higher concentration of pyrroline) had lower resistance towards Epitrix sp.

infestation. Furthermore, since pyrroline could be positively related to plant growth (Chen *et al.*2018; Tamariz *et al.* 2018), it is also possible that *Epitrix* sp. searches for more vigorous plants,
which have more biomass to feed (Agrawal 2005; Wise and Rausher 2013). On the other hand,
we observed that 3-hydroxy-6-tigloyloxytropane negatively affected the infestation levels of *Epitrix* sp. in Ticumán. Then, it seems that the latter alkaloid is providing resistance against this
herbivore in this site.

626 Interestingly, pyrroline affected negatively the infestation levels of *E. parvula* in 627 Teotihuacán. However, negative selection on this compound was detected also in Teotihuacán. 628 Thus, while Epitrix sp. appears to be adapted to this compound in Ticumán, the production of 629 this compound in high concentrations in Teotihuacán may involve physiological costs, as plants 630 also have to allocate resources for production of other compounds (e. g., triterpenoid) to tackle 631 their most harmful herbivore (Lema larvae). Indeed, as we mentioned above, it seems that total 632 alkaloid concentration should be the option to face with E. parvula infestation in Teotihuacán. 633 De-la-Cruz et al. (2020) found that plants from Ticumán have on average higher alkaloid 634 concentration than those in Teotihuacán. Why we did not observe strong selection to increase the 635 IBD to the Ticumán grandparent (higher alkaloid concentration) in Ticumán? Our most 636 parsimonious explanations are, first, as mentioned above, that *Epitrix* sp. (the main herbivore in 637 this site) seems locally adapted to plant chemical defenses (pyrroline) in Ticumán, and that other 638 alkaloids could now be providing defense against this herbivore. Second, since these compounds 639 are expressed constitutively, it is possible that all these powerful chemical weapons are being 640 used to face other natural enemies (virus, bacteria, nematodes, fungi, oomycete, other herbivore 641 species) that we did not detect or that were not present during our experiment. Third, it is also 642 possible that these compounds have other functions in this habitat (e. g., growth, plant-plant

643 communication). For instance, recent genomic evidence from *D. stramonium* indicates that
644 tropane alkaloids such as atropine and scopolamine also act as defenses against pathogens and
645 viruses (De-la-Cruz *et al.* under review).

Finally, the lack of association between the IBD to the Teotihuacán grandparent with
resistance or fitness in Ticumán suggests that the chemical defenses studied here are not induced
(see above; Karban and Baldwin 2007).

649

650

Conclusions

651 The methodology used in this study allowed us to get insights on how natural selection imposed 652 by herbivores drives the genetic underpinnings of plant resistance traits. The lack of association 653 between plant fitness and IBD to the Teotihuacán grandparent (low resistance) in both 654 populations, as well as different magnitude and direction of selection on the IBD to the Ticumán 655 grandparent (high resistance) across populations, provides evidence of how ongoing natural 656 selection operates on plant resistance and promotes local adaptation. Likewise, the results of this 657 study shed some new light on how plants defend themselves against the attack from different 658 herbivores. It seems that in populations where plants are suffering frequent or heavy damage by 659 different herbivores, plants are able to produce and "use" different chemical defensive 660 compounds to face each insect species that feed on them (Wittstock and Gershenzon 2002). The same alkaloids were produced by plants in both populations, but plants possessing the highest 661 662 total alkaloid concentrations were not the most well-defended or fit in either of the populations. 663 Instead, different specific alkaloids appear to be favored by natural selection imposed by 664 herbivores in the two study populations.

665	Our results also revealed that the strong negative selection imposed by one herbivore
666	species (e. g., larvae of L. daturaphila) on plants likely affects interactions with other insects.
667	Most importantly, the results provide evidence for local adaptation by showing that selection
668	favors different loci/alleles related to plant resistance to herbivores in the two populations
669	(Briscoe Runquist et al. 2020). Hence, intraspecific diversity in secondary metabolites of D.
670	stramonium seems to be maintained and selected to cope with varying local conditions among
671	populations (Moore et al. 2014), giving rise to a geographic coevolutionary mosaic (Thompson
672	2005).
673	
674	References
675	Agrawal, A. A. (2005). Natural selection on common milkweed (Asclepias syriaca) by a community of specialized
676	insect herbivores. Evolutionary Ecology Research, 7(5), 651-667.
677	Agrawal, A. A., Hastings, A. P., Johnson, M. T. J., Maron, J. L., and Salminen, JP. (2012). Insect herbivores drive
678	real-time ecological and evolutionary change in plant populations. Science, 338(6103), 113-116.
679	Albrechtsen, A., Moltke, I., and Nielsen, R. (2010). Natural selection and the distribution of Identity by Descent in
680	the human genome. Genetics, 186(1), 295-308.
681	Albrechtsen, A., Sand Korneliussen, T., Moltke, I., van Overseem Hansen, T., Nielsen, F. C., and Nielsen, R.
682	(2009). Relatedness mapping and tracts of relatedness for genome-wide data in the presence of linkage
683	disequilibrium. Genetic Epidemiology, 33(3), 266-274.
684	Álvarez-Caballero, J. M., and Coy-Barrera, E. (2019). Chemical and antifungal variability of several accessions of
685	Azadirachta indica A. Juss. from six locations across the Colombian caribbean coast: identification of
686	antifungal azadirone Limonoids. Plants, 8(12), 555.
687	Anderson, J. T., Wagner, M. R., Rushworth, C. A., Prasad, K. V. S. K., and Mitchell-Olds, T. (2014). The evolution
688	of quantitative traits in complex environments. Heredity, 112(1), 4-12.
689	Andrews, S., 2010 FastQC: a quality control tool for high throughput sequence data.
690	http://www.bioinformatics.babraham.ac.uk/projects/fastqc

