

1 **Recent Advances in Mitigating Membrane Biofouling Using Carbon-**
2 **based Materials**

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23 **Abstract**

24 Biofouling is the Achilles Heel of membrane processes. The accumulation of organic
25 foulants and growth of microorganisms on the membrane surface reduce the
26 permeability, shorten the membrane life, and increase the energy consumption.
27 Advancements in novel carbon-based materials (CBMs) present significant
28 opportunities in mitigating biofouling of membrane processes. This article provides a
29 comprehensive review of the recent progress in the application of CBMs in
30 antibiofouling membrane. It starts with a detailed summary of the different
31 antibiofouling mechanisms of CBM-containing membrane systems. Next,
32 developments in membrane modification using CBMs, especially carbon nanotubes and
33 graphene family materials, are critically reviewed. Further, the antibiofouling potential
34 of next-generation carbon-based membranes is surveyed. Finally, the current problems
35 and future opportunities of applying CBMs for antibiofouling membranes are discussed.

36

37 *Keywords:* Carbon-based materials; Carbon nanotube; Graphene, Biochar; Biofilm;

38 Biofouling

39

40

41 **1. Introduction**

42 Membrane technology is a promising alternative to address water scarcity, one of
43 the most serious challenges of our time [1]. It offers many advantages over the
44 conventional treatment techniques, such as reliable water quality, high flexibility,
45 reduced usage of chemical additives, and relatively low energy consumption in the
46 overall processes [2]. Their easy operation and modular nature have enabled a number
47 of commercially successful membrane-based water/wastewater treatment processes,
48 including membrane bioreactors (MBRs), reverse osmosis (RO), nanofiltration (NF),
49 ultrafiltration (UF), and microfiltration (MF) [3–6]. Many emerging processes (e.g.,
50 forward osmosis and membrane distillation) also show promising niche applications
51 (e.g., for zero liquid discharge) [7,8].

52 Despite their advantages, biofouling is a main obstacle hampering the widespread
53 use of membrane technology. It is caused by waterborne microorganisms and
54 dissolved/particulate organic substances that are retained on the surface of or inside the
55 membrane. Biofouling not only reduces the membrane flux and compromises the
56 permeate quality, but also results in additional energy consumption and shortens the
57 membrane life [9–12]. Numerous strategies have been attempted to address this
58 problem through pretreatment of feed water [13,14], improvement of process design
59 [15], optimization of operational conditions [16,17], membrane cleaning [18,19], and
60 development of antibiofouling membranes [20,21].

61 Carbon based materials (CBMs), such as biochar, carbon nanotubes (CNTs),
62 graphene, mesoporous carbon nanoparticles, and carbon quantum dots, show great

63 promise in developing novel and high-performance membranes [22–25]. These CBMs
64 have many advantageous properties, including excellent separation properties, large
65 surface areas, high mechanical strength, unique electrical and thermal conductivity
66 properties, and superior antibacterial properties (Table 1). Some of these novel CBMs
67 have enabled the fabrication of membranes of improved separation properties [25]. For
68 example, single-layer graphene shows remarkably high water permeability (several
69 orders of magnitude higher than conventional RO membranes) by reducing the
70 membrane thickness to a monoatomic level [26]. Graphene family materials also
71 exhibit exceptional antibacterial and antifouling effects originating from their
72 electrostatic repulsion properties, hydrophilicity, and capability of inducing physical or
73 oxidative damages to cell membrane or metabolism systems [27,28,29]. In this context,
74 the antibiofouling ability of CBMs has attracted great interest in the field of membrane
75 technology.

76 There are several recent critical reviews devoted to the advances in the application
77 of CBMs in membrane fabrication [30–32]. These reviews mainly focused on the usage
78 of CBMs in tuning membrane structure, physiochemical, transport and separation
79 properties. Although CBMs demonstrated remarkable antifouling capacity, there has
80 been no comprehensive review that addresses their fouling control mechanisms and
81 future opportunities in membrane technology. Therefore, this article aims to provide the
82 first review to summarize the state-of-the-art research results on the applications of
83 CBMs in mitigating membrane biofouling. A summary of different biofouling control
84 mechanisms is provided. The recent advances in membrane modification using CBMs

85 and the antibiofouling properties of next-generation graphene-based membranes are
86 surveyed. Finally, current challenges and future opportunities are highlighted.

87

88 **2. Fundamentals of biofouling control using CBMs**

89 Membrane biofouling is triggered by the synergistic effects of microbial cells and
90 abiotic organic foulants, such as extracellular polymeric substances (EPS) and natural
91 organic matter. Because of the unique and tunable structures and physicochemical
92 properties of CBMs, they open a door to develop membranes with better anti-adhesion
93 and bactericidal properties. Novel carbon-based nanocomposites have also been
94 developed to control biofilm formation by manipulation of bacterial signaling system.

95

96 *2.1 Improvement of anti-adhesion properties*

97 Adhesion of polysaccharides, proteins, and microbial cells on a membrane surface
98 depends on their interactions (e.g., van der Waals and electrostatic forces and acid-base
99 interactions) with the membrane surface [33]. These interactions are affected by the
100 physicochemical properties of the membrane surface, such as roughness, functional
101 groups, surface charge, and hydrophobicity [34].

102 In general, membranes with decreased surface roughness, reduced surface charge,
103 and higher hydrophilicity tend to experience less biofouling. CBMs can play an
104 important role in creating anti-adhesion membranes. Incorporation of CBMs such as
105 GO as nanofillers in polymeric membranes induces nucleation and growth of polymer,
106 which resulted in a smoother membrane surface [35,36]. During interfacial

107 polymerization, GO can retard the diffusion of monomer solution into organic solvent,
108 which reduces the ridge formation on the membrane surface [37]. For example, the
109 mean roughness of a thin-film composite (TFC) membrane was decreased about 80%
110 after the addition of 800 ppm GO in the monomer aqueous solution [38]. When GO was
111 grafted on membrane surfaces, they occupied the valleys on the surface, which flattened
112 the surface [39,40]. Although pristine CBMs, such as CNTs and graphene, are
113 hydrophobic (Table 1), hydrophilic functional groups can be introduced to improve the
114 compatibility between CBMs and polymers. The hydrophilic CBMs can also induce the
115 formation of a hydrated layer, which exerts steric exclusion effects to inhibit the
116 adsorption of organics [41].

117 As bacterial cells are prone to adhere on surfaces with a water contact angle of 40–
118 70° [42], superhydrophobic membranes with a water contact angle greater than 150°
119 exhibit effective antibiofouling properties. The superhydrophobic surface minimizes
120 the membrane-water contact area, which largely decreases the probability of organic
121 foulants adhering to the membrane surface [43]. Superhydrophobic surfaces can be
122 fabricated by the deposition of multiwall CNTs (MWCNTs) [44] or by growing a
123 network of CNTs *in situ* using chemical vapor deposition [45]. The silane-treated rGO
124 also showed a contact angle of 157° which can be applied as superhydrophobic coating
125 [46].

126

127 *2.2 Antimicrobial effects*

128 Many nano-size CBMs, such as CNTs and graphene, have exhibited superior

129 antimicrobial properties (Figure 1 & Table 1). Membranes functionalized with these
130 CBMs similarly exhibit excellent antimicrobial effects. These antimicrobial effects can
131 be classified according to their different mechanisms into direct physical damage,
132 oxidative stress, and reactive oxygen species (ROS)-independent approaches.

133 The sharp edges and nanostructures of nano-size CBMs can pierce microbial
134 membranes, causing direct physical damage (Figure 1C). Recent studies found that GO
135 can extract phospholipids from the bacterial membranes onto its own surface [47]. The
136 loss of bacterial membrane integrity results in the release of vital intracellular
137 substances, and eventually causes microbial deactivation. The induction of oxidative
138 stress by nano-size CBMs is another crucial antimicrobial mechanism. Some
139 nanomaterials such as CNT, GO, and fullerene can promote the generation of ROS via
140 the reduction of adsorbed O₂ by cellular enzymes or metabolites (Figure 1D).
141 Photosensitizing materials such as fullerene C60 can generate ROS under ultraviolet
142 irradiation [48]. Excessive levels of ROS lead to the oxidation of fatty acids in the cell
143 membrane, and disrupt the membrane integrity and vital cellular processes.

144 Antimicrobial effects can also be induced by ROS-independent approaches.
145 Carbon-based nanomaterials were found to hinder cell growth by disrupting vital
146 cellular functions and metabolic processes. For example, CNTs can inhibit the
147 pyoverdine production of *Pseudomonas aeruginosa* PAO1, which is essential to its
148 survival under iron-limited conditions [49]. The GO sheets with a large lateral size were
149 shown to prevent bacterial nutrient uptake when wrapped around bacterial cells [27]
150 (Figure 1E). Moreover, the decoration of carbon nanomaterial with other bactericidal

151 substances, such as Ag nanoparticles, can further improve their antimicrobial activity
152 [50].

