



## ORIGINAL ARTICLE

# Assessment of enhanced influenza vaccination finds that FluAd conveys an advantage in mice and older adults

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## Abstract

**Objectives.** Enhanced inactivated influenza vaccines (eIIV) aim to increase immunogenicity and protection compared with the widely used standard IIV (S-IIV). **Methods.** We tested four vaccines in parallel, FluZone high dose, FluBlok and FluAd versus S-IIV in a randomised controlled trial of older adults and in a mouse infection model to assess immunogenicity, protection from lethal challenge and mechanisms of action. **Results.** In older adults, FluAd vaccination stimulated a superior antibody profile, including H3-HA antibodies that were elevated for up to 1 year after vaccination, higher avidity H3HA IgG and larger HA stem IgG responses. In a mouse model, FluAd also elicited an earlier and larger induction of HA stem antibodies with increased germinal centre responses and upregulation and long-term expression of B-cell switch transcription factors. Long-term cross-reactive memory responses were sustained by FluAd following lethal heterosubtypic influenza challenge, with reduced lung damage and viral loads, coinciding with increased T- and B-cell recall. Advantages were also noted for the high-dose FluZone vaccine in both humans and mice. **Conclusion.** The early, broadly reactive and long-lived antibody response of FluAd indicates a potential advantage of this vaccine, particularly in years when there is a mismatch between the vaccine strain and the circulating strain of influenza viruses.

**Keywords:** adjuvant, antibody, B cell, HA stem, infection, influenza vaccine

## INTRODUCTION

Vaccination against influenza viruses aims to mitigate morbidity and mortality associated with

infection. Adults over 65 years of age are the most susceptible group to complications of influenza infection, account for the majority of influenza-associated mortality and are widely

recommended for annual vaccination.<sup>1</sup> Influenza vaccine effectiveness varies from year to year and can be impacted by antigenic drift,<sup>2</sup> mismatch,<sup>3,4</sup> pandemic emergence,<sup>5</sup> egg adaptations<sup>6</sup> and age.<sup>7</sup> Standard-dose inactivated influenza vaccines (S-IIV) contain 15 µg of haemagglutinin (HA) per strain and are the most widely used vaccine formulation to reduce the burden of seasonal influenza epidemics.<sup>8</sup> Prioritising the use of newly available enhanced inactivated influenza vaccines (eIIVs) may increase the longevity, breadth and quality of vaccine-mediated protection against influenza virus infections and associated morbidity and mortality in older adults.<sup>9,10</sup>

Available eIIVs include vaccines that have a higher antigen content than S-IIV, such as recombinant FluBlok (recombinant HA protein, three times of HA content) or FluZone high dose (subunit vaccine, four times of HA content), or contain adjuvants, such as FluAd (MF59). The adjuvanted eIIV (A-eIIV), FluAd, contains MF59, an oil-in-water emulsion acting as both adjuvant and carrier for the antigens, previously increasing the breadth, diversity and avidity of HA antibodies.<sup>11</sup> A recombinant HA-only high-dose IIV (R-eIIV), FluBlok, has a reported higher vaccine efficacy than S-IIV.<sup>12</sup> However, focusing the FluBlok response to a single HA protein target, whilst a critical protein for viral entry, may diminish the breadth of vaccine-mediated protection, neglect NA-mediated protection and affect CD4<sup>+</sup> T-cell-mediated help directed at internal proteins.<sup>13</sup> The high-dose inactivated IIV (H-eIIV), FluZone, induces significantly higher antibody responses and provides better protection against laboratory-confirmed influenza than S-IIV among the elderly.<sup>14</sup> H-eIIV has also shown improved immunogenicity by HAI measures in HIV-immunocompromised patients, despite their low overall CD4<sup>+</sup> T-cell counts. This indicates that increasing the antigenic dose may result in a correspondingly increased immune response,<sup>15</sup> which is particularly important to older adults who experience immunosenescence.

Structural and post-translational modifications of the HA as a result of vaccine production methods, antigen dose and the inclusion of adjuvants may impact serological and cellular immunity.<sup>16,17</sup> Initial results of HAI and T-cell responses from our ongoing trial have shown R-eIIV elicits higher titre microneutralisation antibody titres against influenza A (H3N2) and CD4<sup>+</sup> T-cell boosting, followed by H-eIIV, then

A-eIIV.<sup>18</sup> Parallel comparison of eIIVs through experimental immunogenicity studies may help elucidate their mechanism of action, as large-scale efficacy studies are rare for already-licensed vaccines. In this study, we extended serological results from our ongoing randomised trial of eIIVs in older adults to a vaccination-challenge mouse model to assess eIIV-mediated protection.

## RESULTS

### A-eIIV increases the avidity, stem and long-term H3-HA-specific IgG responses in vaccinated human subjects

A randomised trial of community-dwelling older adults between 65 and 82 years old was initiated in 2017/2018 in Hong Kong to evaluate the immunogenicity of eIIVs (ClinicalTrials.gov NCT03330132).<sup>18</sup> Older adults (Supplementary table 1) were randomly allocated to receive the 2017/2018 Northern Hemisphere formulation of either standard FluQuadri (S-IIV, Sanofi Pasteur, Lyon, France) or one of three eIIVs, including adjuvanted (FluAd, A-eIIV, Seqirus, Maidenhead, UK), high dose (FluZone high dose, H-eIIV, Sanofi Pasteur) and recombinant HA (FluBlok, R-eIIV, Sanofi Pasteur) (Table 1).<sup>18</sup> Sera were collected for baseline responses (day 0) and then after vaccination at days 7 (acute), 30 (short-term memory) and 365 (long-term memory) (Figure 1a). Standard HAI serological responses and T-cell responses are reported elsewhere.<sup>18</sup>

We assessed anti-HA IgG titres (Figure 1b) and avidity (Figure 1c) against a representative vaccine strain H3N2-2013. Antibody responses were equivalent between all vaccine groups at days 7 and 30, but importantly, A-eIIV and R-eIIV H3N2-2013 HA IgG responses were elevated to long-term memory 1 year after vaccination compared with S-IIV. The A-eIIV vaccine also elicited a significant selective increase in high-avidity H3N2-2013 HA IgG at day 7, which was maintained 1 year after vaccination (ns, day 7 versus day 365; Figure 1c). A-eIIV also elicited an increase in HA stem-specific antibodies at day 7 from both H3-stem (Figure 1d,  $P = 0.0015$ ), which were unchanged at 1 year later (ns, day 7 versus day 365; Figure 1d), and H1-stem (Supplementary figure 1a). Furthermore, H-eIIV also mounted a short-lived H1-stem response at day 7 post-vaccination (Supplementary figure 1a). Therefore, A-eIIV induced the most superior quality IgG

**Table 1.** IIV composition and dosage for human and mouse vaccination

Abbreviation	Type	Vaccine name	Valence	Strains included				HA content per vial ( $\mu\text{g}$ )		HA content per mouse dose ( $\mu\text{g}$ )	
				H1	H3	B/Vic	B/Yam	Per strain	Total	Per strain	Total
S	Standard	FluQuadri	QIV	●	●	●	●	15	60	5	20
A	Adjuvant	FluAd	TIV	●	●	●	–	15	45	5	15
H	High dose	FluZone	TIV	●	●	●	–	60	180	20	60
R	Recombinant HA protein	FluBlok	QIV	●	●	●	●	45	180	15	60

responses in terms of long-term memory, high-avidity H3-IgG and H3-stem-specific responses.

Subsequent experiments focused on H3-HA-specific responses as a result of the challenge of antigenic matching for current vaccines and reduced vaccine efficacy.<sup>3,6</sup> Our initial study also found improved responses to enhanced vaccines for H3N2 viruses,<sup>18</sup> and we also observed similar trends for H1-HA-specific responses (Supplementary figures 1 and 5) and influenza B responses with limited reagents (data not shown).

### A-eIIV in mice induces superior influenza antibody responses for vaccine antigens, cross-reactive proteins and neutralisation

To understand the protective potential of high-avidity H3-HA and H3-stem IgG by eIIV in older adults, especially A-eIIV, we evaluated the same vaccines in mice (Figure 1e). Mice were vaccinated twice, and post-vaccination immune responses were assessed at days 7 and 21. The A-eIIV group had significantly increased day 7 and 21 post-vaccination magnitude of HA IgG responses to vaccine strains, H1N1-2009 and H3N2-2013 HA (Figure 1f), whilst other eIIVs were equivalent to S-IIV for these HA IgG responses. In addition, the H1N1-2009 NA IgG response was elevated for A-eIIV at days 7 and 21 post-vaccination, whilst H-eIIV was delayed to until day 21 post-vaccination compared with S-IIV (Figure 1f). As R-eIIV also does not contain NA, no substantial NA-specific IgG responses were detected. Furthermore, virus neutralisation antibodies (VNAs) at day 7 post-vaccination (Figure 1g) were significantly increased for A-eIIV against H1N1-2009 virus ( $907 \pm 174$  versus  $293 \pm 27$  S-IIV) and H-eIIV and A-eIIV against H3N2-2013 virus compared with S-IIV.