- Akaike, H. (1974). A new look at the statistical model identification. IEEE Transactions on Automatic Control
 19(6), 716-723.
- Baldwin, I. T. (1998). Jasmonate-induced responses are costly but benefit plants under attack in native populations.
 Proceedings of the National Academy of Sciences, 95(14), 8113-8118.
- Bello-Bedoy, R., and Núñez-Farfán, J. (2010). Cost of inbreeding in resistance to herbivores in *Datura stramonium*.
 Annals of Botany, 105(5), 747-753.
- Berenbaum, M. R., Zangerl, A. R., and Nitao, J. K. (1986). Constraints on chemical coevolution: wild parsnips and
 the parsnip webworm. Evolution, 40(6), 1215-1228.
- Briscoe Runquist, R. D., Gorton, A. J., Yoder, J. B., Deacon, N. J., Grossman, J. J., Kothari, S., Lyons, M. P., Sheth,
- 700S. N., Tiffin, P., and Moeller, D. A. (2020). Context dependence of local adaptation to abiotic and biotic
- 701 environments: a quantitative and qualitative synthesis. The American Naturalist, 195(3), 412-431.
- Bolger, A. M., Lohse, M., and Usadel, B. (2014) Trimmomatic: a flexible trimmer for Illumina sequence data.
 Bioinformatics. 30:2114-20.
- Browning, S. R., and Browning, B. L. (2012). Identity by Descent between distant relatives: detection and
 applications. Annual Review of Genetics, 46(1), 617-633.
- Cantelo, W. W., and Jacobson, M. (1979). Phenylacetaldehyde attracts moths to bladder flower and to blacklight
 traps. Environmental Entomology, 8(3), 444-447.
- 708 Castillo, G., Cruz, L. L., Tapia-López, R., Olmedo-Vicente, E., Carmona, D., Anaya-Lang, A. L., Fornoni, J.,
- 709 Andraca-Gómez, G., Valverde, P. L., and Núñez-Farfán, J. (2014). Selection mosaic exerted by specialist and
- 710 generalist herbivores on chemical and physical defense of *Datura stramonium*. PLoS ONE, 9(7), e102478.
- 711 Castells, E., Berhow, M. A., Vaughn, S. F., and Berenbaum, M. R. (2005). Geographic variation in alkaloid
- 712 production in *Conium maculatum* populations experiencing differential herbivory by *Agonopterix*
- 713 *alstroemeriana*. Journal of Chemical Ecology, 31(8), 1693-1709.
- Charlesworth, B. (1998). Measures of divergence between populations and the effect of forces that reduce
 variability. Molecular Biology and Evolution, 15(5), 538-543.
- 716 Chen, D., Shao, Q., Yin, L., Younis, A., & Zheng, B. (2019). Polyamine Function in Plants: Metabolism, Regulation
- 717 on Development, and Roles in Abiotic Stress Responses. *Frontiers in Plant Science*, 9.

- Cogni, R., and Futuyma, D. J. (2009). Local adaptation in a plant herbivore interaction depends on the spatial scale.
 Biological Journal of the Linnean Society, 97(3), 494-502.
- Daniels, R. F., Schaffner, S. F., Wenger, E. A., Proctor, J. L., Chang, H.-H., Wong, W., Baro, N., Ndiaye, D., Fall,
- 721 F. B., Ndiop, M., Ba, M., Milner, D. A., Taylor, T. E., Neafsey, D. E., Volkman, S. K., Eckhoff, P. A., Hartl,
- 722 D. L., and Wirth, D. F. (2015). Modeling malaria genomics reveals transmission decline and rebound in
- 723 Senegal. Proceedings of the National Academy of Sciences, 112(22), 7067–7072.
- Dawkins, R., Krebs, J. R., Maynard Smith, J., and Holliday, R. (1979). Arms races between and within species.
- 725 Proceedings of the Royal Society of London. Series B. Biological Sciences, 205(1161), 489-511.
- 726 De-la-Cruz, I. M., Cruz, L. L., Martínez-García, L., Valverde, P. L., Flores-Ortiz, C. M., Hernández-Portilla, L. B.,
- and Núñez-Farfán, J. (2020). Evolutionary response to herbivory: population differentiation in microsatellite
- 128 loci, tropane alkaloids and leaf trichome density in *Datura stramonium*. Arthropod-Plant Interactions, 14(1),
- 729 21-30.
- De-la-Cruz, I. M., Merilä, J., Valverde, P. L., Flores-Ortiz, C. M., and Núñez-Farfán, J. (2020). Genomic and
 chemical evidence for local adaptation in resistance to different herbivores in *Datura stramonium*, Dryad,
- 732 Dataset, https://doi.org/10.5061/dryad.bnzs7h489.
- de Villemereuil, P., Gaggiotti, O. E., Mouterde, M., and Till-Bottraud, I. (2016). Common garden experiments in the
 genomic era: New perspectives and opportunities. Heredity, 116(3), 249-254.
- Doyle, J. J., and Doyle, J. L. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue.
 Phytochemical Bulletin, 19, 11-15.
- 737 Ehrlich, P. R., and Raven, P. H. (1964). Butterflies and plants: a study in coevolution. Evolution, 18(4), 586-608.
- Falconer, D. S., and Mackay, T. (1996). Introduction to quantitative genetics. Harlow: Longmans Green. 386 pp.
- Flood, P. J., and Hancock, A. M. (2017). The genomic basis of adaptation in plants. Current Opinion in Plant
 Biology, 36, 88-94.
- Fornoni, J., and Núñez-Farfán, J. (2000). Evolutionary ecology of *Datura stramonium*: genetic variation and costs
 for tolerance to defoliation. Evolution, 54(3), 789-797.
- Fornoni, Juan, Valverde, P. L., and Núnez-Fárfan, J. (2004). Population variation in the cost and benefit of tolerance
- and resistance against herbivory in *Datura stramonium*. Evolution, 58(8), 1696-1704.