153

154 *2.3 Biofilm signaling disruption*

155 With the tunable carbon backbone, novel carbon nanocomposites have been
156 developed to manipulate bacterial biofilm signaling to mitigate membrane biofouling.
157 Quorum sensing (QS) is a crucial signaling system for biofilm formation. The
158 accumulation of QS signals upregulates the expression of biofilm formation genes and
159 promotes the development of mature biofilms [51]. Therefore, quenching QS signals is
160 a favorable approach to inhibit biofilm formation on the membranes. When AHL-
161 acylase was immobilized on GO, the modified membrane was able to hydrolyze QS
162 signals, which subsequently reduced EPS production and biofilm development [36].

163

164 *2.4 Other mechanisms*

165 CBMs also facilitate other antibiofouling mechanisms. For example, embedding
166 conductive CBMs enables electrically assisted fouling mitigation. Through like-charge
167 electrostatic repulsion, gram-negative bacterial cells are repelled from negatively
168 charged membranes [52]. Moreover, the electrochemical reduction of water generates
169 hydrogen bubbles at the membrane surface to float the foulants (Figure 2A) [53,54].
170 The fouled organics on membrane surface can also be decomposed by electrochemical
171 oxidation (Figure 2B) [55,56]. To prevent foulants from clogging the water transport
172 channel, electrophoretic pumping cationic complexes can act as molecular brushes for

173 membrane cleaning (Figure 2C) [57].

174

175 **3. Dosing CBMs for antibiofouling**

176 Dosing of powdered activated carbon (PAC) is a traditional method for wastewater
177 treatment and membrane biofouling control. Due to its large adsorptive surface,
178 activated carbon can adsorb organics and microbes from the bulk solution. When PAC
179 was applied in membrane bioreactors as an adsorbent, the integrity of sludge granules
180 was increased by enhanced agglomeration with PAC and less organic foulants were
181 available in the solution [58]. Therefore, dosing of PAC is considered an indirect
182 approach to improve the filterability of wastewater. For instance, the addition of PAC
183 in membrane bioreactor (MBR) reduced the total fouling resistance (R_t) by 15.9%
184 compared to that of the control group without PAC [59]. A low PAC dosage (0.75 g/L)
185 was found to result in a better fouling resistance than a high dosage (1.5 g/L). At a low
186 PAC dosage, the EPS in the bulk liquid and fouled on the membrane were 84.3% and
187 90.0% of that in the reactor with high dosage. Consequently, the MBR with a lower
188 dosage exhibited less irreversible fouling and a higher flux recovery rate. Further study
189 found the lower fouling propensity was caused by the higher integrity of the sludge. At
190 a low dosage of PAC, 84.3% less polysaccharides were released when exposed to
191 additional shear [58].

192 As activated carbon is an expensive and non-regenerable material, its high
193 operational costs hinder its practical application. In recent years, biochar has emerged
194 as a low-cost carbon-rich by-product of pyrolysis that can be designed/engineered for

195 multiple applications [60,61]. The π -electron rich sites and dense polar functional
196 groups of biochar endow it with a higher affinity towards humic acid than PAC [62],
197 which can significantly alleviate the irreversible fouling of membranes (Figure 3). It
198 has been reported that dosing biochar in MBR can achieve fouling mitigation effects
199 comparable to those of activated carbon [63]. In comparison, the cost of producing
200 biochar is less than one-tenth that of activated carbon [64]. The key physicochemical
201 properties of biochar can be controlled by altering the pyrolysis conditions. For example,
202 biochar generated under pure nitrogen was found to have a higher adsorption capacity
203 than biochar prepared under low oxygen condition and PAC [62]. When applied in
204 humic acid ultrafiltration, 12.9% less flux decline and 4.1% higher rejection were
205 achieved by biochar compared to the use of PAC. Although CBMs dosing increases the
206 operating cost, it can be offset by the decreased cost for membrane cleaning and the
207 application of low-cost alternatives to PAC. Moreover, CBMs dosing can be beneficial
208 to the wastewater treatment process, which provide substrata for biofilm growth and
209 improve the sludge settleability and dewaterability .

210

211 **4. Application of CBMs in membrane modification**

212 CBMs with different antibiofouling mechanisms can be incorporated into
213 membranes to improve their fouling resistance. CNTs and graphene are the most widely
214 applied CBMs in membrane fabrication. They possess excellent thermal and
215 mechanical properties and superior antimicrobial activities, which provide a non-
216 leaching and non-depleting alternative to metal nanoparticles. Recent advances in their

217 applications in membrane modification are highlighted.

218

219 *4.1 Carbon nanotubes*

220 Single-wall CNTs (SWCNTs) are graphite sheets with a hollow cylindrical
221 structure, while multi-wall CNTs (MWCNTs) consist of multiple rolled layers. CNTs
222 demonstrate tremendous potential in water technology thanks to their excellent
223 electrical, mechanical, and antifouling properties. Moreover, the hydrophobic and
224 atomically smooth interior wall facilitates nearly frictionless water flow through the
225 CNT core [65]. Based on the orientation and location of the CNTs in membrane, CNT-
226 containing membranes can be further classified into vertically aligned, mixed in
227 membranes, and coated on membrane surface (Figure 4).

228 Vertically-aligned (VA) CNT membranes can be prepared by casting and
229 deposition (Figure 4A) (Baek et al., 2014; Madaeni et al., 2013). Due to the fast water
230 transport through the CNTs, the water permeability of the modified membranes was
231 significantly enhanced (Table 2) [66]. The flow velocity through the CNT core was at
232 least 1000 times faster than conventional no-slip flow (Kalra et al., 2003). Due to the
233 hydrophobic properties of pristine CNTs, the surfaces of VACNT membranes become
234 more hydrophobic and rougher [44,66]. When filtering a bacterial suspension, the
235 number of bacteria attached on the surface of the VACNT membrane was 2 log less
236 than that on the control membrane without VACNT, which corresponds to 15% less
237 reduction in the permeate flux (Baek et al., 2014). By modifying the pore size and the
238 functional groups at the ends of the CNTs, the perm selectivity of the VACNT can be

239 further improved [67]. When zwitterions were functionalized onto the ends of the CNTs,
240 strong electrostatic interactions and a stable hydration layer were generated on the
241 membrane surface, which resulted in less protein fouling and a higher recovery rate in
242 the reverse osmosis (RO) membrane (Figure 5) [67]. Moreover, a packed VACNT array
243 without a polymeric matrix can be used directly as a membrane. The densified CNT
244 wall was found to enhance water permeability by 2,366% compared with other CNT
245 membranes [68]. Both the VACNT wall and VACNT membrane exhibited good
246 antifouling potential via decreased attachment of bacteria and inhibition of biofilm
247 formation on the membrane surface [66,68]. An assessment of the cell viability of the
248 membranes revealed that their antibiofouling effects were attributed to growth
249 inhibition rather than deactivation.

250 Due to the hydrophobic properties and strong π - π interactions between CNTs,
251 pristine CNTs have a low dispersibility in solution and poor interfacial interaction with
252 polymers. When pristine CNTs are used as a membrane additive, incompatibility
253 between the CNTs and the polymer generates nanocorridors that compromise the solute
254 rejection [69]. Therefore, chemical treatment is commonly applied to introduce
255 hydrophilic moieties such as carboxyl, hydroxyl, and amino groups on the CNTs surface
256 to improve their dispersibility and compatibility (Figure 5). CNT mixed-matrix (MM)
257 membranes can then be prepared by introducing the modified CNTs into the active layer
258 or support layer of membrane via interfacial polymerization or phase inversion (Figure
259 4B). The incorporation of CNTs into either the support or active layer enhances the
260 water permeability and fouling resistance of the resulting membrane by preventing

261 clogging in the skin layer and substrate pores [69,70]. The addition of CNTs to the
262 active layer provides more effective biofouling reduction, in which the degree of the
263 total flux loss (R_t) caused by protein fouling can be reduced by 78.7% . The bacterial
264 growth inhibition is directly correlated with the amount of CNTs in the active layer.
265 When the weight percentage of MWCNTs in the active layer was increased from 0.02
266 to 1%, the *Escherichia coli* growth inhibition rate increased from 55% to 80% [71].

267 Inclusion of carboxyl- and amine-surface modified CNTs into a membrane
268 enhanced surface hydrophilicity and reduced roughness, thus increasing pure water flux
269 and improving membrane fouling resistance against proteins and polysaccharides
270 [70,72–74]. The smooth surface of CNT MM membranes results from the increased
271 viscosity of the casting solution, which hinders diffusion into the organic phase during
272 dissolution [71,74]. Grafting carboxyl and hydroxyl groups onto the CNT surface also
273 enables further functionalization with other functional groups. For example,
274 dodecylamine and hyperbranched poly(amine-ester) groups were functionalized on
275 carboxylated MWCNTs, which further enhanced the protein fouling resistance and
276 hydrophilicity of the resulting membranes [41,75]. Sulfonated CNTs produce a
277 negatively charged surface for electrostatic repulsion [76,77]. In order to further
278 enhance the antibacterial effect, biocidal groups have been grafted onto CNTs, such as
279 (3-chloro-2-hydroxypropyl)-(5,5-dimethylhydantoinyl-1-ylmethyl)-
280 dimethylammonium chloride (CDDAC), which inactivated more than 90% of *E. coli*
281 and *Staphylococcus aureus* in 24 hours [78].