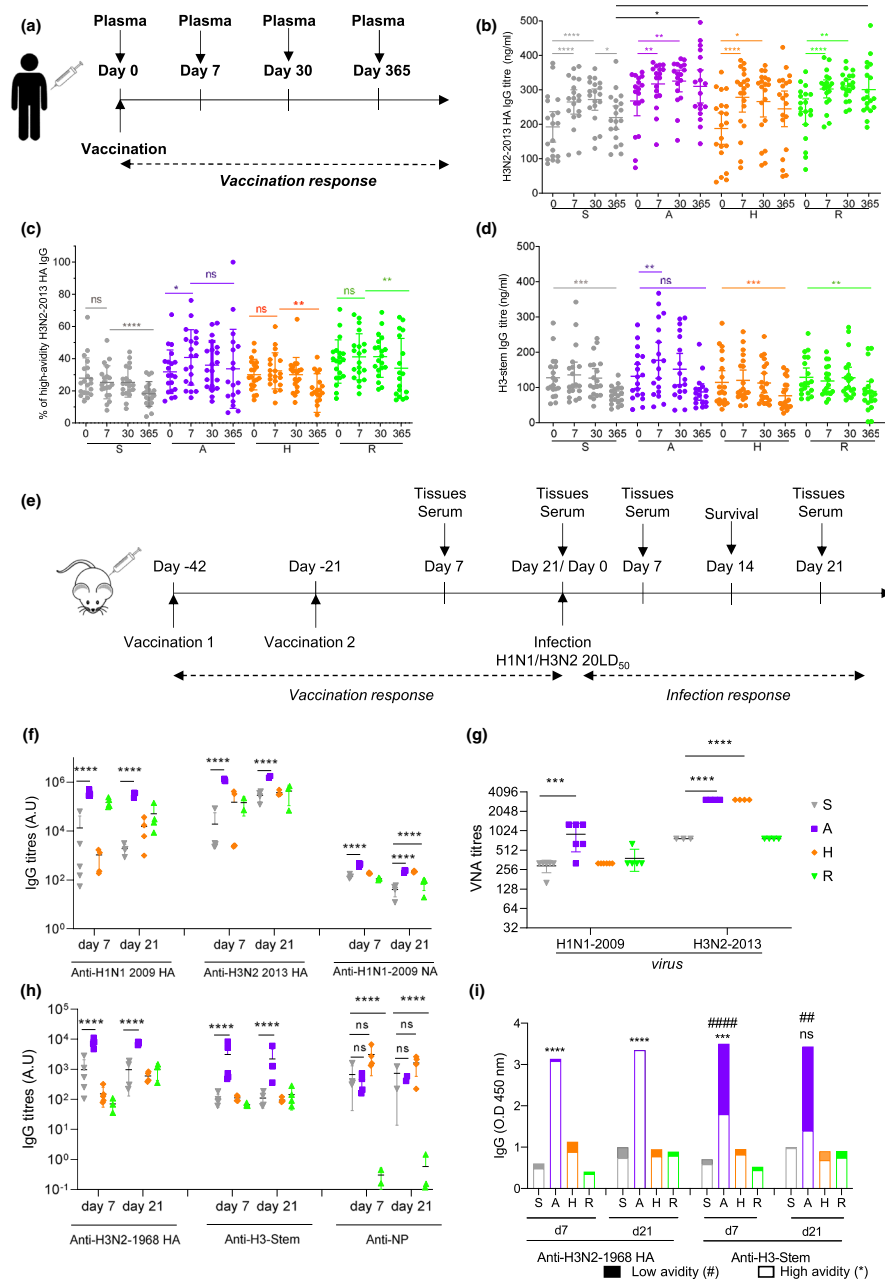
Cross-reactive antibodies for an antigenically older H3N2 virus (H3-1968), conserved proteins (NP) and functionally conserved targets (H3-stem

and H1-stem) were also assessed (Figure 1h and Supplementary figure 1b). Again, only A-eIIV compared with S-IIV had significantly elevated IgG for H3N2-1968 HA, H3-stem (Figure 1h) and H1-stem (Supplementary figure 1b). Cross-reactive NP-specific IgG responses were comparable for S-IIV, A-eIIV and H-eIIV (Figure 1h). As the recombinant HA protein vaccine, R-eIIV, also does not contain NP, no substantial NP-specific IgG responses were detected. Cross-reactive H3N2-1968 HA IgG responses by A-eIIV were predominantly high-avidity and significantly increased in magnitude (Figure 1i), whilst both high- and low-avidity H3-stem-specific IgG were increased for A-eIIV ( $P = 0.003$ ; Figure 1i).

### A-eIIV increases memory B-cell and long-term antibody responses in mice

We next investigated the effects of eIIV on the generation of isotype-switched memory B-cell responses ( $\text{B220}^+ \text{CD95}^+ \text{Fas}^{\text{low}} \text{IgD}^- \text{IgG/IgM}^+$ ) (Figure 2a), which are important for long-term antibody production. Shortly after vaccination (day 7), there was no significant increase in total  $\text{IgM}^+$  or  $\text{IgG}^+$  B memory cells in the vaccination site draining inguinal lymph node (iLN) for S-IIV compared with unvaccinated PBS mock-treated mice (Figure 2b and c). Meanwhile, there was a significant induction of both  $\text{IgG}^+$  and  $\text{IgM}^+$  memory B cells in by A-eIIV compared with S-IIV at day 7, which was maintained at 21 days post-vaccination.

To assess post-vaccination waning, the kinetics of antibody responses at 2, 4 and 6 months were assessed longitudinally (Figure 2d–f). All vaccines induced stable long-term H3-specific IgG until 6 months post-vaccination; however, A-eIIV was consistently superior for increased magnitude and avidity of cross-reactive H3N2-1968 HA (Figure 2e) and H3-stem responses (Figure 2f).



**Figure 1.** A-eIV induces high-avidity H3-2013 and H3-stem IgG in humans and mice. A randomised clinical trial to assess eIV immunogenicity compared with S-IIV, sampled blood plasma at days 0 (baseline), 7, 30 and 365 after vaccination **(a)**. H3N2-2013 HA titres **(b)**, high-avidity H3N2-2013 HA **(c)** and H3-stem-specific **(d)** IgG responses were measured by ELISA. The same vaccines were assessed in a mouse model of vaccination and infection **(e)**. Vaccine strains, H1N1-2009 and H3N2-2013 HA-specific and H1N1-2009 NA-specific, day 7 and 21 post-vaccination IgG titres by ELISA **(f)** and day 21 post-vaccination by VNA **(g)**. Cross-reactive, H3N2-1968, H3-stem HA-specific and NP-specific, day 7 and 21 post-vaccination IgG responses by ELISA **(h)**. Proportion of low- and high-avidity H3N2-1968 HA and H3-stem-specific IgG **(i)**. Data represent the mean, SEM and individual responses, **(b–d)**  $n = 20$  per group and **(f–i)**  $n = 4$  or 5 per group. For **b–d**, \*shows statistical significance by one-way ANOVA with the Friedman test in each vaccine group day 0 versus days 7, 30 and 365. For **f–i**, \*shows statistical significance by one-way ANOVA (mixed model) for eIV versus S-IIV by Tukey’s multiple comparison test. # or \* $P < 0.05$ , ## or \*\* $P < 0.01$ , ### or \*\*\* $P < 0.005$ , #### or \*\*\*\* $P < 0.001$ , \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.005$ , experiments were repeated twice, ns, not significant. Human serological experiments were performed with  $n = 20$  individuals/group, experiments were repeated twice. Mouse experiments were performed with individual mouse serum from 4–5 mice, and experiments were repeated twice.

### Increased class switch and hypermutation-associated transcripts in A-eIIIV B cells

A-eIIIV elicited stable long-lived high-avidity antibodies to vaccine strains and cross-reactive HA proteins, H3N2-1968 and H3-stem. Therefore, we investigated further the germinal centre (GC) B-cell reaction and associated CD4<sup>+</sup> T follicular helper (Tfh)-cell responses in the iLN induced by the eIIIVs (Figure 3a). A-eIIIV elicited a significantly increased Tfh response, which was more than double that of S-IIV at day 7 and still ongoing at day 21 (Figure 3b). Concurrently, the GC B-cell response was also doubled at day 7 in the A-eIIIV group compared with S-IIV responses (Figure 3c). At day 21, Tfh and GC cell counts decreased by 30-fold in most vaccine groups, but the A-eIIIV group still maintained significantly higher GC responses than S-IIV ( $P = 0.02$ ; Figure 3c). In comparison with S-IIV, H-eIIIV and R-eIIIV vaccination did not display any significant increase in GC or Tfh cells (Figure 3b and c).

A-eIIIV increased the avidity and magnitude of cross-reactive antibody responses, which was accompanied by increased Tfh and GC cellular responses in the iLN, suggesting an early and enhanced somatic hypermutation (SHM) and class switch recombination (CSR) response. Therefore, we assessed the transcription levels of *Aicda* for activation-induced cytidine deaminase (AID) enzyme, which is responsible for the B-cell receptor maturation through the SHM process in the light zone of LN.<sup>19</sup> Expression of AID was increased in iLN B cells at 7 days after A-eIIIV compared with S-IIV ( $P < 0.0001$ ; Figure 3d). Additional transcription factors controlling GC formation and AID activation, namely *Bcl6*, *Pax5* and *c-Myc*,<sup>20</sup> were also significantly increased in A-eIIIV B-cell responses compared with S-IIV (Figure 3d).

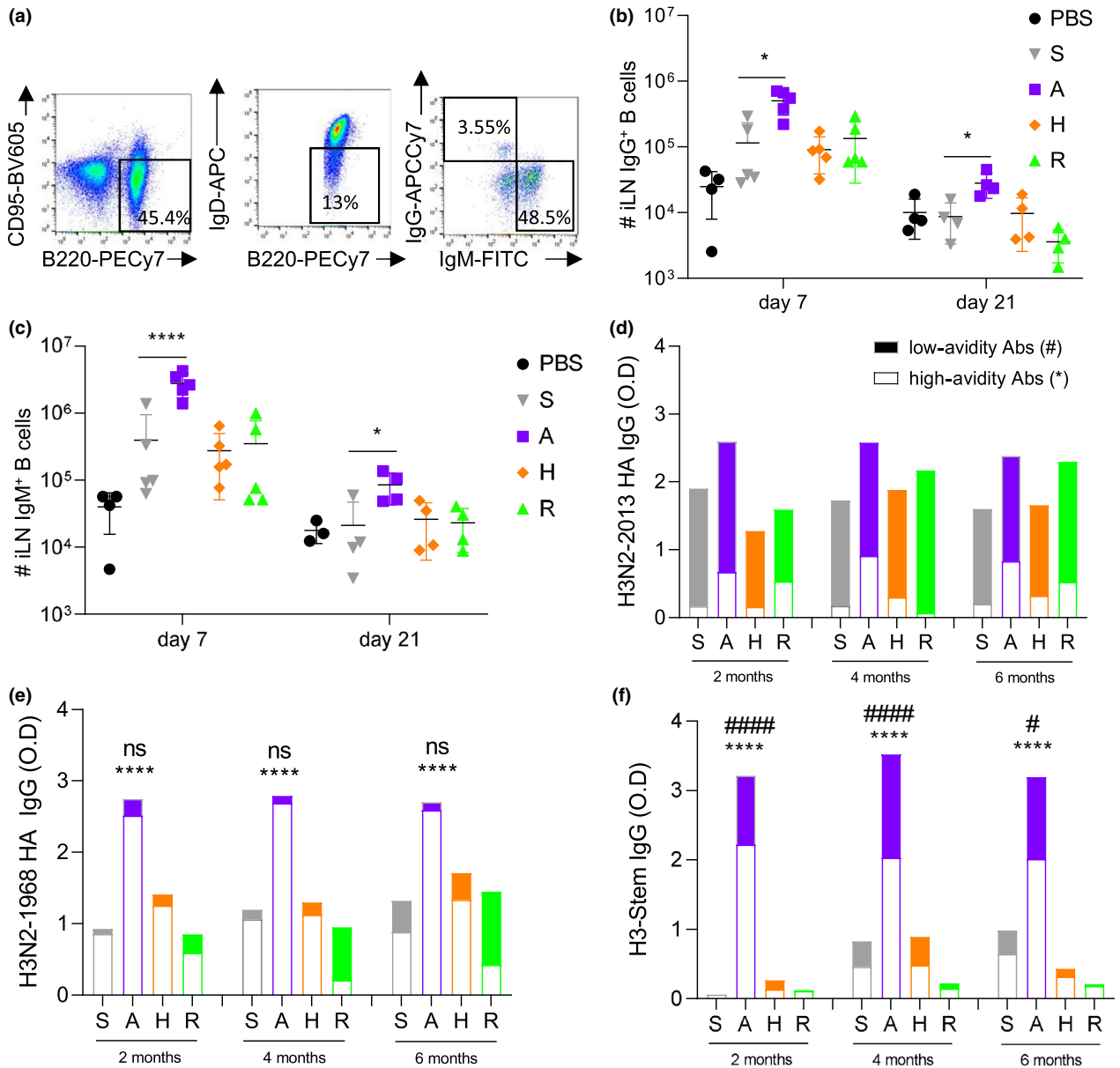
As AID is also responsible for the CSR that takes place in the light zone of LN, we determined the level of class switching (from IgM to IgG responses) at days 7 and 21 post-vaccination using ELISA endpoint titrations of H3-2013 HA-specific IgG and IgM responses in mouse serum (Figure 3f and g and Supplementary figure 2). At day 7, the S-IIV H3N2-2013 HA response was predominantly IgM (66%) rather than IgG (34%), whereas the A-eIIIV response was predominantly IgG (59%) rather than IgM (41%) (Figure 3h). This indicates an earlier switch from the default IgM isotype towards the IgG isotype by A-eIIIV. By day 21, the S-IIV H3N2-2013 HA IgG response increased (55% IgM, 45% IgG) but