- 745 Gomulkiewicz, R., Thompson, J. N., Holt, R. D., Nuismer, S. L., and Hochberg, M. E. (2000). Hot Spots, Cold
- 746 Spots, and the Geographic Mosaic Theory of Coevolution. The American Naturalist, 156(2), 156–174.
- 747 González-Coloma, López-Balboa, A., Santana, C., Reina, O., and Fraga, B. M. (2011). Triterpene-based plant
- 748 defenses. Phytochemistry Reviews, 10(2), 245-260.
- Hahn, P. G., Agrawal, A. A., Sussman, K. I., and Maron, J. L. (2019). Population variation, environmental gradients,
- and the evolutionary ecology of plant defense against herbivory. The American Naturalist, 193(1), 20-34.
- Henden, L., Lee, S., Mueller, I., Barry, A., and Bahlo, M. (2018). Identity-by-descent analyses for measuring
 population dynamics and selection in recombining pathogens. PLoS Genetics, 14(5), e1007279.
- Ionkova, I., Witte L., and Alfermann H.-A. (1994). Spectrum of tropane alkaloids in transformed roots of *Datura innoxia* and *Hyoscyamus x gyorffyi* cultivated in vitro. Planta Medica, 60, 382-384.
- Janz, N. (2011). Ehrlich and Raven revisited: Mechanisms underlying codiversification of plants and enemies.
- Annual Review of Ecology, Evolution, and Systematics, 42(1), 71-89.
- 757 Karban, R., and Baldwin, I. T. (2007). Induced responses to herbivory. University of Chicago Press.
- Kariñho-Betancourt, E., Agrawal, A. A., Halitschke, R., and Núñez-Farfán, J. (2015). Phylogenetic correlations
 among chemical and physical plant defenses change with ontogeny. New Phytologist, 206(2), 796-806.
- 760 Kawecki, T. J., and Ebert, D. (2004). Conceptual issues in local adaptation. Ecology Letters, 7(12), 1225-1241.
- Lande, R. (1979). Quantitative genetic analysis of multivariate evolution applied to brain: body size allometry.
 Evolution, 33(1Part2), 402-416.
- Lande, R., and Arnold, S. J. (1983). The measurement of selection on correlated characters. Evolution, 37(6), 12101226.
- Li, H., and Durbin, R. (2009) Fast and accurate short read alignment with Burrows-Wheeler Transform.
- 766 Bioinformatics, 25:1754-60.
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., Durbin, R., and 1000
- Genome Project Data Processing Subgroup. The sequence alignment/map (SAM) format and SAMtools,
 Bioinformatics (2009) 25(16) 2078-9.
- TO Lowry, D. B., Lovell, J. T., Zhang, L., Bonnette, J., Fay, P. A., Mitchell, R. B., Lloyd-Reilley, J., Boe, A. R., Wu,
- 771 Y., Rouquette, F. M., Wynia, R. L., Weng, X., Behrman, K. D., Healey, A., Barry, K., Lipzen, A., Bauer, D.,
- 572 Sharma, A., Jenkins, J., ... Juenger, T. E. (2019). QTL × environment interactions underlie adaptive