282 CNTs can also be immobilized onto membrane surfaces by direct deposition or

283 interfacial polymerization. Coating of the membrane surface with CNTs significantly
284 promoted its antibacterial effect. After 24 h of cross-flow filtration, little biofilm was
285 formed on a CNT-coated membrane surface, and more than 99% of the bacteria in the
286 feed were inactivated [79]. Antibacterial and antifouling activities were further
287 improved by doping with AgNPs (Figure 5) [50,80]. Deposition of the CNT-AgNP
288 composites on hollow fiber membrane was found to significantly decrease R_f by 98% .

289 Due to the extraordinary electrical conductivity of CNTs, the resistivity of surface-
290 modified membranes was found to be reduced by nearly an order of magnitude [55].
291 Therefore, electro-kinetic systems can be introduced to improve the antifouling
292 performance. By combining an electric field and a negative charge, organic foulants
293 were repelled by the negatively charged surface and electrophoresis to achieve superior
294 antifouling resistance [81,82]. Moreover, microbubbles were generated by periodic
295 electrolysis, enabling *in situ* cleaning of fouled membranes [83]. By contrast, when a
296 positive charge was applied, fouling was mitigated by electro-oxidation, in which the
297 organics adsorbed on the membrane were decomposed by electrochemical reaction
298 [55,56]. The removal of organic foulants such as humic acids was found to be a function
299 of the applied potential. At +1.5V, the permeate flux and removal efficiency were about
300 1.6 and 3.0-fold higher than those of the corresponding uncharged membrane [56].
301 Electrochemical assistance also allowed a positively charged CNT membrane to
302 inactivate bacteria and even bacteriophages attached to the membrane surface
303 [52,84,85]. Under a positive applied potential, the rate of flux declined during bacterial
304 filtration was three times lower than that of the control with no applied voltage [52].

305 Full recovery of the initial flux was achieved after a short flushing.

306

307 *4.2 Graphene family materials*

308 Graphene is an atomically thin sheet of sp^2 -bonded carbon atoms in a hexagonal
309 arrangement. Chemically-modified graphene derivatives, such as GO and reduced
310 graphene oxide (rGO) are referred to graphene family materials (GFMs) [26]. Due to
311 their low-cost mass production, chemical inertness, high tensile strength, and
312 antimicrobial properties, GFMs have generated tremendous interest in the field of
313 membrane modification. Similar to CNTs, GFMs can be incorporated into membranes
314 via interfacial polymerization and phase inversion. Alternatively, they can be
315 immobilized on the membrane surface by deposition, covalent functionalization, and
316 layer-by-layer approaches. Recent investigations have exploited the alignability of GO
317 on membranes. In a magnetic field, GO nanosheets could be vertically aligned on the
318 membrane surface which maximized the edge exposure and antimicrobial activities
319 [86].

320 Due to the strongly hydrophobic nature of pristine graphene, GO is prepared by
321 oxidative modification, which introduces oxygen-containing moieties such as hydroxyl,
322 epoxy, carbonyl, and carboxyl groups [26]. These functional groups make the use of
323 modified graphene as a membrane additive more feasible (Table 3). A TFC membrane
324 embedded with GO exhibited an increased hydrophilicity, negative surface potential,
325 and decreased roughness, which resulted in greater than 95% reduction in the volume
326 of biofilm on the membrane [37,87]. These fouling mitigation properties are directly

327 correlated to the oxygen-containing groups. On the contrary, the use of rGO in a MM
328 membrane showed a detrimental effect on the antifouling properties [88]. With its
329 higher specific surface and greater amount of oxygen-containing functional groups, a
330 GO-blended PVDF membrane demonstrated 30% higher pure water permeability than
331 a comparable membrane containing oxidized MWCNTs [89]. The hydrophilic
332 membrane surface also contributed to a decrease in R_t by 6% and an enhancement of
333 flux recovery rate by up to 5-fold (Table 3). However, a high loading of GO in a
334 membrane induced aggregation and produced flaws in the membrane [37,38]. This
335 negative effect can be alleviated by the synergistic interactions between CNTs and GO.
336 In such a hybrid system, CNT-bridged GO formed a 3-D architecture that prevented
337 aggregation and strengthened the interactions between the polymer and the
338 nanomaterials [90,91].

339 Anchoring of other functional groups can further enhance antifouling performance
340 (Figure 6). For example, due to the stronger hydrogen-bonding forces and electrostatic
341 repulsion of the sulfonic groups, the use of sulfonated GO increased the flux recovery
342 ratio of a GO MM membrane by 18.3% [57]. Grafted antimicrobial groups such as
343 tannic acid, Co_3O_4 , and AgNPs improved the bacterial inactivation rate of GO
344 membranes [92–94]. The loading of GO- Co_3O_4 composites was found to decrease R_t
345 caused by sludge fouling by 20.8% and increase the flux recovery rate by 45.4%. In
346 addition to antimicrobial activity, antibiofouling can be achieved by influencing the
347 biofilm signaling system as discussed before. When acylase was decorated onto the
348 surface of GO sheets, the nanohybrid membrane was capable of hydrolyzing biofilm

349 signaling molecules [36] (Figure 6). Consequently, although acylase had negligible
350 effects on organic fouling mitigation, the formation of biofilm on the membrane was
351 reduced by 83.9%.

352 The unique attributes of GFMs render them a promising material to tailor the
353 surface features of membranes. GFMs are anchored onto the membrane surface by
354 covalent bonding and layer-by-layer (LbL) assembly. After the GFMs are deployed on
355 the membrane surface, the hydrophobic area of the basal plane of GO can cause
356 enhanced organic adsorption, without affecting the water flux around the edges of the
357 GO [95]. Moreover, GO functionalization can shield the carboxyl groups on the original
358 membrane surface and maintain the volume charge density [96]. Therefore, surface
359 coating with GO improves the antimicrobial effect without affecting the water
360 permeability, in which 50% reduction in R_t was achieved [97–99]. Comparing the two
361 mainstream surface coating approaches for TFC membranes, the density of GO grafted
362 on surfaces by covalent crosslinking is higher than that of LbL membranes, which
363 improves the surface properties and antibiofouling effects [40]. In pressure-retarded
364 osmosis (PRO) membranes, the LbL approach enables the formation of thin films on
365 both sides of the membrane support, which prevents irreversible fouling inside the
366 porous support [95].

367 Due to the inertness of GO, the chemical resistance of GO-containing membranes
368 is increased. The coating layer can protect RO membranes against chlorine attack by
369 preventing the active chlorine species from diffusing toward the selective layers [100].
370 Such membranes showed higher durability under frequent oxidative cleaning [101].

371 The physicochemical properties of GO also facilitate its use in photocatalytic
372 antibiofouling processes. An integrated GO-TiO₂ membrane showed 51% and 74%
373 higher protein photodegradation efficiency than membranes containing TiO₂ or GO
374 alone, respectively [102]. Photocatalysis can also be utilized for *in situ* synthesis of
375 AgNPs, which provides an alternative method to regenerate the antimicrobial properties
376 of the membrane [103].

377

378 *4.3 Other CBMs*

379 Other antimicrobial carbon-based nanomaterials such as fullerene, mesoporous
380 carbon nanoparticles (MCNs), and carbon quantum dots (CQDs) have also been
381 embedded into membranes to enhance their antifouling performance. The deposition of
382 the fullerene C₆₀ on the membrane was found to reduce bacterial attachment and inhibit
383 microbial respiration [104]. MCNs have a higher specific surface area than GO and
384 CNTs, which makes them a promising membrane filler. A membrane containing
385 carboxylated MCN exhibited an increase in hydrophilicity and surface roughness,
386 which resulted in 80% less protein adsorption and 90% less bacterial attachment [105].
387 The CQDs have emerged as a new class of carbon nanomaterials. Their antibacterial
388 effects are caused by physical damage and ROS induction [106]. When CQDs were
389 immobilized on a membrane surface via covalent linkage, the resulting membrane
390 demonstrated better antifouling and antibacterial properties than a similar GO
391 membrane [106]. After 12-h filtration of a bacterial suspension, the flux drop of the
392 CQD membrane was 24.3%, while the permeate decrease for the GO membrane was

393 65.7%.

394 Attempts have also been made to incorporate CBMs other than carbon
395 nanomaterials for membrane fabrication. Incorporation of PAC improved the
396 morphology and porosity of the composite microfiltration membrane, and the
397 selectivity of the membrane also increased at a low carbon loading [107]. Blending PAC
398 and hydrophilic PEG resulted in improved permeability, hydrophilicity, roughness, and
399 organic fouling resistance [108]. As an alternative to PAC, biochar has also been applied
400 in membrane modification in the latest studies. Comparable antibiofouling performance
401 was achieved, which indicated the great potential for biochar in practical applications
402 [109].