was still significantly lower than the A-eIIIV response (31% IgM, 69% IgG) (Figure 3h and Supplementary figure 2). Similar results were obtained for the cross-reactive H3N2-1968 HA-specific antibodies, consolidating an earlier class switch induced by the A-eIIIV for cross-reactive responses as well (Supplementary figure 2). The H-eIIIV and R-eIIIV, H3N2-2013 HA IgM and IgG switch ratios were comparable to S-IIV at both time points (Figure 3h). Also, the H3N2-1968 HA IgG subclass response observed in the A-eIIIV group of mice was increased for both IgG1 and IgG2a compared with the S-IIV, and for H3-stem for IgG1, IgG2a and IgG2b, which are associated with greater effector functions and maturation of B-cell selection,<sup>21</sup> suggesting an increased IgG subclass maturation in response to A-eIIIV (Supplementary figure 3).

### Increased HA stem neutralisation by A-eIIIV but comparable HA landscape to S-IIV

The A-eIIIV vaccine in mice and humans elicited significantly higher magnitude of antibodies binding to the H3-stem (Figure 1d, h, and i) and H1-stem (Supplementary figure 1b). We determined whether these antibodies were also neutralising using a chimeric H9/1N3 virus that contains a H9 HA head domain (A/guinea fowl/Hong Kong/WF10/99) and a H1-stem domain (A/Puerto Rico/8/1934) for H1-stem-mediated neutralisation.<sup>22</sup> The day 7 VNA titres against cH9/1N3 by A-eIIIV were significantly increased compared with S-IIV response (Figure 4a); therefore, HA stem binding antibodies stimulated by A-eIIIV are also neutralising.

A H3N2-1968 HA yeast surface display (YSD) library in combination with mouse serum from day 7 post-vaccination was used to confirm and map the presence of cross-reactive anti-H3N2-1968 HA antibodies from the 2017/2018 vaccination. Assay controls showed specific binding by H3N2-1968 HA convalescent serum (0.82% HA-YSD<sup>+</sup>) and minimal background from PBS-vaccinated mice (0.05% HA-YSD<sup>+</sup>) (Figure 4b). However, we did not detect enough HA-YSD binding in the serum from S-IIV, H-eIIIV or R-eIIIV (Figure 4b) for further HA-YSD mapping, whilst A-eIIIV serum had a reliable population for HA-YSD mapping (1.4% HA-YSD<sup>+</sup>; Figure 4bc). HA-YSD sequence analysis showed similar H3N2-1968 HA binding of the A-eIIIV serum compared with the H3N2-1968-positive serum (Figure 4d), without an enrichment of HA stem antibodies despite cH9/1N3 neutralisation and H3-stem binding.

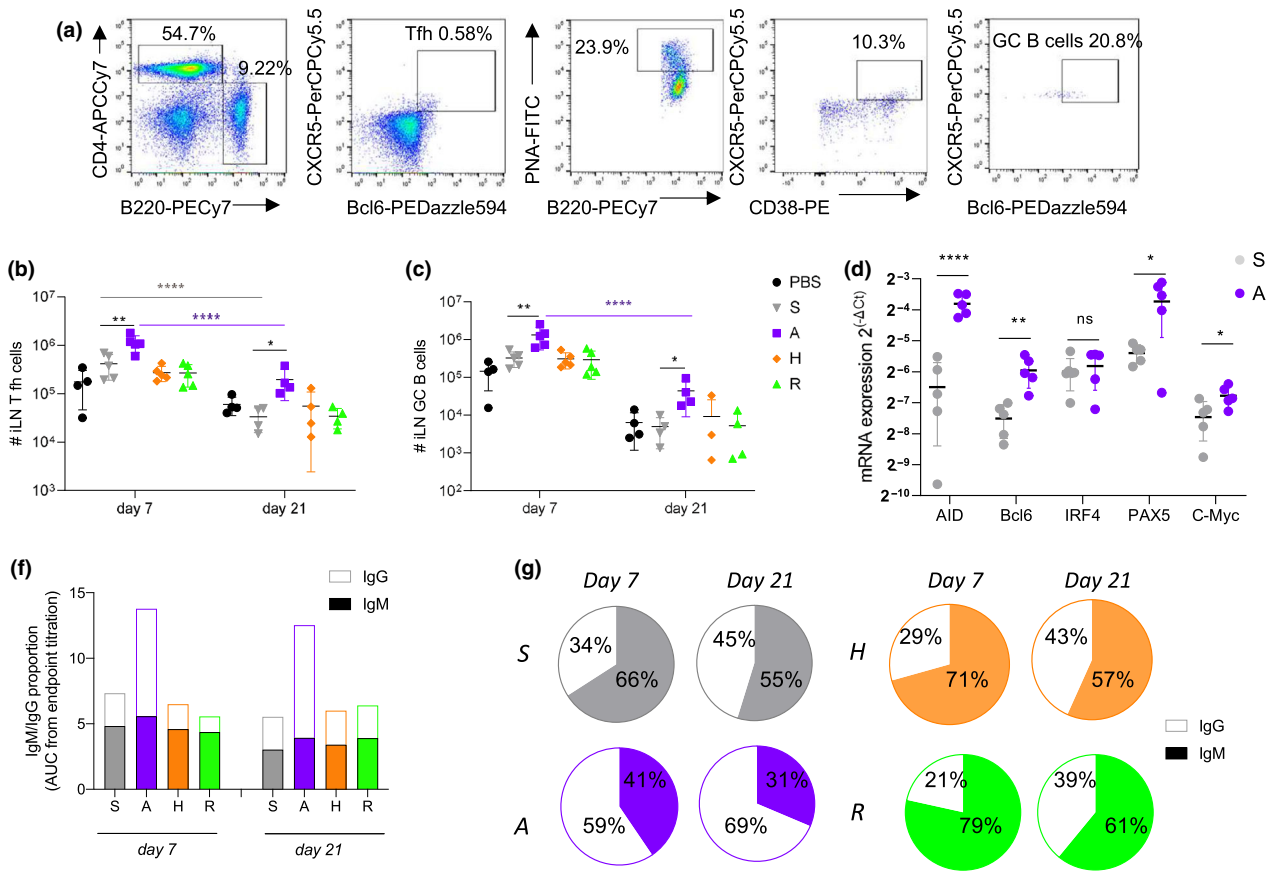


**Figure 2.** Memory B-cell responses and long-term antibody responses are increased in A-elIV-vaccinated mice. BALB/c mice were vaccinated twice i.m. with either PBS, S-IIV, A-elIV, H-elIV or R-elIV vaccines 3 weeks apart. iLN was collected at days 7 and 21 post-vaccination to quantify by FACS (a), IgG<sup>+</sup> (b) and IgM<sup>+</sup> (c) B memory cells. Longitudinal serum samples were serially sampled from individual mice at 2, 4 and 6 months post-vaccination for H3N2-2013 (d), H3N2-1968 (e) and H3-stem (f) IgG by ELISA for avidity. Data represent the mean, SEM and individual responses, *n* = 4–8 mice per group. For b, c, \*shows statistical significance for elIV versus S-IIV by one-way ANOVA (mixed model) and Tukey’s multiple comparison test. For d–f, #shows statistical significance for low-avidity antibodies and \*for high-avidity antibodies for elIV versus S-IIV by one-way ANOVA (mixed model) and Tukey’s multiple comparison test. # or \**P* < 0.05, and ##### or \*\*\*\**P* < 0.001, ns, not significant. Experiments were repeated twice. Representative FACS plots were gated on lymphocytes for FSC/SSC and live/dead (a).