- divergence in switchgrass across a large latitudinal gradient. Proceedings of the National Academy of Sciences,
 116(26), 12933-12941.
- 775 Martins, C. H. Z., Cunha, B. P., Solferini, V. N., and Trigo, J. R. (2015). Feeding on host plants with different
- concentrations and structures of pyrrolizidine alkaloids impacts the chemical-defense effectiveness of a
 specialist herbivore. PLoS ONE, 10(10): e0141480.
- 778 Machado, B. B., Orue, J. P. M., Arruda, M. S., Santos, C. V., Sarath, D. S., Goncalves, W. N., Silva, G. G., Pistori,
- H., Roel, A. R., and Rodrigues-Jr, J. F. (2016). BioLeaf: a professional mobile application to measure foliar
 damage caused by insect herbivory. Computers and Electronics in Agriculture, 129, 44-55.
- Mauricio, R., and Rausher, M. D. (1997). Experimental manipulation of putative selective agents provides evidence
 for the role of natural enemies in the evolution of plant defense. Evolution, 51(5), 1435-1444.
- Miller, A. J., and Chamberlain, W. F. (1989). Azadirachtin as a larvicide against the horn fly, stable fly, and house
 fly (Diptera: Muscidae). Journal of Economic Entomology, 82(5), 1375-1378.
- 785 Miranda-Pérez, A., Castillo, G., Hernández-Cumplido, J., Valverde, P. L., Borbolla, M., Cruz, L. L., Tapia-López,
- R., Fornoni, J., Flores-Ortiz, C. M., and Núñez-Farfán, J. (2016). Natural selection drives chemical resistance
 of *Datura stramonium*. PeerJ, 4.
- 788 Moore, B. D., Andrew, R. L., Külheim, C., and Foley, W. J. (2014). Explaining intraspecific diversity in plant
- secondary metabolites in an ecological context. New Phytologist, 201(3), 733-750.
- Motten, A. F., and Antonovics, J. (1992). Determinants of outcrossing rate in a predominantly self-fertilizing weed,
 Datura stramonium (Solanaceae). American Journal of Botany, 79(4), 419-427.
- Muola, A., Mutikainen, P., Lilley, M., Laukkanen, L., Salminen, J.-P., and Leimu, R. (2010). Associations of plant
 fitness, leaf chemistry, and damage suggest selection mosaic in plant–herbivore interactions. Ecology, 91(9),
- *2650-2659.*
- Núnez-Farfán, J., Cabrales-Vargas, R. A., and Dirzo, R. (1996). Mating system consequences on resistance to herb
 ivory and life history traits in *Datura stramonium*. American Journal of Botany, 83(8), 1041-1049.
- Núñez-Farfán, J., and Dirzo, R. (1994). Evolutionary ecology of *Datura Stramonium* L. in central Mexico: natural
 selection for resistance to herbivorous insects. Evolution, 48(2), 423-436.
- 799 Núñez-Farfán, J., Fornoni, J., and Valverde, P. L. (2007). The Evolution of resistance and tolerance to herbivores.
- Annual Review of Ecology, Evolution, and Systematics, 38(1), 541-566.

- 801 Rastas, P. (2017). Lep-MAP3: Robust linkage mapping even for low-coverage whole genome sequencing data, 802 Bioinformatics, 33(23), 3726-3732.
- 803 Oliveira, D. A. B. de, and Niculau, A. V. da S. and E. dos S. (2019). Molecular docking of Azadirachtin in nuclear 804 Ecdysone receptor. Current Physical Chemistry, 6(1), 50-57.
- 805 Pedrol, N., and Tiburcio, A. F. (2001). Polyamines determination by TLC and HPLC. In M. J. Reigosa Roger (Ed.), 806
- Handbook of Plant Ecophysiology Techniques (pp. 335-363). Springer Netherlands.
- 807 Qamar, A., Mysore, K. S., & Senthil-Kumar, M. (2015). Role of proline and pyrroline-5-carboxylate metabolism in 808 plant defense against invading pathogens. Frontiers in Plant Science, 6.
- 809 Savolainen, O., Lascoux, M., and Merilä, J. (2013). Ecological genomics of local adaptation. Nature Reviews 810 Genetics, 14(11), 807-820.
- 811 Shonle, I., and Bergelson, J. (2000). Evolutionary Ecology of the tropane alkaloids of Datura stramonium L.
- 812 (Solanaceae). Evolution, 54(3), 778-788.
- 813 Sokal, R. R., and Rohlf, F. J. (1994). Biometry: The principles and practice of statistics in biological research. W H 814 Freeman & Co.
- 815 Stam, J. M., Kroes, A., Li, Y., Gols, R., van Loon, J. J. A., Poelman, E. H., and Dicke, M. (2014). Plant interactions 816 with multiple insect herbivores: from community to genes. Annual Review of Plant Biology, 65(1), 689-713.
- 817 Tamariz, J., Burgueño-Tapia, E., Vázquez, M. A., and Delgado, F. (2018). Pyrrolizidine alkaloids. In H.-J. Knölker 818 (Ed.), The alkaloids: chemistry and biology (Vol. 80, pp. 1-314). Academic Press.
- 819 Thompson, J. N., and Cunningham, B. M. (2002). Geographic structure and dynamics of coevolutionary selection. 820 Nature, 417(6890), 735-738.
- 821 Thompson, E. A. (2013). Identity by descent: variation in meiosis, across genomes, and in populations. Genetics, 822 194(2), 301-326.
- 823 Thompson, J. N. (2005). Coevolution: the geographic mosaic of coevolutionary arms races. Current Biology, 15(24), 824 R992-R994.
- 825 Ujváry, I. (2010). Chapter 3-pest control agents from natural products. In R. Krieger (Ed.), Hayes' Handbook of 826 Pesticide Toxicology (Third Edition) (pp. 119-229). Academic Press.
- 827 Valverde, P. L., Fornoni, J., and Núñez-Farfán, J. (2001). Defensive role of leaf trichomes in resistance to
- 828 herbivorous insects in Datura stramonium. Journal of Evolutionary Biology, 14(3), 424-432.