403 The membranes modified by CBMs have exhibited superior antibiofouling
404 properties which outperform the commercial membranes. Among different
405 modification approaches, the MM membrane can be readily realized for industrial scale
406 production. The fabrication procedure requires to load CBMs into the active or support
407 layer, which can be integrated into the existing production lines of polymeric
408 membranes. The low price of CBMs like MWCNTs makes MM membrane feasible to
409 scale up at a competitive cost. Compared with MM membrane, the processing of CBMs
410 into the primary rejection layer, for example VA CNT membranes, is often complicated
411 and time consuming. The incorporation of CBMs may also compromise the salt
412 rejection capability by forming defects or decreasing crosslinking density . Moreover,
413 the leachability of nano-size CBMs into aquatic environments or product water should
414 be considered, especially for the surface-modified membranes. The deposited CBMs

415 on membrane surfaces directly interact with the feed water which are more likely to
416 release into environments.

417

418 **5. Next-generation membranes fabricated using carbon-based materials**

419 The excellent separation properties of GFMs offer great opportunities to design
420 novel membrane processes. Nanoporous graphene (NPG) and graphene oxide
421 frameworks (GOFs) have been proposed as next-generation membranes for
422 desalination. To overcome the impermeability of pristine graphene, NPG is generated
423 by introducing nanometer pores via plasma etching or bombardment [110]. Due to its
424 well-defined nanopores and monoatomic thickness, NPG exhibits outstanding size
425 exclusion properties and water permeability. However, the scale-up of NPG membranes
426 remains challenging, and the antibiofouling properties of NPG are largely unexplored.
427 It has been recognized that rGO and graphite have been found to induce more intense
428 oxidative stress than GO and graphite oxide [111]. Functional groups can be introduced
429 to decorate the nanopores to mitigate fouling and prevent clogging. Therefore, NPG is
430 expected to have good biofouling resistance properties.

431 GOFs, which comprise stacked GO nanosheets, are another advanced alternative
432 to the existing desalination membranes. The GO laminates can be simply prepared by
433 filtration or LbL deposition to produce freestanding or substrate-supported GOF
434 membranes. Unlike membranes produced by surface modification, the stacked GO
435 itself serves as the selective layer in GOF membranes. Water molecules are transported
436 through the nanochannels between adjacent GO sheets, while the solute can be excluded.

437 To achieve the trade-off between permeability and membrane selectivity, GO-CNT
438 composite membranes were developed in which CNTs control the interlayer space
439 between graphene sheets . The pure water permeability was enhanced by more than 2
440 times, while high salt rejection ratio was maintained. The intercalation of CNT with GO
441 can also enhance the mechanical stability of GOF membranes against cross flow . The
442 ability of GO laminates to sustain long-term filtration of Kraft black liquor has been
443 demonstrated [112]. The accumulation of organic foulants on GO laminates was
444 dependent on their interlayer spacing, the hydrophilicity of the GO, and the chemical
445 properties of the cross-linker [113]. When more GO layers were introduced, the organic
446 fouling propensity of the membrane was reduced and its antimicrobial activity was
447 enhanced [114]. As stacked GO tends to disperse in water, rGO nanosheets are applied
448 to reduce the swelling of the GOF. To minimize fouling on the hydrophobic rGO surface,
449 hydrophilic coatings can be applied via hydrophilic adhesive polydopamine (pDA)
450 deposition [115]. Moreover, the chemistry and morphology of GO can be tuned to
451 optimize GO laminates for different purposes [116]. For example, the hydrophobicity
452 of GO laminates is controlled by adjusting their oxidation state via photoreduction. A
453 negative charge can be introduced to the nanochannels via *in situ* post-treatment with
454 free chlorine, which offers great potential to generate electrostatic repulsion [117].

455 Although the emerging GFMs have showed promising performance, most of these
456 applications are limited to small-scale devices. The poor mechanical stability of GFMs
457 under practical hydrodynamic flow condition is another major challenge in long-term
458 application. In order to resolve these issues, recent advancements have improved the

459 scalability of GOF membrane via spray coating and shear alignment . The resistance to
460 shear stress was enhanced by the additional interfacial adhesive layer between GO and
461 support layers. With exceptional resistance to chemical cleaning, the GOF membrane
462 can be a scalable alternative to commercial polymeric membranes in the desalination
463 industry.

464

465 **6. Conclusions and future prospects**

466 In recent years, substantial advances have been made in the application of various
467 CBMs for membrane antibiofouling. The use of CBMs shows great promise in
468 addressing the problem of biofouling and revolutionizing conventional membrane
469 processes. Dosing CBMs can effectively reduce membrane contamination by foulants
470 from the bulk liquid. Surface modification has shown promising antibiofouling activity
471 via tailoring the membrane surface roughness, hydrophilicity, electrostatic potential,
472 and antimicrobial properties. Based on synergistic effects of CBMs with other
473 nanomaterials, additional highly desirable characteristics and functionalities can be
474 introduced. Next-generation graphene-based membranes provide ultrafast water
475 transport and antibiofouling potential, which outperform current membrane processes.
476 However, many challenges still limit the practical application of these novel membrane
477 processes. In order to tackle the water crisis, further scientific and technical
478 contributions will be vital to fully explore the potential of membrane systems.

479 Firstly, industrialization of these cutting-edge membrane systems is still
480 problematic. For example, due to the synthetic complexity and difficulties in scalable

481 processing, large-scale production of NPG is still in its infancy [30]. The
482 commercialization of modified membranes using CBMs is relatively easier compared
483 to NPG and GOF [23]. Therefore, novel technologies and production techniques are
484 required to close the gap between research and industrial utilization. The application of
485 sustainable and cost-effective CBMs such as biochar is one alternative to promote
486 system sustainability and reduce production cost.

487 Secondly, the long-term durability and antifouling performance of these
488 membranes should be further evaluated. Over a long period of operation, microbes can
489 condition the membrane surface by EPS and dead cell components that facilitate
490 subsequent adhesion of microbes and eventually cause severe biofouling. Therefore,
491 more resources need to be invested to provide a comprehensive understanding of long-
492 term antibiofouling activity of next-generation membranes, whose superior rejection
493 and water permeability properties do not necessarily translate into long-term
494 antibiofouling activity.

495 Finally, appropriate antibiofouling characterization approaches should be adopted.
496 Measurement of the number of colony forming units (CFUs) is the mainstream method
497 to evaluate the antimicrobial effect of a membrane. However, these results can be
498 misleading due to cell aggregation under stress conditions [118]. Moreover,
499 nondestructive and real-time approaches should be applied to monitor the biofouling
500 processes of membrane. The most widely applied technique is to stain the bacteria
501 attached on the membrane after disassembly of the membrane module and observe them
502 under a confocal microscope. However, the resultant observation provides only a

503 snapshot of the fouling process, and artificial effects are induced during the examination.
504 To address this issue, nondestructive methods to assess biofouling have been developed
505 using optical coherence tomography and confocal microscope compatible microfluidics
506 [119,120].

507

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514

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1172

1173 Table 1. Material properties of CBMs used in antibiofouling membrane

	Activated carbon	Biochar	SWCNT	MWCNT	GO	Fullerene	Mesoporous carbon nanoparticle	Carbon quantum dot
Material feature	3D material	3D (nano)material	1D nanomaterial	1D nanomaterial	2D nanomaterial	0D nanomaterial	3D nanomaterial	0D nanomaterial
Water transport	NR ^a	NR	frictionless flow through CNT core (hydrophobic, smooth inner core)	frictionless flow through CNT core (higher water permeability than SWCNT) [121]	a) permeate through nanopore of GO sheet; b) permeate through interlayer space between adjacent GO nanosheets	No	NR	NR
Separation properties	NR	NR	Size exclusion (pore diameter: 0.6-100 nm) [122]	Size exclusion (pore diameter: 1-100 nm) [122]	Size exclusion: a) minimum pore size: 0.26 nm; b) 0.3-0.9 nm interspacing [123]	NR	NR	NR
Antibacterial	No	No	Physical damage, oxidative stress, metabolism disruption	Physical damage, oxidative stress, metabolism disruption (weaker than SWCNT)	Physical damage, ROS-dependent and ROS-independent oxidative stress	ROS production, metabolism disruption	No significant effects [105,124,125]	Physical damage, oxidative stress (GOQD ^b) (Zeng et al., 2012)
Adhesion affinity	High affinity to high molecular weight organics [126]	Similar with activated carbon	High affinity to various organic chemical [126]	High affinity to various organic chemical	Antiadhesion due to hydrophilicity and charge repulsion	Lower affinity and adsorption capability than CNT [127]	High affinity to various organic chemical [128]	Antiadhesion due to hydrophilic and charge repulsion (GOQD) [129]
Electrical conductivity (S/m)	30-80 [130]	10 ⁻⁶ -400 [131]	10 ⁶ [132]	10 ⁴ -10 ⁵ [133]	electrical insulation	10 ⁻⁶ – 10 ⁻¹² (Mokarova et al., 2001)	30-500 [134]	Electrical insulation (GOQD)
Market Price (\$/g)	0.3-2.1*10 ⁻³ [135]	0.1-4.0*10 ⁻³ [136,137]	25-280	0.6-25	125 -300	35-400	30-160	2000-7500

1174 ^aNR: no reference; ^bGOQD: graphene oxide quantum dot.