**In vitro protection by mouse and human serum by A-elIV against heterologous viruses**

Because of the presence of HA stem antibodies and cross-reactivity against H3N2-1968, we

assessed further cross-reactivity against an avian influenza virus, H7N7, another group 2 influenza virus. Firstly, A-elIV elicited significantly increased H7-HA-specific IgG response compared with S-IIV (Figure 5a), which correlated with the group 2 HA stem responses (Figure 5b, *R*<sup>2</sup> = 0.6246). An

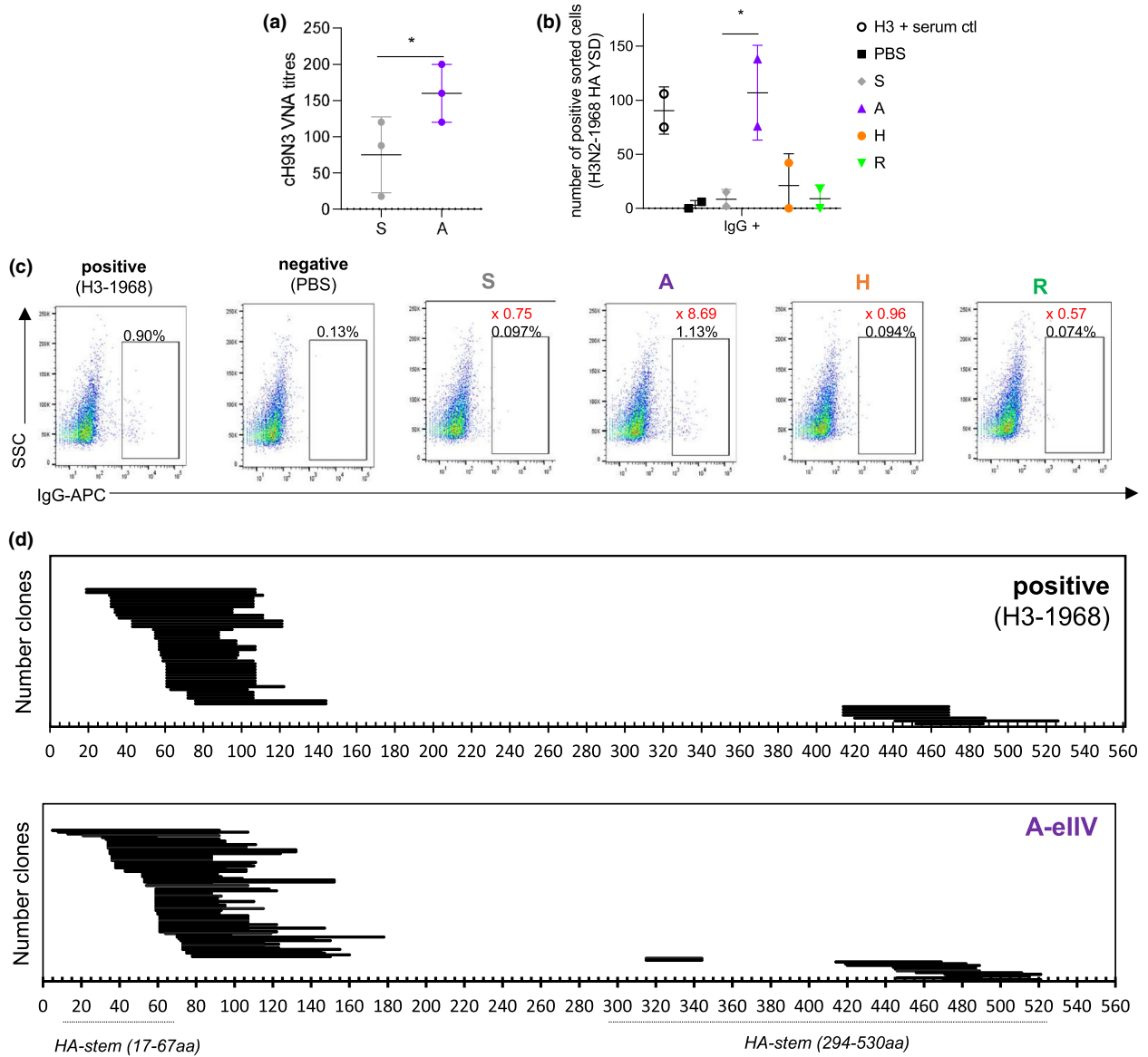


**Figure 3.** Elevated CD4<sup>+</sup> Tfh and GC B cells by increased expression of SHM and CSR genes lead to early IgG class switch for A-eIV. Vaccine responses were assessed by FACS **(a)** for CD4<sup>+</sup> Tfh cells **(b)** and GC B cells **(c)** at days 7 and 21 post-vaccination in the iLN. Expression of AID, Bcl6, IRF4, Pax5 and c-Myc in B cells from iLN (relative expression to GAPDH expression, expressed in 2<sup>ΔCt</sup>) at day 7 post-vaccination **(d)**. Quantity **(e)** and proportion **(f)** of H3N2-1968 HA-specific IgM and IgG at days 7 and 21 post-vaccination, measured by ELISA AUC endpoint titres (Supplementary figure 2). Data represent the mean, SEM and individual responses, n = 4 or 5 mice per group. \*shows statistical significance by one-way ANOVA (mixed model) for eIV versus S-IIV, \*P < 0.05, \*\*P < 0.01 and \*\*\*\*P < 0.001 experiments were repeated twice, ns, not significant. Representative FACS plots were gated on lymphocytes for FSC/SSC and live/dead **(a)**.

*in vitro* serum protection assay by FACS (Figure 5c) showed that the increased A-eIV cross-reactive antibody responses also lead to a significant reduction of H7N7-infected cells relative to naïve mouse serum, whilst S-IIV had no *in vitro* serum protection against H7N7 and was comparable to naïve mouse serum (Figure 5d). In the same *in vitro* protection FACS assay, the human antibody responses after S-IIV and A-eIV vaccination showed equivalent serum protection against homologous H3N2-2013 virus; however, A-eIV responses significantly reduced cells infected with heterologous viruses, H3N2-1968 and H7N7, reflecting the high level of antibody cross-reactivity induced by A-eIV (Figure 5e and f).

### Recall of H3-specific T and B cells by A-eIV during heterologous influenza challenge in mice

Human and mouse data for A-eIV have shown increased H3-HA-specific antibodies that are high-avidity and cross-reactive, but the protective potential of these responses is not known. Therefore, to determine vaccine-mediated protection we challenged eIV mice with a lethal dose of H3N2-1968. We selected the H3N2-1968 virus for mouse challenge as a well-established model of heterosubtypic protection in T cell<sup>23</sup> and HA stem-based vaccine models.<sup>24</sup> The H3N2-1968 virus has 85.3% amino acid similarity to the HA of the H3N2-2014 virus in the 2017/2018 vaccines



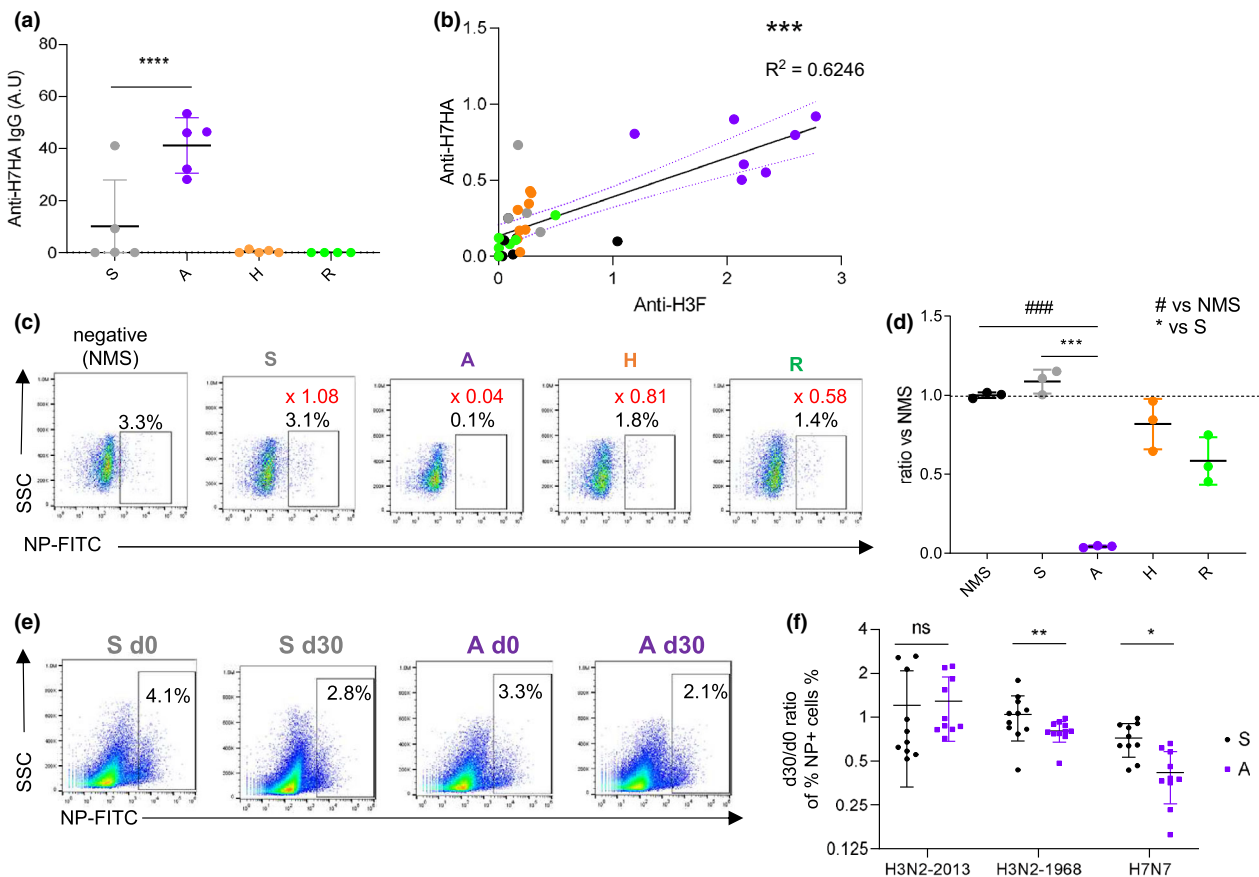
**Figure 4.** HA stem antibodies are neutralising and map to the HA stem for A-eIV responses. VNA titres for cH9/1N3 virus measuring HA stem-mediated neutralisation of A-eIV and S-eIV serum from day 7 post-vaccination **(a)** ( $n = 4$  pooled serum in triplicate). YSD H3N2-1968 library binding to eIV and control pooled H3N2-1968 convalescent mouse serum **(b)** for FACS sorting. Mapping of the yeast clone HA amino acid sequence by serum binding for A-eIV ( $n = 85$  clones) and recovered H3N2-1968 ( $n = 48$  clones) serums **(c)**. YSD binding **(b, c)**, pooled serum from  $n = 5$  mice per group. In **(d)**, the percentage of the population of interest is shown in black on the FACS plots, and the fold change of the percentage versus PBS is shown in red. \* $P < 0.01$ , experiments were repeated twice, and HA stem amino acid positions are as previously described.<sup>29</sup>

used. Challenge experiments were performed at 6 months post-vaccination (long-term memory) to study the potential waning of vaccine responses (Figure 6), and at 21 days after vaccination (short term; Supplementary figure 4a–c).