- Valverde, P. L., Fornoni, J., and Núñez-Farfán, J. (2003). Evolutionary ecology of *Datura stramonium*: Equal plant
 fitness benefits of growth and resistance against herbivory. Journal of Evolutionary Biology, 16(1), 127-137.
- 831 Verçosa, D., Cogni, R., Alves, M. N., and Trigo, J. R. (2019). The geographical and seasonal mosaic in a plant-
- 832 herbivore interaction: Patterns of defences and herbivory by a specialist and a non-specialist. Scientific
- Reports, 9(1), 15206.
- 834 Wickham, H. (2016). ggplot2: Elegant graphics for data analysis (2nd ed.). Springer International Publishing.
- Wise, Michael Joseph. (2009). Competition among herbivores of *Solanum carolinense* as a constraint on the
 evolution of host-plant resistance. Evolutionary Ecology, 23(3), 347–361.
- Wise, Michael J. (2010). Diffuse interactions between two herbivores and constraints on the evolution of resistance
 in horsenettle (*Solanum carolinense*). Arthropod-Plant Interactions, 4(3), 159-164.
- 839 Wise, M. J., and Rausher, M. D. (2013). Evolution of resistance to a multiple-herbivore community: genetic
- correlations, diffuse coevolution, and constraints on the plant's response to selection. Evolution, 67(6), 17671779.
- Wittstock, U., and Gershenzon, J. (2002). Constitutive plant toxins and their role in defense against herbivores and
 pathogens. Current Opinion in Plant Biology, 5(4), 300-307.
- Witte, L., Müller, K., and Arfmann, H. A. (1987). Investigation of the alkaloid pattern of *Datura innoxia* plants by
 capillary gas-liquid-chromatography-mass spectrometry. Planta Medica, 53:192-197.
- 846 Wong, W., Griggs, A. D., Daniels, R. F., Schaffner, S. F., Ndiaye, D., Bei, A. K., Deme, A. B., MacInnis, B.,
- 847 Volkman, S. K., Hartl, D. L., Neafsey, D. E., and Wirth, D. F. (2017). Genetic relatedness analysis reveals the
- 848 cotransmission of genetically related *Plasmodium falciparum* parasites in Thiès, Senegal. Genome Medicine,
- **849** 9(1), 5.
- Yamanaka, N., Rewitz, K. F., and O'Connor, M. B. (2013). Ecdysone control of developmental transitions: lessons
 from drosophila research. Annual Review of Entomology, 58(1), 497-516.
- 852 Zar, J. H. (1999). Biostatistical Analysis. Prentice Hall.
- Zhou, W., Kügler, A., McGale, E., Haverkamp, A., Knaden, M., Guo, H., Beran, F., Yon, F., Li, R., Lackus, N.,
- Köllner, T. G., Bing, J., Schuman, M. C., Hansson, B. S., Kessler, D., Baldwin, I. T., and Xu, S. (2017).
- 855 Tissue-specific emission of (E)-α-Bergamotene helps resolve the dilemma when pollinators are also
- 856 herbivores. Current Biology, 27(9), 1336-1341.

- 857 Züst, T., Heichinger, C., Grossniklaus, U., Harrington, R., Kliebenstein, D. J., and Turnbull, L. A. (2012). Natural
- 858 enemies drive geographic variation in plant defenses. Science, 338(6103), 116-119.

Table 1. Analyses of natural selection testing the effects of (a) Identity by Descent to Ticumán grandparent, (b) Identity by Descent to Teotihuacán grandparent, (c) resistance, (d) the seven alkaloids, (e) total alkaloid concentration, (f) herbivores at Teotihuacán and (g) *Epitrix* sp. infestation at Ticumán. N = number of individuals, d.f. = degrees of freedom, β_i = selection gradients (generalized linear coefficients), se = standard error, *t* = t-ratio. Significant *p*-values (*p*) are in bold. Pop = effect of the experimental site.