1175

1176 Table 2. Summary of the use of CNTs for membrane modification

Category ^a	Base polymer	Filtration process ^b	Carbon material ^c	Fabrication method	Pure water permeability (modified vs control)	Antibacterial effect	Antifouling effect ^d	Feed solution ^e	Reference
Vertically aligned	Epoxy	UF	SWCNT	Casting	231%	-	2 log less cell attachment	<i>P. aeruginosa</i> PAO1	[66]
	-	UF	SWCNT wall	Densification	2,466.7%	-	Qualitative	<i>P. aeruginosa</i> PAO1	[68]
	PVDF	NF	MWCNT	Deposition	-	-	FRR 82%	BSA	[44]
	Polyamide	RO	SWCNT-Z	Interfacial polymerization	-	-	FRR increased by 44.9%, R _t decreased 28.2%	BSA	[67]
MM in active and support layer	PSF	NF and FO	CNT	Interfacial polymerization	143.4%	-	FRR increased by 55.4% (NF), 15.6% (FO), R _t decreased by 39.8% (NF), 50% (FO)	HA	[138]
MM in support layer	PES	RO	MWCNT-COOH	Phase inversion	-	-	FRR increased by 6.64%, R _t increased by 6.48%	BSA	[70]
MM in active layer	Pluronic F-127 modified PVA	RO	MWCNT	Interfacial polymerization	-	80% growth inhibition	-	<i>E. coli</i>	[71]
	PES/PVDF	UF (HFM)	MWCNT-COOH	Phase inversion	463%	-	R _t decreased by 68.9%	BSA	[139]
	PSU	UF (HFM)	MWCNT-COOH	Phase inversion	196%	-	R _t decreased by 78.7%	BSA	[140]
	CA	FO	MWCNT-COOH	Phase inversion	-	-	R _t decreased by 23.5%	Alginate	[72]
	Polyamide	RO	MWCNT-COOH	Interfacial polymerization	-	-	FRR increased by 7%, R _t decreased by 10.8%	BSA	[141]
	PAES and APAES	UF	MWCNT-COOH	Phase inversion	150%	-	FRR increased by 17.7%, R _t decreased by 13.6%	Ovalbumin, lysozyme	[142]
	PVDF	RO	MWCNT-COOH	Interfacial polymerization	-	-	R _t decreased 58.9%	BSA	[73]
	PES	NF	MWCNT-NH ₂	Phase inversion	174.3%	-	FRR increased by 46.1%	BSA	[143]
	PES	UF	MWCNT-NH ₂	Phase inversion	148%	-	FRR increased by 70.4%, R _t decreased by 3.9%	BSA	[144]
	Polyamide	NF	MWCNT-NH ₂	Interfacial polymerization	-	-	R _t decreased by 44.6%	BSA	[145]
	PAES/S-PAES	UF	MWCNT-SO ₃ H	Phase inversion	176%	-	FRR increased by 14.3%, R _t decreased by 24%	BSA	[142]
	PES	NF	MWCNT-SO ₃ H	Interfacial polymerization	160%	-	FRR increased by 11.2%, R _t decreased 40%	BSA	[77]
	PVDF	UF	MWCNT-HPAE	Phase inversion	485%	-	FRR increased by 22.1%, R _t increased by 6.2%	BSA	[41]

	PSF	NF	MWCNT-DDA	Phase inversion	275%	-	FRR increased by 45.6%, R _t decreased by 43.1%	BSA	[75]
	PVDF	UF (HFM)	MWCNT-CDDAC	Phase inversion	211%	Sterilization ratio 92.7% (<i>E. coli</i>) and 95.2% (<i>S. aureus</i>)	FRR increased by 52.4% (BSA)	BSA, <i>E. coli</i> , and <i>S. aureus</i>	[78]
	PES	UF	MWCNT-PANI	Phase inversion	564.5%	-	FRR 100%, R _t 65%	HA	[146]
	-	MF	MWCNT-AgNP	Sintering	-	100% removed/killed	-	<i>E. coli</i> K12	[147]
	PVDF	Charged UF	CNT	Phase inversion	-	-	R _t decreased by 85.8%	SRFA	[82]
	PSF	Charged MF	MWCNT-COOH	Phase inversion	-	Inactivated 99.999%	-	<i>E. coli</i> , <i>S. aureus</i>	[148]
Surface modification	PVDF	MF	SWCNT, MWCNT	Deposition	-	-	TMP decreased by 7.03 bar	Prefiltered natural surface water	[149]
	PVDF	Charged MF	MWCNT	Phase inversion	110%	-	TMP decreased by 14.8%	SA, BSA, HA mixture	[81]
	PES	UF	MWCNT-PEG	Deposition	98%	-	R _t decreased by 60% (HA); 41.8% (BSA); 9.4% (SA)	HA, BSA, and SA solution	[150]
	PSF	RO	MWCNT-COOH	Deposition	-	Cell viability less than 1%	R _t decreased by 67%	<i>P. aeruginosa</i> PAO1	[79]
	PAN	UF (HFM)	MWCNT-AgNP	Deposition	96%	86.7% growth inhibition	R _t decreased by 98%, FRR 91.8%	<i>E. coli</i>	[50]
	Styrene, acrylic acid	MF	MWCNT-AgNP	Deposition	-	Inhibition zone diameter increased by 3.67 mm (<i>S. aureus</i>) and 2.53 mm (<i>E. coli</i>)	-	<i>S. aureus</i> and <i>E. coli</i>	[80]

PAN	Charged MF	CNT-COOH	Deposition	-	-	FRR 98%, R _t 15%	HA	[56]
Polyamine	Charged UF	MWCNT-COOH	Interfacial polymerization	-	-	FRR 92.9%	BSA	[55]
PVDF	Charged MF	MWCNT	Deposition	-	-	R _t 56.6%	Yeast	[83]
Polyamide	Charged NF	MWCNT-COOH	Interfacial polymerization	667%	-	FRR 100%; flux decline rate three times lower than control	<i>P. aeruginosa</i> PA01	[52]
PTFE	Charged MF	MWCNT	Deposition	-	7.4 log removal, 3.4 log inactivation	No fouling when filtering NOM	MS2 bacteriophage	[85]
Ceramic	Charged MF (HFM)	CNT-COOH	Deposition	-	Qualitative	R _t 12.1%	<i>E. coli</i>	[84]

1177 ^a MM: mixed matrix.

1178 ^b UF: ultrafiltration; RO: reverse osmosis; MF: microfiltration; FO: forward osmosis; HFM: hollow fiber membrane.

1179 ^c PANI: polyaniline; PEG: polyethylene glycol; Z: zwitterionic group; HPAAE: hyperbranched poly(amine-ester); DDA: dodecylamine; CDDAC: (3-chloro-2-hydroxypropyl)-(5,5-dimethylhydantoinyl-1-ylmethyl)-dimethylammonium chloride.

1181 ^d FRR: flux recovery rate; R_t: degree of the total flux loss caused by total fouling.

1182 ^e SRFA: Suwannee river fulvic acid.

1183

Table 3. Summary of the application of graphene for membrane modification

Category	Base polymer ^a	Filtration process	Carbon material ^b	Fabrication method	Pure water permeability (modified vs control)	Antibacterial effect	Antifouling effect	Feed	Reference
MM in active and support layer	PSF	RO	GO	Interfacial polymerization	-	-	Biovolumes decreased by 99%	<i>P. aeruginosa</i> PAO1	[37]
MM in active layer	PPSU	UF	GO	Phase inversion	143.7%	-	R _f decreased by 58.3%, FRR increased by 72.7%	BSA	[151]
	PSF	UF	GO	Interfacial polymerization	-	-	R _f decreased by 44.2%, FRR increased by 70%	BSA	[152]
	PES	NF	GO	Phase inversion	248.8%	-	R _f decreased by 31.6%, FRR increased by 158.6%	Protein	[153]
	PVDF	UF	GO, OMWCNTs	Phase inversion	351.7%	-	R _f decreased by 6.1%, FRR increased by 554%	BSA	[89]
	PVDF	UF	GO, OMWCNTs	Phase inversion	340%	-	R _f decreased by 9.2%, FRR increased by 177.7%	BSA	[89]
	PVDF	UF	GO	Phase inversion	196.4%	-	FRR increased by 23%	BSA	[154]
	PVDF	UF	GO	Phase inversion	321.8%	-	FRR increased by 26.6%	BSA	[155]
	PA	FO	GO	Interfacial polymerization	163.9%	-	FRR increased by 60%	SA	[38]
PA	UF	GO	Interfacial polymerization	180%	78.4% inactivation	-	<i>E. coli</i>	[156]	