Challenge of mice with H3N2-1968 virus at 6 months after vaccination induced a significant weight loss in unvaccinated mice, whilst all vaccinated mice were still protected with no

lethality and no differences between weight loss between S-eIV and eIV (Figure 6a). Therefore, our definition of protection is based on lung viral loads and damage to the lung by total protein concentration. Importantly, A-eIV and H-eIV exhibited earlier viral clearance in the lungs by day 7 than S-eIV (Figure 6b). Meanwhile, S-eIV-mediated protection had waned and viral loads were comparable to unvaccinated PBS mice





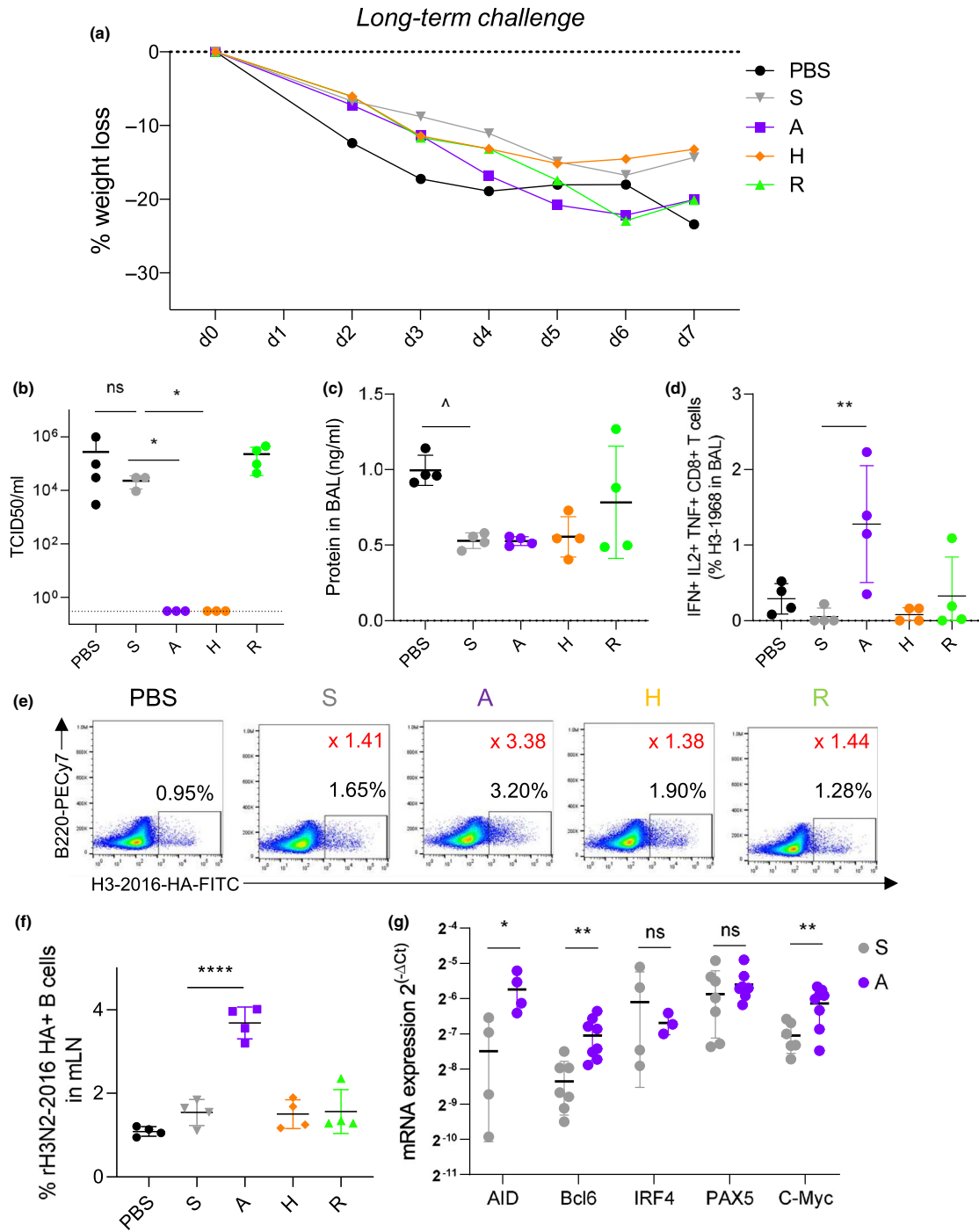
**Figure 5.** H7N7 cross-reactive A-eIV antibodies in mice and humans. H7N9-HA-specific IgG titres measured by ELISA in mice at 7 days post-vaccination ( $n = 5$  mice per group) (a). Correlation of H7N9-HA and H3-stem IgG titres in mice serum by the Pearson correlation,  $R^2 = 0.624$ ,  $P < 0.0001$  (b). *In vitro* protection assay of H7N7-infected Raji cells with mouse serum from 21 days post-vaccination (pooled serum in triplicate,  $n = 3$  mice per group) assessed by FACS (c), and responses were normalised to naïve mouse serum (d). *In vitro* protection assay of homologous H3N2-2013, heterologous H3N2-1968 or H7N7-infected Raji cells with human pre- and 30-day post-vaccination sera (e), individual responses ( $n = 10$  per group) for S-IIV and A-eIV, and fold change post-vaccination (f). Experiments were repeated twice. Data represent the mean, SEM and individual responses. In c, the percentage of the population of interest is shown in black on the FACS plots, and the ratio of the mean percentage versus naïve mouse serum (NMS) is shown in red. #shows statistical significance versus NMS, and \*shows statistical significance versus S-IIV. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.005$ , and \*\*\*\* $P < 0.001$ , ns, not significant.

(Figure 6b); however, there was a trend for less damage to the lung (by total protein concentration) for S-IIV mice compared with the PBS groups ( $P = 0.009$ ; Figure 6c).

Similar observations were made at short-term challenge with H3N2-1968 and H1N1-2009. In the short-term challenge, no difference was observed between S-IIV and eIV in terms of weight loss, but the local lung damage (by total protein concentration) was reduced in A-eIV and H-eIV compared with S-IIV, whereas R-eIV was elevated compared with S-IIV (Supplementary figure 4). Homologous short-term challenge with H1N1-2009 virus also showed protection for all vaccines and a strong superiority for the A-eIV in terms of VNA

titres and H1N1-2009 HA-specific antibodies but comparable serum antibody ADCC activity (Supplementary figure 5).

Differences in inflammation and viral clearance after influenza challenge at long term may indicate changes in cellular responses; therefore, we assessed recall of influenza-specific T cells (Figure 6d) and B cells (Figure 6e and f). Following influenza virus challenge, A-eIV mice showed increased H3N2-1968-specific polyfunctional  $CD8^+$  T cells ( $IFN-\gamma^+ TNF-\alpha^+ IL-2^+$ ) in the bronchoalveolar lavage (BAL), whilst no other vaccination group had any substantial T-cell response (Figure 6d, A-eIV versus S-IIV,  $P = 0.005$ ), whereas H3N2-1968-specific  $CD4^+$  T-cell responses were comparable



**Figure 6.** Influenza-specific T- and B-cell recall responses increased by A-eIIV after H3N2-1968 challenge. Vaccinated mice were challenged i.n. with 20LD<sub>50</sub> of H3N2-1968 virus at 6 months (long-term memory) post-vaccination, and at day 7 post-infection, lungs and BALF were collected. Long-term memory H3N2-1968 challenge weight loss (a), lung viral load (b) and protein concentration in BALF (c). By FACS, polyfunctional H3N2-1968-specific CD8<sup>+</sup> T cells (IFN- $\gamma$ <sup>+</sup> TNF- $\alpha$ <sup>+</sup> IL-2<sup>+</sup>) were measured in the BAL (d). By FACS (e), fluorescent-labelled recombinant HA probes for H3-2016-specific B cells were measured in the mLN (f). Expression of AID, Bcl6, IRF4, Pax5 and c-Myc in B cells from spleen (relative expression to GAPDH expression, expressed in 2<sup>- $\Delta\Delta Ct$</sup> ) (g). Data represent the mean, SEM and individual responses of n = 4 mice per group. In e, the percentage of the population of interest is shown in black on the FACS plots, and the ratio of the mean percentage versus PBS is shown in red. ^shows statistical significance of S-IIV versus PBS. \*shows statistical significance of eIIV versus S-IIV. \*P < 0.05, \*\*P < 0.01 and \*\*\*\*P < 0.001, experiments were repeated twice, ns, not significant.

between the vaccination groups in the lung draining mediastinal lymph node (mLN) (Supplementary figure 4d and e). Furthermore, in the mLN, only A-eIIV elicited a substantial recall of the H3N2-2016 HA-specific B-cell response (Figure 6e and f), which was twice the magnitude of S-IIV (Figure 6f,  $P < 0.0001$ ). The A-eIIV B-cell recall responses at 6 months post-vaccination also maintained elevated expression of B-cell CSR- and SHM-promoting responses for AID, Bcl6 and c-Myc versus S-IIV (Figure 6g).

### Recall of high-quality antibody responses of A-eIIV short- and long-term memory

To investigate the local inflammatory response, we performed a 12-panel cytokine bead array for Th1, Th2 cytokines and chemokines (Figure 7a) in BAL after challenge. S-IIV had a significant fold-change increase in pro-inflammatory IFN- $\gamma$ , IL-6, TNF- $\alpha$  and IL-2 and anti-inflammatory IL-4 compared with unvaccinated PBS mice (Figure 6d), whilst A-eIIV compared with S-IIV induced a significant increase in pro-inflammatory IFN- $\gamma$ , TNF- $\alpha$ , IL-9 and IL-2, which may drive viral clearance in the lung, and IL-4 driving a Th2 response. Further, H-eIIV had an opposite effect compared with S-IIV, with a significant reduction of both types of Th1 and Th2 cytokines, and also of the CCL2 and CCL4 chemokines, which may lead to reduced recruitment of immune cells to the lung. Finally, R-eIIV had comparable responses to S-IIV.