Response variable	Effects	Ν	d.f.	βi	se	t	р
(a) w_i fitness	IBD-Ticumán grandparent	135	3	2.19	0.99	2.20	0.0302
	Рор	135	3	-0.13	0.10	-1.22	0.2262
	$Pop \times IBD$	135	3	1.39	0.99	1.40	0.1646
(b) w_i fitness	IBD-Teotihuacán grandparent	135	3	0.41	0.89	0.46	0.6484
	Рор	135	3	-0.17	0.11	-1.49	0.1390
	$Pop \times IBD$	135	3	-0.71	0.89	-0.80	0.4252
(c) w_i fitness	Resistance	177	1	0.11	0.27	0.43	0.6624
	Рор	177	1	-0.15	0.31	-0.50	0.6143
	Pop × Resistance	177	1	0.64	0.27	2.36	0.0179
(d) w_i fitness	3-hydroxy-6-tigloyloxytropane	136	1	-0.36	0.21	-1.71	0.0899
	Anisodamine	136	1	-0.07	0.28	-0.25	0.7961
	Atropine	136	1	-0.07	0.12	-0.56	0.5719
	Triterpenoid	136	1	0.26	0.18	1.46	0.1459
	Scopolamine	136	1	-0.24	0.19	-1.24	0.2151
	Phenylacetaldehyde	136	1	-0.21	0.21	-0.98	0.3254
	Pyrroline	136	1	0.32	0.14	2.19	0.0304
	Рор	136	1	-0.12	0.09	-1.29	0.1997
	Pop × 3-hydroxy-6-tigloyloxytropane	136	1	0.13	0.21	0.62	0.5362
	Pop × Anisodamine	136	1	-0.06	0.28	-0.24	0.8104
	$Pop \times Atropine$	136	1	0.19	0.12	1.55	0.1224
	$Pop \times Triterpenoid$	136	1	0.38	0.18	2.10	0.0378
	Pop × Scopolamine	136	1	-0.04	0.19	-0.20	0.8375
	Pop × Phenylacetaldehyde	136	1	-0.14	0.21	-0.67	0.5015
	Pop × Pyrroline	136	1	-0.43	0.14	-2.93	0.0040
(e) w_i fitness	Total alkaloid concentration	144	1	-0.05	0.08	-0.58	0.5657
	Pop	144	1	-0.12	0.09	-1.34	0.1838
	Pop × Total alkaloid concentration	144	1	0.09	0.08	1.04	0.2988
(f) w_i fitness	Adults of Lema daturaphila	113	1	0.08	0.09	0.90	0.3692
	Adults of Epitrix parvula	113	1	0.09	0.09	0.97	0.3324
	Adults of Trichobaris soror	113	1	0.09	0.08	1.03	0.3070
	Larvae of Lema daturaphila	113	1	-0.27	0.08	-3.22	0.0017
(g) w_i fitness	Adults of Epitrix sp.	63	1	0.30	0.081	3.75	0.0004

Table 2. General linear models testing the effect of (a) Identity by Descent to the Ticumán grandparent, (b) Identity by Descent to the Teotihuacán grandparent, (c) herbivore infestation at Teotihuacán and (d) *Epitrix* sp. infestation at Ticumán, on whole plant resistance. N = number of individuals, d.f. = degrees of freedom, Estimate = generalized linear coefficients, se = standard error, t = t-ratio. Significant p-values (p) are in bold. Pop = effect of the experimental site.

Response	e variable	Effects	Ν	d.f.	Estimate	se	t ratio	р
(a)	Resistance	IBD-Ticumán grandparent	182	3	0.98	0.64	1.52	0.1256
		Рор	182	3	-0.40	0.06	-5.95	0.0001
		Pop × IBD	182	3	2.06	0.64	3.18	0.0015
(b)	Resistance	IBD-Teotihuacán grandparent	182	3	0.31	0.48	0.64	0.5182
		Рор	182	3	-0.40	0.07	-5.69	0.0001
		Pop × IBD	182	3	0.51	0.48	1.05	0.2896
(c)	Resistance	Adults of Lema daturaphila	185	1	0.02	0.05	0.38	0.7009
		Epitrix parvula	185	1	0.27	0.06	4.51	0.0001
		Trichobaris soror	185	1	0.03	0.05	0.53	0.5944
		Larvae of Lema daturaphila	185	1	-0.52	0.05	-9.00	0.0001
(d)	Resistance	<i>Epitrix</i> sp.	82	1	-0.40	0.10	-4.00	0.0001

Figure legends

Fig. 1. Relationships between plant fitness (log scale) and identity by descent (IBD) relationship to the Ticumán grandparent for (a) plants grown in the Teotihuacán population and (b) plants grown in the Ticumán population. Relationships between plant fitness and IBD relationship to the Teotihuacán grandparent for (c) plants grown in the Teotihuacán population and (d) plants grown in the Ticumán population. (e) Box plot of IBD to the Ticumán grandparent for plants that survived and died. (f) Box plot of IBD to the Teotihuacán grandparent for plants that survived and died. (g) Distribution of the relatedness between all F_2 fullsibs, and relatedness of F_2 plants to each grandparent. A relatedness of ~0.5 is the mean expected value between all F_2 fullsibs. A relatedness of ~0.5 is the maximum expected value between the F_2 progeny and each one of the grandparents (Falconer and Mackay 1996). *p*-values of full GLMs are shown in each plot (a-f). Each dot depicts observation for an individual. See also Table 1.

Fig. 2. Relationships between resistance to herbivory and identity by descent (IBD) relationship to the Ticumán grandparent for (a) plants grown in the Teotihuacán population and (b) plants grown in the Ticumán population. Relationships between resistance to herbivory and IBD relationship to the Teotihuacán grandparent for (c) plants grown in the Teotihuacán population and (d) plants grown in the Ticumán population. Relationship between resistance and (e) adults of *Epitrix parvula* in Teotihuacán, (f) larvae of *Lema daturaphila* in Teotihuacán, (g) adults of *Epitrix* sp. in Ticumán. *p*-values of GLMs are shown in each plot (a-g). Each dot depicts observation for an individual. Pop = population. See also Table 2.