PA	RO	GO	Interfacial polymerization	-	-	Biovolume decreased by 98%	<i>P. aeruginosa</i> PAO1	[87]
PSF	UF	GO	Phase inversion	151.4%	-	Biofilm thickness decreased by 43%	<i>P. aeruginosa</i> PAO1	[46]
PVDF	MF	GO	Phase inversion	151.5%	-	R _f decreased by 22%	Wastewater	[157]
PA	UF	GO	Interfacial polymerization	124.3%	-	R _f decreased by 71%	Natural water	[113]
PVDF (HFM)	UF	GO-SO ₃ H	Phase inversion	198.4%	-	R _f decreased by 36.6%, FRR increased by 8.7% (after five cycles)	HA	[101]
PVDF	UF	GO-SO ₃ H	Phase inversion	255.2%	-	R _f decreased by 7.9%, FRR increased by 76.3%	BSA	[57]
PVDF	UF	GO-APTS	Phase inversion	170.8%	-	R _f decreased by 63%, FRR increased by 203%	BSA	[158]
PSF	UF	GO-EDA	Phase inversion	303%	-	R _f decreased by 60.9%, FRR increased by 812.9%	BSA	[159]
PES	UF	GO-PAA	Phase inversion	244.1%	-	R _f decreased by 39.5% and 59.1%	Spent wash effluent	[160]
PES	NF	GO-PSBMA	Phase inversion	185.8%	-	R _f decreased by 68.6%, FRR increased by 68.8%	BSA	[161]
PA	RO	GO-TA	Interfacial polymerization	-	Viability decreased by 47.9%	-	<i>E. coli</i>	[93]
PES	NF	GO-ZIF8	Interfacial polymerization	152.6%	Inactivated 84.3%	-	<i>E. coli</i>	[162]

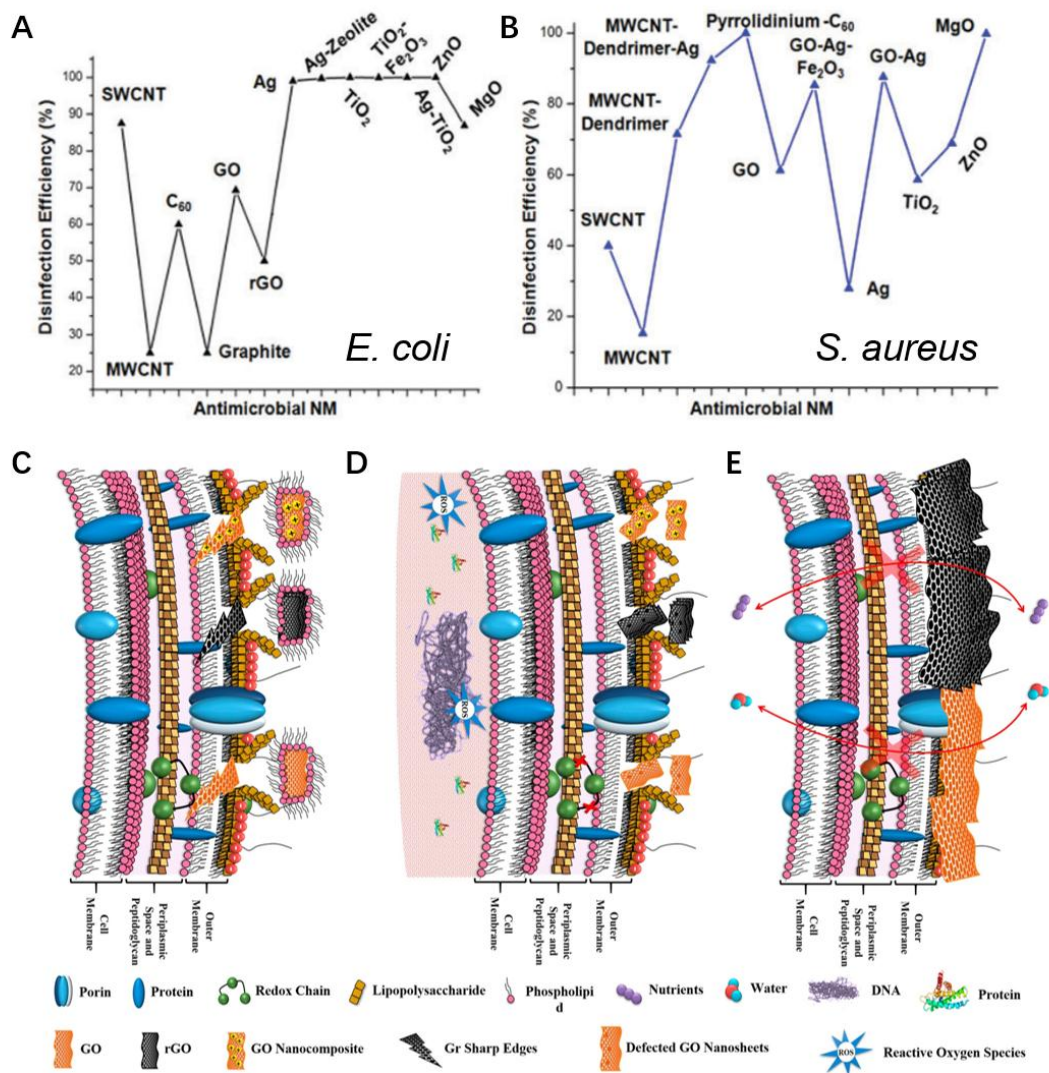
PSF	UF	GO-SiO ₂	Phase inversion	182.2%	-	R _i increased by 5.8%, FRR increased by 16.1%	BSA	[163]
PVDF	UF	GO-SiO ₂	Phase inversion	162.1%	-	R _i decreased by 37.3%, FRR increased by 23.2%	BSA	[35]
PVDF	UF	GO-SiO ₂	Phase inversion	252.9%	-	R _i decreased by 28.9%, FRR increased by 103.6%	BSA	[164]
PSF	NF	GO-ZnO	Phase inversion	574.2%	Qualitative	R _i decreased by 9.2%, FRR increased by 16.5% (HA)	HA, <i>E. coli</i>	[165]
PES	NF	GO-TiO ₂	Interfacial polymerization	100%	-	R _i decreased by 52.9%	BSA	[154]
PVDF	Photocatalytic UF	GO-TiO ₂	Phase inversion	308.5%	-	R _i decreased by 52.1%, FRR increased by 90.5%	BSA	[102]
PES	NF	rGO-TiO ₂	Phase inversion	194.8%	-	FRR increased by 28.7%	BSA	[166]
PSF	UF	GO-TiO ₂	Phase inversion	104.5%	-	R _i decreased by 37.5%, FRR increased by 29.6%	HA	[167]
PES	Photocatalytic UF	GO-TiO ₂ -AgNP	Filtration	4.2%	3 log less live cells attached on membrane	-	<i>E. coli</i> K12	[92]
PA	RO	rGO-TiO ₂	Interfacial polymerization	-	-	R _i decreased by 61.2%	BSA	[168]

	PES	UF	GO-Co ₃ O ₄	Phase inversion	344.1%	Inactivated 89.8% (<i>E. coli</i>)	R _t decreased by 20.8%, FRR increased by 45.4% (activated sludge)	<i>E. coli</i> , activated sludge	[94]
	PVDF	UF	GO-AC	Phase inversion	88.3%	Biovolume decreased by 83.9%	R _t decreased by 2.8%, FRR increased by 1.8% (BSA)	<i>P. aeruginosa</i> , BSA	[36]
Surface modification	PA	RO	GO/GO-NH ₂	Layer-by-layer assembly	-	-	R _t decreased by 55.9%	BSA	[100]
	PA	FO	GO-pDA	Covalent bonding	121.5%	ATP level decreased by 98.5%	R _t decreased by 37.1%	Surface water	[169]
	pDA/TMC	MF	GO	Layer-by-layer self-assembly	184.2%	-	R _t decreased by 47.2%	Alginate	[96]
	PA	PRO	GO	Layer-by-layer assembly	-	-	R _t decreased by 44.3%, FRR increased by 32.7%	Alginate	[95]
	PA	FO	GO	Covalent bonding	120%	ATP level decreased by 99.9%	R _t decreased by 18.5%, biofouling resistance increased by 33%	Surface water	[170]
	PA	FO	GO	Covalent bonding	-	Viability decreased by 32.6%, cell attachment decreased by 36%	R _t decreased by 50%	<i>P. aeruginosa</i>	[98]
	PE	MF	GO	Covalent bonding	-	99% bacterial inactivation	-	<i>E. coli</i>	[97]
	PA	RO	GO	Covalent bonding	96.7%	65% bacterial inactivation	-	<i>E. coli</i>	[99]
	PA	RO	GO-Azide	Covalent bonding	-	Cell adhesion reduced by 94.1%, 90% inactivated	R _t reduced by 42.9% (BSA)	<i>E. coli</i> , BSA solution	[171]