As antibody quality is a consequence of AID activity, the properties of immune sera at influenza challenge time points were further assessed. The H3N2-1968 VNA titre at day 7 post-infection was comparable between unvaccinated PBS mice and S-IIV (Figure 7b), suggesting no recall of existing vaccine antibodies by heterologous H3N2-1968 challenge; however, A-eIIV had significantly increased early VNA responses during infection. Our FACS-based *in vitro* serum protection assay also showed an early increased day 7 neutralisation response for A-eIIV and H-eIIV (Figure 7c), whilst other vaccine groups only developed *in vitro* neutralisation activity later by day 21 post-challenge when virus is well cleared. Importantly, the local mucosal IgA H3N2-1968 HA response from the BAL of the lung of A-eIIV mice showed significant early recall of vaccine antibody responses, with a significant H3N2-1968 HA IgA response at the site of infection at short-term post-vaccination challenge,

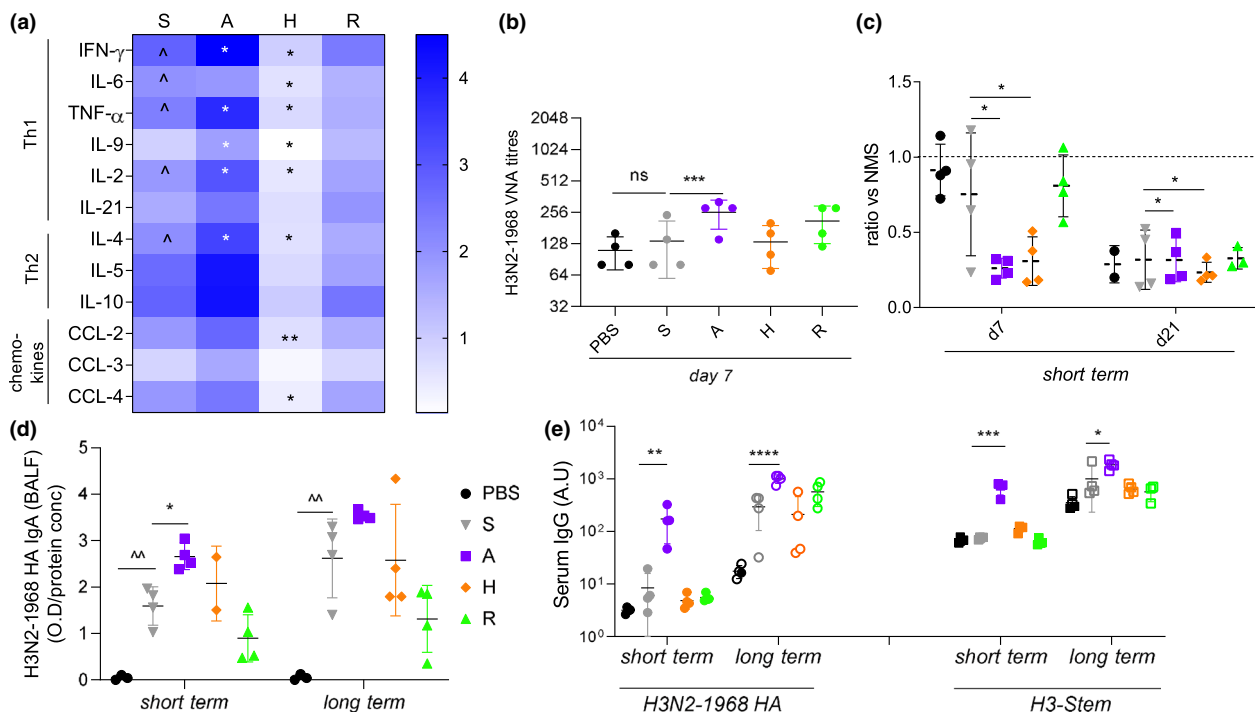
which was not observed in other vaccination groups, and a trend for increased IgA responses at long-term challenge compared with S-IIV (Figure 7d). Challenge at both short- and long-term post-vaccination time points showed A-eIIV had significantly increased IgG titres and avidity of cross-reactive antibody responses against H3N2-1968 HA (Figure 7e and Supplementary figure 4g), H3-stem (Figure 7e) and NP (Supplementary figure 4h) compared with S-IIV.

## DISCUSSION

Influenza virus is the only pathogen that is recommended for yearly vaccination, with over 500 million doses used globally every year.<sup>25</sup> IIV has limited protection against infecting strains that are antigenically distinct from those contained in the vaccine. This can create a public health problem, such as recent H3N2 viruses, which have been more difficult to match and control with S-IIV.<sup>26</sup> Newly available vaccine approaches, including eIIVs, could be utilised to provide longer duration and breadth of protection, as broadly protecting vaccines are needed, but no universal influenza vaccine is licensed yet.<sup>27,28</sup> We therefore compared the immunogenicity, mechanism of action and protective potential of three commercially available enhanced vaccines, FluAd (A-eIIV), FluZone-HD (H-eIIV) and FluBlok (R-eIIV), with a standard seasonal influenza vaccine, FluQuadri (S-IIV), in humans and mice. We found evidence to indicate that A-eIIV and H-eIIV may provide longer-lasting and broader cross-protection against influenza viruses than S-eIIV and R-eIIV.

Results from our randomised trial of eIIVs in older adults showed that A-eIIV increased the long-term avidity of H3-2013-specific HA IgG early after vaccination in conjunction with an increased IgG response towards H1-stem and H3-stem and, to a lesser extent, H-eIIV also increased H1-stem IgG responses. The HA stem of the HA protein harbours functionally conserved neutralising epitopes, which may enable broader protection against different influenza strains,<sup>29</sup> rendering the induction of anti-HA stem antibodies by A-eIIV and partly H-eIIV of particular interest for increasing the breadth of protection by influenza vaccination strategies.

Our randomised clinical trial was designed to assess immunogenicity, not vaccine efficacy against clinical outcomes. Therefore, we assessed



**Figure 7.** Lung elevated cytokine production and secretory-specific IgA, along with high serum H3-stem antibodies in A-eIV mice after challenge. Mice were challenged with 20LD<sub>50</sub> H3N2-1968 virus 21 days or 6 months post-vaccination, and inflammatory responses were sampled at day 7 post-infection. Inflammatory cytokine and chemokine concentrations in individual lung homogenates ( $n = 4$ ) by CBA were normalised to fold change from PBS response **(a)**. H3N2-1968 VNA endpoint titres from serum from short-term challenge at day 7 post-infection **(b)**. *In vitro* protection assay of H3N2-1968-infected Raji cells with mouse serums from 7 and 21 days post-infection ( $n = 3$  or 4), responses were normalised to naïve mouse serum **(c)**. Day 7 post-challenge, H3N2-1968 HA-specific secretory IgA in BALF for short- and long-term memory recall measured by ELISA **(d)**, normalised to total protein concentration by BCA. H3N2-1968 HA and H3-stem-specific IgG responses by ELISA for short- and long-term memory recall at day 21 post-challenge **(e)**. Data represent the mean, SEM, individual responses,  $n = 4$  mice per group. \*shows statistical significance of eIV versus S-IV. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.005$  and \*\*\*\* $P < 0.001$ , experiments were repeated twice, ns, not significant.

S-IV versus three different eIVs in parallel in mice followed by challenge with a lethal heterologous influenza strain, enabling access to infected tissues and immune sites that are constrained in human studies. In addition, the naïve mouse status enables direct comparison of direct vaccine immunogenicity without interference of prior immunity. Whilst IgG subclass function, age, exposure history, vaccine dosage and duration of responses may differ between mice and humans, the mouse model is an important but limited resource to benchmark vaccine responses.

Our mouse vaccination results showed an increase in the avidity and magnitude of antibodies targeting the vaccine H3N2-2013 HA, H1N1-2009 HA and NA, and more importantly an induction of cross-reactive H3N2-1968 HA and H3-stem antibodies by A-eIV. We further confirmed the induction of antibody cross-reactivity by the induction of neutralising anti-HA stem antibodies

using a chimeric H9/1N3 virus.<sup>22</sup> Furthermore, challenge of mice with the heterologous H3N2-1968 virus showed substantial early recall of IgA antibodies to the site of infection. Altogether, these data reinforce the findings on the effect of the MF59 adjuvant of A-eIV increasing the breadth and cross-reactivity of vaccine responses.<sup>11</sup> MF59 is an oil-in-water emulsion that enhances the persistence of the vaccine antigens at the injection site and subsequently increases the activation of antigen-presenting cells.<sup>30</sup> We found that MF59 induces an earlier and higher expression of AID, the enzyme responsible for antibody SHM and CSR in the GC, together with Bcl6, Pax5 and c-Myc, leading to an elevated GC reaction and magnitude of Tfh- and GC B-cell responses. Because of the enhanced expression of AID and related transcription factors involved in GC development, we observed an enhancement of the avidity of cross-reactive H3N2-1968 HA and

H3-stem IgG by A-eIIIV, and an earlier class switch from the default IgM towards the IgG isotype, which did not occur in other vaccine groups. Expression of AID and other genes of the CSR process plays a vital role for protection from influenza infections, as AID<sup>-/-</sup> mice showed severe morbidity with delayed virus clearance compared with AID<sup>+/-</sup> mice.<sup>31</sup> As these important B-cell transcription factors are induced in the local LN by A-eIIIV and not S-IIIV, the adjuvant promotes the GC reaction, SHM and CSR to generate high-quality antibody responses.

The earlier and more intense induction of GC B cells by A-eIIIV and not H-eIIIV or R-eIIIV may also be as a result of the persistent deposition of the antigen in the LN by follicular dendritic cells facilitated by the MF59 adjuvant, as sustained antigen availability has previously been shown to increase the number of GC and Tfh in the LN.<sup>32,33</sup> A-eIIIV also induced larger memory B-cell responses than the high-dose vaccines, H-eIIIV and R-eIIIV. Previous vaccine studies for hepatitis B and pneumococcal vaccines have also reported that low doses of antigen tend to favor immune memory<sup>34–36</sup> over higher antigen doses, whilst we did not see this for S-IIIV, the lower dose and use of adjuvant of A-eIIIV may have favored the establishment of long-term immune memory responses. The formation of long-term memory is an advantage for extended seasonal protection against influenza infection.

Vaccine-mediated protection was further assessed by lethal H3N2-1968 challenge at long-term (6 months) and short-term (21 days) memory time points post-vaccination. Consistent with our vaccine immunogenicity results, A-eIIIV showed superiority during infection by reducing lung inflammation and increasing Th1 and Th2 cytokines that are key for viral clearance.<sup>37,38</sup> The H3N2-1968 HA-specific antibody response from A-eIIIV, most importantly lung IgA, was recalled at a higher magnitude and avidity compared with other vaccine groups. Influenza-specific polyfunctional CD8<sup>+</sup> T cells and B cells were also selectively recalled by the A-eIIIV long-term memory response, unlike other vaccine groups.