Supplementary Information (Figures and Tables) of the manuscript "Genomic and chemical evidence for local adaptation in resistance to different herbivores in *Datura stramonium*"

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Evolution

S1. Depiction of the experimental design used to produce the F₂ generation progeny used in the study (see Methods for details).



Experiment design to select the parents and to produce the F₂ generation progeny

S2. Alkaloids identified in leaves of *Datura stramonium*. RT, the retention time of each alkaloid; m/z = mass/charge, MS = mass spectrometry reference.

Alkaloid	Formula	RT (min)	m/z	MS Ref.
3-Tigloyloxy-6-Hydroxytropane	$C_{13}H_{21}NO_3$	10.1	240.1594	Witte 1987
6-Hydroxyhyoscyamine (Anisodamine)	$C_{17}H_{23}NO_4$	13.2	306.1699	Ionkova <i>et al</i> . 1994
Atropine (Hyosciamine)	$C_{17}H_{23}NO_3$	8.7	290.1751	Witte 1987
Scopolamine	$C_{17}H_{21}NO_4$	15	304.1543	Witte 1987
Pyrroline	C_4H_7N	5.5	70.0655	Pedrol and Tiburcio 2001
Phenylacetaldehyde	C_8H_8O	9.4	121.0641	Cantelo and Jacobson 1979
Triterpenoid	$C_{28}H_{36}O_4$	15.6	437.2721	González-Coloma et al. 2011

S3. Plant damage (in percentage) experienced by each F_2 plant during four sampling dates at (A) Teotihuacán and (B) Ticumán. Plants experienced the most severe damage at 45 days after transplanting in the two localities, and damage levels were higher in Teotihuacán than in Ticumán. (C) Violin plot showing total infestation accounted by different herbivore (log scale). The circle inside each violin depicts the mean value. Overall, *Lema daturaphila* larvae was the most abundant insect herbivore. A quasirandom jittering was used to reduce datapoint overlap. TEO = Teotihuacán, TIC = Ticumán. *p*-values of ANOVAs are showed in each plot. For figures (A) and (B): black line = mean.



S4. Mean differentiation in the level of damage (expressed as percentage) between sampling sessions in (a) Teotihuacán and (b) Ticumán. (c) Mean differentiation in the level of herbivore infestation (Log transformed data). N = number of individuals, se = standard error, d.f. = degree of freedoms, SS = Sum of Squares, MS = Mean Square, F = Fisher-statistic, p = p-values of ANOVAs.

ANOVA	Ν	Mean	se	Source	d.f.	SS	MS	F	р
(a) Damage at Teotihuacán									
15 days	164	19.55	0.07	Model	3	309.61	103.20	99.04	0.0001
30 days	179	48.84	0.07	Error	706	735.63	1.04		
45 days	184	90.08	0.07	Total	709	1045.24			
60 days	183	56.86	0.07						
(b) Damage at Ticumán									
15 days	83	6.40	0.12	Model	3	211.30	70.43	50.33	0.0001
30 days	83	9.96	0.12	Error	317	443.61	1.39		
45 days	73	14.51	0.13	Total	320	654.91			
60 days	82	12.15	0.13						
(c) Herbivore									
E. parvula (TEO)	185	1.74	0.06	Model	4	666.49	166.62	215.32	0.0001
<i>Epitrix</i> sp. (TIC)	47	1.87	0.12	Error	541	418.65	0.77		
Adults of L. daturaphila (TEO)	143	2.15	0.07	Total	545	1085.14			
Larvae of L. daturaphila (TEO)	131	4.36	0.07						
T. soror	40	1.16	0.13						

S5. Correlations between infestation by each herbivore and plant leaf average damage. a = Teotihuacán and b = Ticumán. ALd = adults of *Lema daturaphila*, Ep = *Epitrix parvula*, Ts = *Trichobaris soror*, LLd = larvae of *Lema daturaphila*.

Variable	by Variable	Correlation	Lower 95%	Upper 95%	<i>p</i> -value
		(a) 7	Feotihuacán		
Damage	LLd	0.615779	0.517482	0.698035	1.079E-20
Ts	Ep	0.241138	0.100361	0.37245	9.446E-04
Ep	ALd	0.168386	0.024718	0.305239	2.195E-02
Ts	ALd	0.137232	-0.00718	0.276036	6.250E-02
LLd	ALd	-0.03421	-0.1776	0.110605	6.439E-01
Damage	Ts	-0.123	-0.26261	0.021649	9.531E-02
Damage	ALd	-0.13037	-0.26956	0.014169	7.694E-02
LLd	Ts	-0.15396	-0.29175	-0.00991	3.640E-02
LLd	Ep	-0.31356	-0.43802	-0.17731	1.386E-05
Damage	Ep	-0.32261	-0.44611	-0.18704	7.516E-06
		(b)	Ticumán		
Damage	<i>Epitrix</i> sp.	0.387549	0.187536	0.556704	2.949E-04

S6. Generalized linear models (GLMs) between herbivore infestation levels and the seven alkaloids and total alkaloid concentration: (a) infestation by adults of *E. pavula*, (b) stepwise GLM between larvae of *Lema* and alkaloids. (c) stepwise GLM between adults of *Lema* and alkaloids, (d) stepwise GLM between adults of *T. soror* and alkaloids. (e) GLM between *Epitrix* sp. and alkaloids in Ticumán. N = number of individuals, d.f. = degree of freedoms, Estimate = generalized linear coefficients, se = standard error, t = t-ratio, Significant *p*-values are in bold. Notice that for the total alkaloid concentration was performed a different model for each herbivore (*i. e.*, the effect of this variable was not included in the GLMs between each herbivore and the seven alkaloids (see methods).