Polypropylene	UF	GO-alkynyl; GO-azide	Layer-by-layer assembly	182%	Viability decreased by 66.7%	R _t decreased by 26.1%, FRR increased by 86% (BSA)	<i>E. coli</i> , BSA	[114]
PA	FO	GO-PLL	Covalent bonding	105.9%	99% inactivation	R _t decreased by 21.7%	Surface water	[40]
Cellulose nitrate	UF	GO-PVK	Deposition	-	Viability reduced by 88.6% and 93.6%	-	<i>E. coli</i> , <i>B.</i> <i>subtilis</i>	[172]
PA	RO	GO-Cs	Covalent bonding	-	-	R _t decreased by 66.7%, FRR increased by 12.8%	BSA	[173]
PA	FO	GO-AgNP	Covalent bonding	-	Viability decreased by 80%, Live cell biovolume decreased by 41%	R _t decreased by 58.2%	<i>P.</i> <i>aeruginosa</i>	[174]
PES	UF	GO-TiO ₂ -AgNP (<i>in situ</i>)	Covalent bonding	4.2%	3 log less live cells attached on membrane	-	<i>E. coli</i> , <i>B.</i> <i>subtilis</i>	[103]
PA	FO	GO-AgNP	Covalent bonding	98%	Viable cells decreased by 96%	-	<i>E. coli</i>	[39]
PANi, PA	Charged MF	rGO	Interfacial polymerization	15%	-	R _t decreased by 6.0%	Yeast	[175]
Polyester	Charged MF	rGO-PPy	Interfacial polymerization	-	-	R _t decreased by 5.2%	Yeast	[21]

1185 ^a PA: polyamide; PPSU: polyphenylsulfone; PANi: polyaniline; pDA: polydopamine; TMC: 1,3,5-benzenetricarbonyl trichloride; PE: polyethylene.

1186 ^b OMWCNTs: oxidized multiwall carbon nanotubes; TA: tannic acid; APTS: 3-aminopropyltriethoxysilane; EDA: ethylenediamine; PAA: polyacrylic acid; PSBMA:
1187 poly(sulfobetaine methacrylate); ZIF8: zeolitic imidazolate framework-8; Cs: chitosan; PLL: poly L-Lysine; PVK: poly(N-vinylcarbazole); PPy: polypyrrole; HNTs:
1188 halloysite nanotubes.



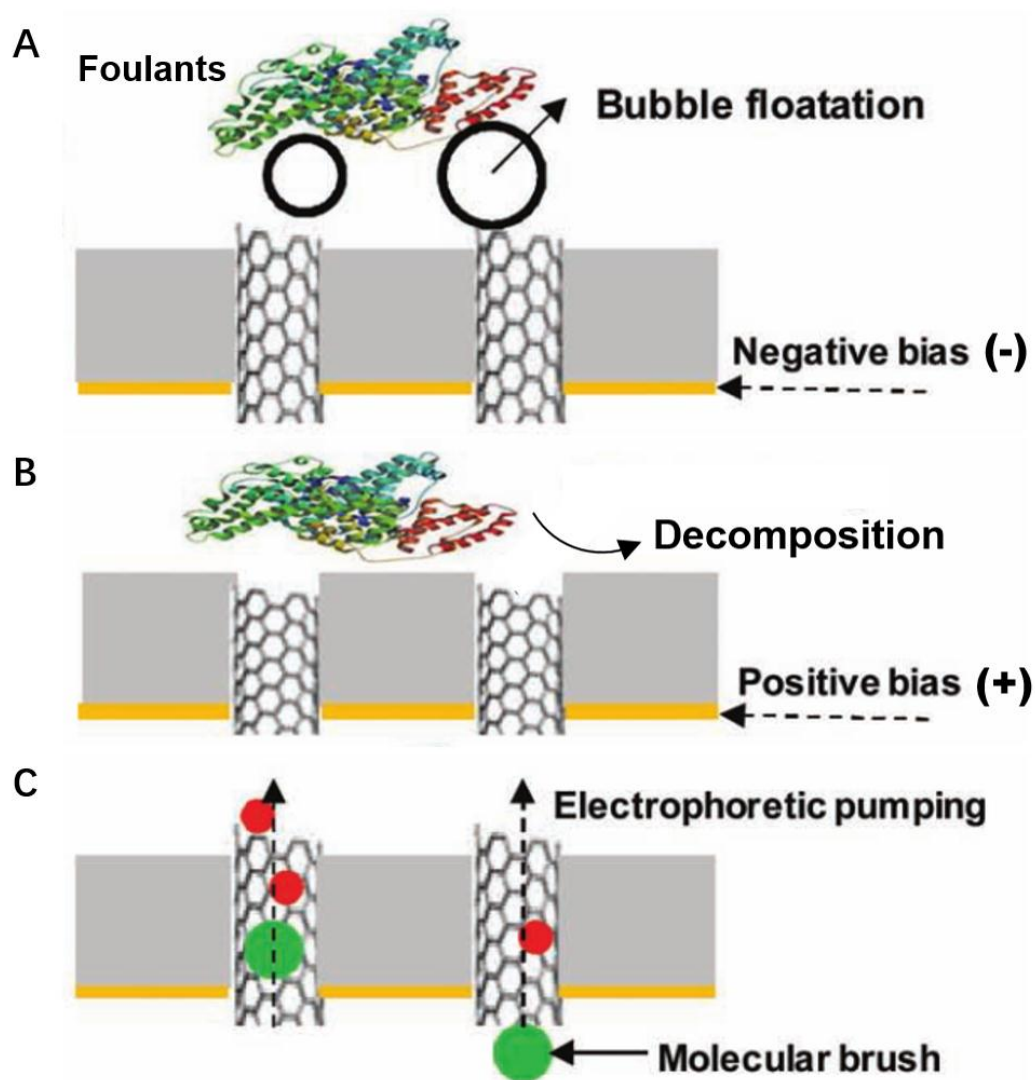
1189
 1190 **Figure 1.** The disinfection efficiency of different nanomaterials and main antimicrobial mechanisms
 1191 of graphene-family materials. GO, CNT and their derivatives displayed excellent antimicrobial
 1192 activities against *E. coli* (A) and *S. aureus* (B). Direct physical damage (C); ROS-mediated oxidative
 1193 stress (D); and bacterial isolation via wrapping around the bacterial surface (E). The figure is
 1194 reprinted with copyright permission [27,29].

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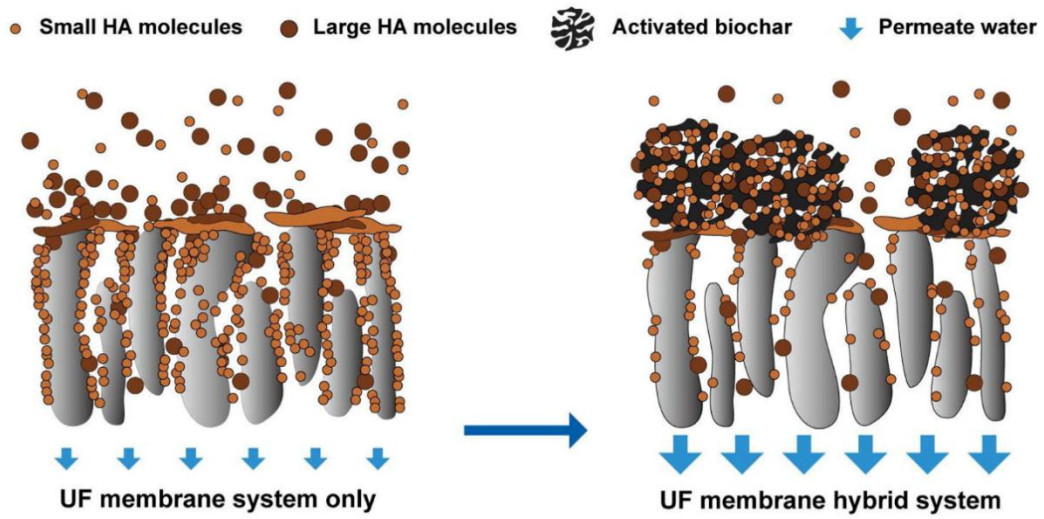
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1199

1200 **Figure 2.** Schematic diagram of electrically-assisted fouling mitigation: membrane cleaning via
 1201 bubbles generated by electro-reduction (A), foulants decomposed by electro-oxidation (B) and
 1202 membrane cleaning via ionic pumping (C). The figures are reprinted with copyright permissions
 1203 [2,54].