HA stem-specific antibodies have shown remarkable cross-reactivity in some cases for different strains, subtypes and even pan-influenza reactivity,<sup>39</sup> and clinical trials are underway for HA stem vaccines to induce broadly reactive antibodies.<sup>40</sup> A-eIIIV induced HA stem antibodies that may contribute to H3N2-1968 cross-reactivity

and also bring an additional level of protection against other IAV subtypes such as avian H7N7.<sup>41</sup> From our *in vitro* protection assay, we saw that A-eIIIV dramatically reduced H7N7-infected cells after vaccination in mice and humans, whilst other vaccine responses did not. These *in vitro* antiviral responses could possibly be contributed to by antibodies specific to the H3-stem and NP proteins. Antibodies targeting the highly conserved internal NP were also highly induced after vaccination in most vaccines, excluding R-eIIIV that does not contain NP, and A-eIIIV-treated mice had greater recall of NP antibodies during influenza challenge. The NP protein has an even higher level of sequence conservation than the HA stem (95% versus 70–40%), and is expressed on the surface of infected cells and internally of virions,<sup>42</sup> and could therefore be an additional important universal vaccine target.

Whilst A-eIIIV stimulated the strongest cross-reactive responses, H-eIIIV also had some promising results. H-eIIIV displayed interesting protective features in the challenge experiments with a reduction of the lung damage and all inflammatory cytokines, particularly the chemokine responses, CCL2 (MCP-1) and CCL4 (MIP-1 $\beta$ ), thus potentially reducing the local recruitment of inflammatory cells and deleterious inflammation, along with efficient viral clearance. H-eIIIV had an equivalent protective effect to A-eIIIV for reduction of the viral loads after long-term challenge, which illustrates sustained vaccine memory responses. Also, H-eIIIV compared with S-IIIV elicited significant day 21 post-vaccination antibody responses against the NA, NP proteins and H3-2013 virus by VNA in mice and H1-stem in humans.

Our study focused on H3N2 responses stimulated by eIIIV because of the recent lower vaccine efficacy against this virus subtype.<sup>26</sup> Some limitations of our study include the use of mismatched antigens to characterise vaccine immune responses. Because of the lack of commercial full-length HA proteins for the H3N2-2014 virus (A/Hong Kong/4801/2014), we used the H3N2-2013 HA protein from a recent virus (A/Switzerland/9715293/2013), which has over 98% amino acid similarity and high level of cross-reactivity. Furthermore, H3N2-2016 HA probes from B-cell staining at long-term challenge were from a different H3N2 strain again, H3-2016 (A/Singapore/INFIMH-16-0019/2016) – as these are precious non-commercial trimeric-HA fluorescent probes, we used that which was available to us. Challenge of

mice with a more closely related H3N2 virus may have yielded different results, but the use of H3N2-1968 virus is warranted to illustrate vaccine mismatch and heterologous protection. Despite a number of antigenic differences between the H3N2 vaccine and antigens within our immunological characterisation, A-eIIV showed continued superiority. Another limitation is the use of young mice (6 weeks old) and older adults (> 65 years) for the randomised clinical trial. The use of the mouse model was to study vaccine immunogenicity and protective potential directly rather than effect of age on vaccine usage and immunosenescence.

In Australia since 2018, H-eIIV and A-eIIV have been preferentially given to older adults,<sup>43</sup> whilst the UK switched to the A-eIIV for older adults in 2018/19. In the United States, the H-eIIV is now given to 40% of vaccinated older adults.<sup>44</sup> Therefore, no current consensus on vaccine utilisation has been achieved. In our initial report,<sup>18</sup> we identified an improved microneutralisation response against cell-grown H3N2 by R-eIIV compared with the A-eIIV and H-eIIV. Therefore, combining those earlier findings with the results of this study, we have found that each of the eIIVs has an advantage over S-IIV, with indications that the R-eIIV could provide better protection against vaccine-matched H3N2, whilst A-eIIV and H-eIIV provide better cross-protection against a broader array of influenza viruses. Future studies could examine whether combination strategies, for example alternating use of different eIIVs from year to year, could take advantage of the distinct strengths of the different eIIVs.

## METHODS

### Human subjects and randomised clinical trial of eIIV

We conducted a randomised trial to determine the vaccine immunogenicity of standard versus enhanced IIV in community-dwelling older adults, 65–82 years old, which was initiated before the 2017/2018 winter influenza season in Hong Kong (ClinicalTrials.gov NCT03330132). Older adults (Supplementary table 1) were randomly allocated to receive the 2017/2018 WHO-recommended IIV formulation of either standard FluQuadri IIV (S-IIV) or one of three enhanced vaccines adjuvanted (FluAd, A-eIIV), recombinant HA (FluBlok, R-eIIV) and high dose (FluZone high dose, H-eIIV) (Table 1).<sup>18</sup> Plasma was isolated from blood samples at baseline (day 0), and after vaccination at days 7 (acute) and 30 (short-term memory) (Figure 1a). Standard HAI

serological responses and T-cell responses are reported elsewhere.<sup>18</sup> All participants provided written informed consent. The study protocol was approved by the Institutional Review Board of the University of Hong Kong (UW:16-2014).

### Vaccination and infection of mice

To assess eIIV immunogenicity and vaccine-mediated protection, female 6-week-old BALB/c mice were vaccinated twice 21 days apart via the intramuscular (i.m.) route with either the S-IIV, eIIV or PBS (Figure 1e), with a third of a human dose in 100  $\mu$ L (Table 1). Mice were vaccinated with the Northern Hemisphere 2017/2018 containing A/Michigan/45/2015 (H1N1)pdm09-like virus, A/Hong Kong/4801/2014 (H3N2)-like virus and B/Brisbane/60/2008-like virus (Victoria lineage). S-IIV and R-eIIV were quadrivalent and contained a second influenza B virus, B/Phuket/3073/2013 (Yamagata lineage). For assessment of vaccination responses, mice were culled at day 7 or 21 after the second booster vaccination dose.

Vaccinated mice were challenged 21 days or 6 months after the booster dose, and anaesthetised mice were infected intranasally with a lethal dose (20LD<sub>50</sub>) of either H1N1-2009 (A/California/07/2009,  $1.375 \times 10^6$  pfu  $25 \mu$ L<sup>-1</sup>) or H3N2-1968 (A/Hong Kong/1/1968 MAC20,  $1 \times 10^4$  pfu  $25 \mu$ L<sup>-1</sup>), and culled at day 7 or 21 after challenge (Figure 1e). H1N1 (A/California/04/2009) virus was used for 'homologous' challenge representing the A/Michigan/45/2015 H1N1 vaccine component (HA amino acid conservation of 97%). Alternatively, to assess the breadth of vaccine-mediated protection from eIIV versus S-IIV, mice were challenged with a 'drifted' H3N2 strain, from 46 years earlier of the vaccine strain (A/Hong Kong/4801/2014) with A/Hong Kong/1/1968 (HK/68) for 'heterologous' challenge (HA amino acid conservation of 85.3%).

Blood was collected by cardiac puncture and clotted (MiniCollect, Greiner Bio-one, Kremsmünster, Austria), and serum was harvested after centrifugation and aliquoted and heat-inactivated, for 30 min at 56°C, before all *in vitro* experiments. To quantify virus replication, antibody and cellular responses, the lungs, BAL and lymphoid organs (spleen, mLN for challenged mice or iLN for vaccinated mice) were processed as previously described.<sup>23</sup> Lung viral titres were determined from homogenates by standard TCID<sub>50</sub> assay on MDCK cells, and viral titres were calculated by the Reed–Muench method, as previously described.<sup>23</sup> All experimental procedures were conducted in accordance with the standards and approved by the Committee on the Use of Live Animals in Teaching and Research, The University of Hong Kong.

### Antibody quantification by enzyme-linked immunosorbent assay (ELISA)

To assess influenza-specific antibodies from vaccination or infection, serum was probed in ELISA. Commercial proteins (Sinobiological, Beijing, China) to represent challenge viruses and vaccine strains included the following: H1N1-2009 HA (A/California/07/2009), H3N2-2013 HA (A/Switzerland/9715293/2013), H3N2-1968 HA (A/Hong Kong/1/1968), H7N9-

2013 HA (A/Anhui/01/2013), and NP (A/Switzerland/9715293/2013). Because of commercial availability, the H3N2-2013 protein (A/Switzerland/9715293/2013(H3N2)) was used to represent the H3N2 vaccine strain (A/Hong Kong/4801/2014 (H3N2)), which shared 98% HA amino acid homology and showed a high level of cross-reactivity.

The H1-stem (A/California/07/2009) and H3-stem (A/Aichi/2/1968) proteins (from Raghavan Varadarajan, Indian Institute of Science) were made as previously described.<sup>24,45</sup> Recombinant HA, NP proteins (at 80 ng mL<sup>-1</sup>) and HA stem protein (at 800 ng mL<sup>-1</sup>) were coated on 96-well flat-bottom immunosorbent plates (Nunc Immuno MaxiSorp, Roskilde, Denmark), in 100 µL coating buffer (PBS with 53% Na<sub>2</sub>CO<sub>3</sub> and 42% NaHCO<sub>3</sub>, pH 9.6) at 4°C overnight. An additional plate coated with a non-specific protein (blocking buffer, PBS with 5% FBS) was used to measure the background binding of each individual mouse serum and used as a background subtraction from protein-specific responses. Following FBS blocking and thorough washing, diluted serum samples (starting at 1:100) were bound for 2 h, further washed and then detected by an anti-mouse or anti-human Ig secondary antibody labelled with HRP specific for selected antibody classes or subclasses as indicated (IgG, or anti-mouse IgG1, IgG2a, IgG2b, IgA or IgM; Invitrogen, Carlsbad, CA, USA).