Teotihuacán								
Response variable		Effects	Ν	d.f.	Estimate	se	t	р
(a)	# of adult of Epitrix parvula	3-hydroxy-6-tigloyloxytropane	153	1	-0.29	0.17	3.09	0.0808
		Phenylacetaldehyde	153	1	-0.32	0.14	5.24	0.0234
		Pyrroline	153	1	-0.26	0.12	4.41	0.0375
		Triterpenoid	153	1	0.46	0.18	6.12	0.0145
		Scopolamine	153	1	-0.22	0.16	1.78	0.1831
		Anisodamine	153	1	0.27	0.21	1.65	0.1997
		Atropine	153	1	0.01	0.11	0.00	0.9272
		Total alkaloid concentration	161	1	-0.15	0.079	-1.98	0.0496
(b)	# of larvae of <i>Lema daturaphila</i>	Phenylacetaldehyde	156	1	0.31	0.13	2.38	0.0188
		Pyrroline	156	1	0.10	0.12	0.88	0.3820
		Triterpenoid	156	1	-0.56	0.17	-3.24	0.0015
		Scopolamine	156	1	0.08	0.16	0.48	0.6326
		Anisodamine	156	1	0.06	0.12	0.55	0.5837
		Total alkaloid concentration	161	1	-0.09	0.08	-1.20	0.2308
(c)	# adult of Lema daturaphila	3-hydroxy-6-tigloyloxytropane	156	1	-0.24	0.09	-2.65	0.0089
		Phenylacetaldehyde	156	1	-0.08	0.09	-0.84	0.4031
		Atropine	156	1	0.25	0.10	2.45	0.0152
		Total alkaloid concentration	161	1	0.05	0.08	0.70	0.4832
(d)	# adult of Trichobaris soror	Phenylacetaldehyde	158	1	-0.17	0.13	-1.29	0.1978
		Triterpenoid	158	1	0.33	0.16	2.05	0.0419
		Scopolamine	158	1	-0.37	0.14	-2.65	0.0088
		Atropine	158	1	0.13	0.11	1.16	0.1978
		Total alkaloid concentration	161	1	-0.02	0.08	-0.33	0.7387
Ticumán								
(e)	# of adult of <i>Epitrix</i> sp.	3-hydroxy-6-tigloyloxytropane	48	1	-0.43	0.20	4.56	0.0388
		Phenylacetaldehyde	48	1	0.044	0.17	0.06	0.7982
		Pyrroline	48	1	0.54	0.18	8.52	0.0057
		Triterpenoid	48	1	-0.19	0.20	0.90	0.3463
		Scopolamine	48	1	-0.21	0.18	1.32	0.2566
		Anisodamine	48	1	0.14	0.24	0.34	0.5596
		Atropine	48	1	-0.00	0.16	0.00	0.9807
		Total alkaloid concentration	50	1	0.11	0.15	0.83	0.4412

S7. Relationships between fitness and resistance in (A) F2 plants grown in Teotihuacán and (B) F2 plants grown in Ticumán. Positive selection to resistance was observed in Teotihuacán while a negative trend was detected in Ticumán. Each dot depicts observation for an individual. p-value of full model is shown in the plots. See Table 1.





S8. Prediction profilers from the GLM between fitness (w_i) and concentrations of the seven alkaloids. Response effects are shown separately for the two experimental sites. Envelops = confidence intervals (95% CI). See table 1.





References

- Cantelo, W. W., and Jacobson, M. (1979). Phenylacetaldehyde attracts moths to bladder flower and to blacklight traps. Environmental Entomology, 8(3), 444-447.
- González-Coloma, López-Balboa, A., Santana, C., Reina, O., and Fraga, B. M. (2011). Triterpene-based plant defenses. Phytochemistry Reviews, 10(2), 245-260.
- Ionkova, I., Witte L., and Alfermann H.-A. (1994). Spectrum of tropane alkaloids in transformed roots of *Datura innoxia* and *Hyoscyamus x gyorffyi* cultivated in vitro. Planta Medica, 60, 382-384.
- Pedrol, N., and Tiburcio, A. F. (2001). Polyamines determination by TLC and HPLC. In M. J. Reigosa Roger (Ed.), Handbook of Plant Ecophysiology Techniques (pp. 335-363). Springer Netherlands.
- Witte, L., Müller, K., and Arfmann, H. A. (1987). Investigation of the alkaloid pattern of *Datura innoxia* plants by capillary gas-liquid-chromatography-mass spectrometry. Planta Medica, 53:192-197.