1204

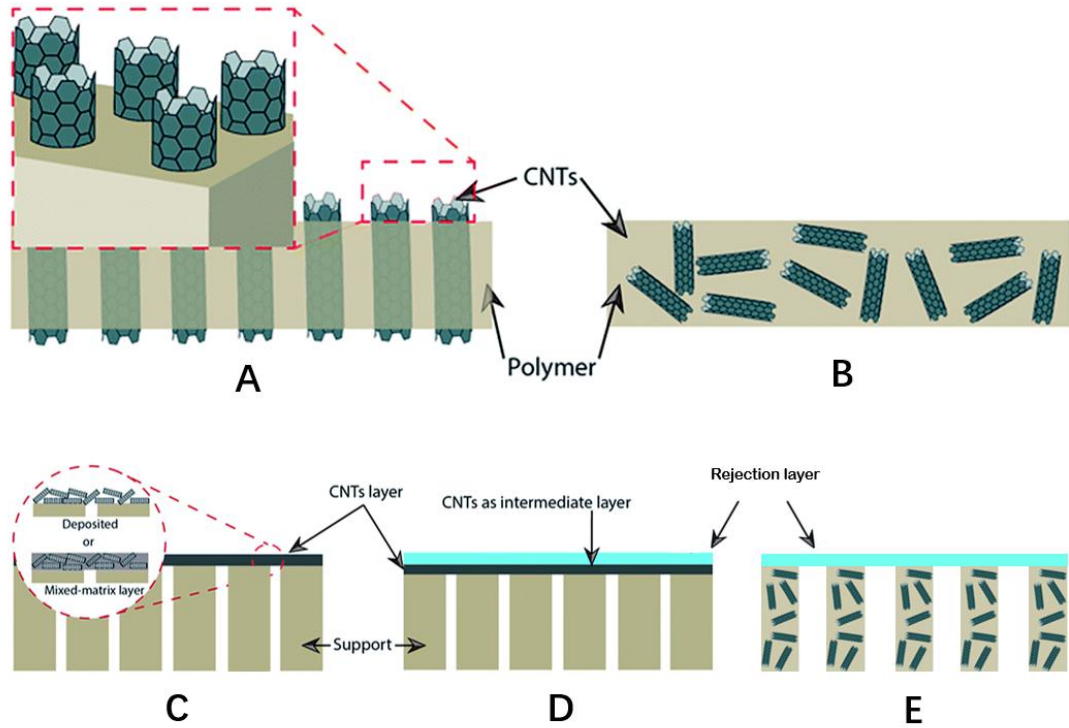


1205

1206 **Figure 3.** Application of biochar as an adsorbent to mitigate biofouling. The figure is reprinted with
 1207 copyright permission [62].

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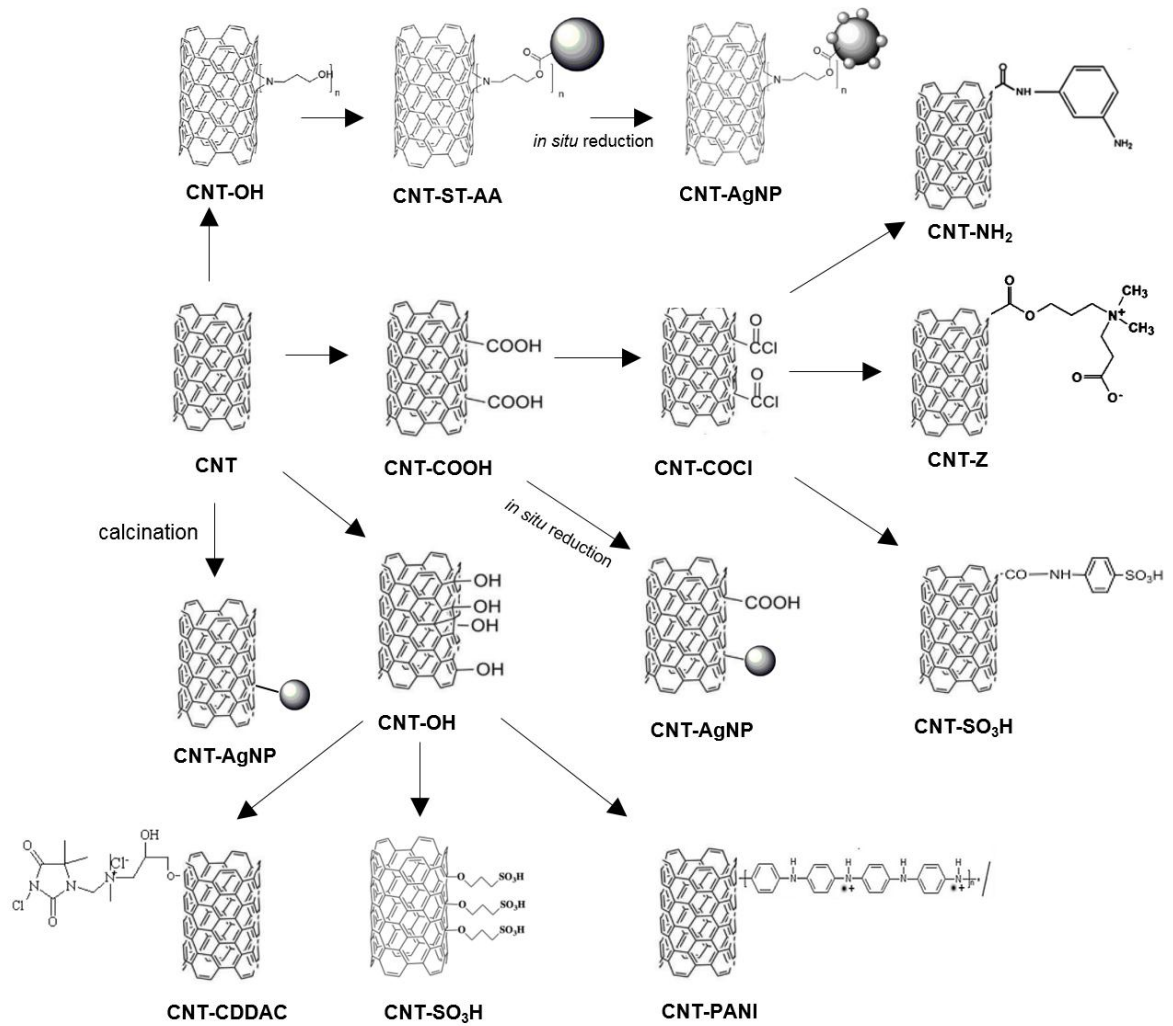
1211 **Figure 4.** CNT membranes with different structures. Vertically aligned (A) and mixed matrix (B)

1212 CNT in membranes. CNT deposited on membrane surface or support (C). CNT coated on

1213 membrane surface (support) as intermediate layer (D). CNT incorporated in support layer (E) [32].

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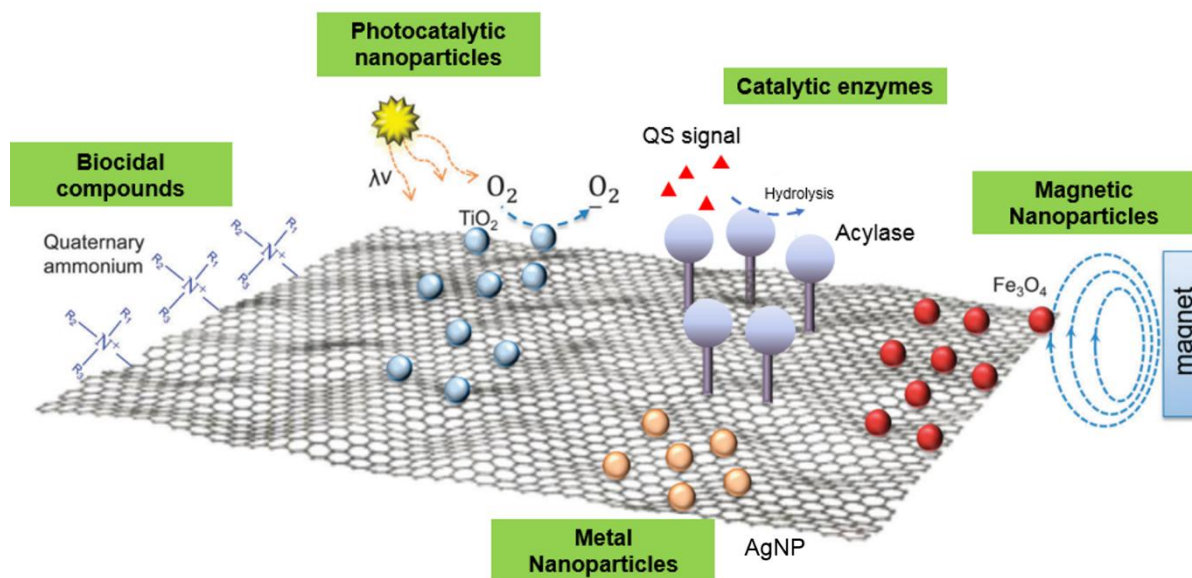


1216

1217 **Figure 5.** Functionalization of CNT for biofouling control. ST-AA: styrene-co-acrylic acid
 1218 microspheres; Z: zwitterionic group; PANI: polyaniline; CDDAC: (3-chloro-2-hydroxypropyl)-
 1219 (5,5-dimethylhydantoinyl-1-ylmethyl)-dimethylammonium chloride.

1220

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1222

1223 **Figure 6.** Different types of graphene-based antibiofouling nanocomposites [176].

1224