High-avidity antibodies were measured by an incubation 15 min with 8 M urea, after serum binding to remove low-avidity antibodies, and then, the remaining IgG response represents high-avidity antibodies.<sup>46</sup> TMB/peroxide was used as substrate, and the reaction was stopped by addition of sulphuric acid (R&D systems, Minneapolis, MN, USA), and optical density (O.D.) absorbance was read at 450 nm. A standard range was performed for all ELISAs as an interexperimental control, using pooled recovered post-infection mouse serum for H1N1-2009, H3N2-1968 or H7N7 viruses, and enabling the expression of results in arbitrary units (A.U.) relative to either H1, H3 or H7 proteins in A.U. Finally, to allow a quantitative comparison of the diverse IgG subclasses, the levels of the IgG1, IgG2a and IgG2b in these positive control serums were measured in ng mL<sup>-1</sup> with total IgG1, IgG2a and IgG2b ELISA kits (Chondrex, Redmond, WA, USA). IgA quantification in the BAL fluid (BALF) was normalised per protein concentration as measured by BCA assay.

### Haemagglutinin yeast surface display (HA-YSD) epitope mapping

To map HA-specific antibodies to the length of the HA protein, we used a fragmented YSD HA library to bind immune mouse serum. A H3N2-1968 HA-YSD library was constructed as previously described.<sup>23</sup> Yeast surface expressing HA (length 100–500 bp) was constructed from random digestion of the HA fragments derived from the H3N2-1968 HA gene. Antibody-positive yeast were isolated by cell sorting (FACSaria I cell sorter, BD Bioscience, Franklin Lakes, NJ, USA) and grown overnight in selective medium. Plasmids were then extracted by Zymoprep yeast plasmid miniprep II kit (Zymo Research, Irvine, CA, USA) and transformed in *E. coli*. A representative of 120 individual clones was picked for sequencing analysis from A-eIV and H3N2-1968 convalescent serums. Sequences were then aligned to the original H3N2-1968 HA sequence.

### Virus neutralisation antibody (VNA) assays

Influenza-specific VNA was measured by standard microneutralisation (MN) assay. Immune serum was pre-treated with receptor-destroying enzyme (RDE) (Denka Seiken, Tokyo, Japan). Twofold serial dilutions from 1:5 to 1:3200 of mouse serum samples were prepared in virus medium (MEM, 1% Pen/Strep). An equal volume of 200TCID<sub>50</sub> 35 µL<sup>-1</sup> of influenza viruses, H1N1-2009, H3N2-2013, H3N2-1968 and H7N7 (low pathogenic avian influenza (LPAI); A/Northern Shoveller/MPF518/2008), was added to the sample dilutions (final serum dilution 1:10 to 1:1280) and incubated for 2 h. Then, 35 µL of antibody/virus was added to MDCK cells and incubated for 72 h. Standard haemagglutination assay and visualisation of cytopathic effect were used to measure virus inhibition.

### Immune cell profiling by flow cytometry

To quantify and profile adaptive immune cells recruited during vaccination and infection, we used multi-parameter flow cytometry using a panel of phenotypic immune cell markers. Single-cell suspensions were prepared from lymph nodes (mLN from infection and iLN from vaccination), spleen or BAL, as previously described.<sup>47</sup> Cells were then stained with Zombie live/dead (Biolegend, San Diego, CA, USA), FcR-blocked (anti-CD16/CD32; BD Bioscience), and stained with one of the three cocktails containing a panel of monoclonal antibodies (all from Biolegend unless otherwise indicated) for 30 min on ice in FACS buffer (PBS, 1% FBS and 0.5% NaN<sub>3</sub>). Cocktail 1 for GC B and Tfh cells contains the following: anti-mouse CD3-BV605, B220-PECy7, CD4-APCCy7, PNA-FITC (Vector Labs, Burlingame, CA, USA), CXCR5-PerCPy5.5, CD38-PE and intracellular Bcl6-PEDazzle594. For intracellular stains, cells were fixed in fixation/permeabilisation buffer (eBioscience, San Diego, CA, USA) for 30 min on ice and stained with anti-Bcl6-PEDazzle594 in Perm/Wash buffer (BD Bioscience). GC B cells were defined as B220<sup>+</sup> CD3<sup>-</sup> PNA<sup>+</sup> CXCR5<sup>+</sup> CD38<sup>low</sup> Bcl6<sup>+</sup>, and Tfh cells were defined as CD3<sup>+</sup> B220<sup>-</sup> CD4<sup>+</sup> CXCR5<sup>+</sup> Bcl6<sup>+</sup>. Cocktail 2 for B memory cells contains the following: anti-mouse B220-PECy7, CD38-PE, IgD-APC, CD95 (Fas)-BV605, IgM-FITC and IgG-APCCy7. B memory cells were defined as B220<sup>+</sup> CD38<sup>+/−</sup> Fas<sup>low</sup> IgD<sup>-</sup> IgM<sup>+</sup> or IgG<sup>+</sup>. Cocktail 3 for influenza-specific B cells contains the following: B220-PECy7 and rHA-probe-SA-APC (H3N2 A/Singapore/INFIMH-16-0019/2016) (from Christopher Mok, HKU and Nicholas Wu, Scripps). Cells were finally fixed with 100 µL<sup>-1</sup> of 4% PFA for 20 min on ice. Samples were acquired by flow cytometry on a FACS Attune (Invitrogen) and analysed with FlowJo software.

### Inflammation and damage by total protein in BAL and cytokine bead array (CBA)

Bronchoalveolar lavage samples were harvested in total 3 mL MEM, and cells were pelleted for immunological staining and supernatant for measuring total protein and cytokine concentrations. Total protein was measured on BAL fluids with the BCA Protein Assay Kit (Thermo Fisher Scientific, Waltham, MA, USA) with a twofold standard range using

BSA, according to the manufacturer's instructions. Optical densities were read on a plate reader at 562 nm. For the measurement of cytokines in the BAL, a custom 12-cytokine LegendPlex (Biolegend) panel included the following: IFN- $\gamma$ , IL-6, TNF- $\alpha$ , IL-9, IL-2, IL-21, IL-4, IL-5, IL-10, CCL2, CCL3 and CCL4. Standard wells were run in duplicates from individual lung homogenate supernatants ( $n = 4$  per group). Samples were acquired by flow cytometry on a FACS Attune (Invitrogen) and analysed with LegendPlex software (Biolegend) as per the manufacturer's instructions.

### Intracellular cytokine staining (ICS) of influenza-specific T cells

To quantify influenza-specific T cells producing polyfunctional antiviral cytokines, single-cell suspensions of BAL and mLN were infected with a MOI of 4 of H3N2-1968 virus plus  $10 \text{ U mL}^{-1}$  rhIL-2 (Roche, Basel, Switzerland), purified CD49d and CD28 (Biolegend) for 6 h before the addition of BFA Golgi-Plug, and then incubated for further 12 h at 37°C. At 18 h post-infection, cells were washed with FACS buffer, stained with anti-mouse CD8-PerCPy5.5 and CD4-APCCy7 (Biolegend) for 30 min on ice, and then fixed (BD Cytoperm/Cytofix) for 20 min on ice. Cells were then stained intracellularly with anti-mouse IFN- $\gamma$ -FITC, TNF- $\alpha$ -APC and IL-2-PE for 30 min on ice. Samples were acquired by flow cytometry on a FACS Attune (Invitrogen) and analysed with FlowJo software. Total cytokine production was calculated by subtracting background fluorescence using the no-virus controls.

### In vitro FACS protection assay

To determine whether vaccination elicited antibodies that reduced or increased infection, we used an *in vitro* FACS protection assay. Raji cells ( $1 \times 10^5$  per well) were infected with a MOI of 4 of H3N2-2013, H3N2-1968 or H7N7 virus in the presence of mice or human sera (neat, 9  $\mu\text{L}$ ) for 16 h in MEM (1% Pen/Strep). Cells were then fixed (BD Cytofix/Cytoperm) and stained with anti-NP-FITC (Abcam, Cambridge, UK) in BD permeabilisation buffer for 30 min on ice. Samples were acquired by flow cytometry on a FACS Attune (Invitrogen) and analysed with FlowJo software. H3N2-1968-recovered mouse serum and naïve mouse serum were used as positive and negative controls, respectively. The % NP<sup>+</sup> cells were normalised to naïve mouse serum, and a ratio above 1 indicates increased infection and below 1 indicates reduced infection. For human samples, the fold change versus day 0 responses was calculated.

### Quantitative RT-PCR (qRT-PCR) of genes associated with SHM and CSR

Quantitative RT-PCR was performed for the murine *Aicda* (AID), *Bcl6*, *Pax5*, *IRF4* and *C-Myc* genes on B cells isolated from mouse iLNs using magnetic B-cell isolation kit (Miltenyi Biotec, Bergisch Gladbach, Germany). RNA was extracted using RNeasy mini kit (Qiagen, Hilden, Germany), and a reverse transcription using oligo-dT primers with PrimeScript RT reagent kit (TAKARA, Kusatsu, Japan) was performed. The cDNA was then used in the qPCR (Fast SYBR Green, ABI, St. Louis, MO, USA)

with gene-specific primers (Supplementary table 2), then run and analysed in the LightCycler 480 (Roche). To quantify the gene expression, relative qRT-PCR was performed by normalising the data to the GAPDH gene expression.

### Statistical analysis

Results represent the individual result, mean  $\pm$  SEM of 4–6 mice per group, unless indicated otherwise. Statistical significance was compared between the S-IV group versus PBS (indicated as  $^{\wedge}P < 0.05$ ,  $^{\wedge\wedge}P < 0.01$  and  $^{\wedge\wedge\wedge}P < 0.0001$ ) and eIV versus S-IV group (indicated as  $*P < 0.05$ ,  $**P < 0.01$  and  $***P < 0.0001$ ) using a one-way ANOVA (unless indicated) on GraphPad Prism software v8. When only two groups were present in an experiment, a Student's *t*-test was performed on GraphPad Prism software v8. The Pearson correlations were also performed on GraphPad Prism software v8. To compare the distributions of the AUC IgM and IgG between groups, a binomial test of the percentage values was performed for each observed value of eIV groups versus the S-IV group on GraphPad Prism software v8 with a Wilson/Brown calculation of the confidence intervals.

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### CONFLICT OF INTEREST

BJC has received honoraria from Sanofi and Roche for advisory committees. The authors report no other potential competing interests.

### AUTHOR CONTRIBUTIONS

NK and SAV designed the study and wrote the first draft of the manuscript. NK, AH, AL, CC, SAV and AWC performed the experiments. NK, SAV, BJC, NHL, VJF and LLMP provided analysis and interpretation of experiments. All



authors critically revised the paper. VJF and BJC were the consulting statisticians.

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## Supporting Information

Additional supporting information may be found online in the Supporting Information section at the end of the article.



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