

New Biomarkers of Chronic Hepatitis B

Lung-Yi Mak¹, Wai-Kay Seto^{1,2,3}, James Fung^{1,2}, and Man-Fung Yuen^{1,2}

¹Department of Medicine, Queen Mary Hospital, The University of Hong Kong, ²State Key Laboratory for Liver Research, The University of Hong Kong, Hong Kong, and ³Department of Medicine, The University of Hong Kong-Shenzhen Hospital, Shenzhen, China

Chronic hepatitis B (CHB) infection leads to clinically heterogeneous disease outcomes. Different viral markers are utilized to monitor treatment effects and predict risk of complications in patients with CHB. Hepatitis B core-related antigen (HBcrAg) is a novel serum composite viral protein whose performance has been proven to be superior to that of existing viral markers. It showed good correlation with intrahepatic covalently closed-circular DNA. Its profile differs drastically in patients in different disease phases, and the level declines with antiviral therapies. HBcrAg may be helpful for predicting hepatocellular carcinoma development and hepatitis B virus (HBV) reactivation in immunosuppressed patients. Another emerging measurable serum marker, HBV RNA, exists in the form of pregenomic RNA-containing virions. Its profile differs between patients in different disease phases in a similar manner to that of HBcrAg. HBV RNA is present in serum at lower levels than HBV DNA in treatment-naïve patients by 1–2 logs. In contrast, its level is higher than HBV DNA in patients receiving nucleos(t)ide analogues (NAs). A significant decline in serum RNA was observed in patients receiving novel antiviral therapies, including core protein allosteric modulators and RIG-1/NOD2 agonists. Both HBcrAg and HBV RNA may be helpful for predicting off-therapy sustained virological control in patients who stop long-term NA treatment. (**Gut Liver 2019;13:589-595**)

Key Words: Hepatitis B core-related antigen; Hepatitis B virus RNA; Biomarkers

INTRODUCTION

Hepatitis B virus (HBV) is the only hepatotropic virus which exists in DNA form. It infects human livers and exerts necro-inflammatory, fibrotic and carcinogenic effects.¹ Most patients with chronic HBV (CHB) infection acquire the virus via vertical/

early-age horizontal transmission. Up to 15%–40% of them will progress to cirrhosis, decompensated liver disease, hepatocellular carcinoma (HCC) or death, and liver transplantation may be required in patients with advanced liver disease.² Current available antiviral therapies including pegylated interferon (PEG-IFN) and nucleos(t)ide analogues (NA)³ are effective in controlling or suppressing viral replication. However, a complete cure, as defined by the total eradication of the virus from the liver, is not achieved due to the persistence of covalently closed circular DNA (cccDNA).^{4,5} The surrogate treatment endpoint of HBV surface antigen (HBsAg) seroclearance, known as a functional cure, is deemed more feasible, and has been shown to be associated with significantly lower risk of liver-related complications,⁶ despite being only achieved in a minority of patients.^{7–9} Ongoing efforts are made to develop novel anti-HBV drugs to act against various steps of the HBV replication cycle, aiming to enhance virological control and promote functional cure.¹⁰ For the majority of patients with CHB who do not achieve functional cure, long term NA is likely needed. In spite of this long-term therapy, liver-related complications can still occur even with sustained viral suppression. To this end, newer virological markers have been developed to predict the risk of liver-related complications in these patients who often have undetectable serum HBV DNA, and the likelihood of achieving functional cure or partial cure, which is defined as off-therapy virological suppression. In the following sections, two novel serum biomarkers developed for these purposes will be discussed: HBV core-related antigen (HBcrAg) and HBV RNA.

HEPATITIS B CORE-RELATED ANTIGEN

1. Overview

Following viral entry into the hepatocyte, the relaxed circular DNA (rcDNA) is converted into cccDNA minichromosome, which is used as a template for subsequent transcription and

Correspondence to: Man-Fung Yuen

Department of Medicine, Queen Mary Hospital, The University of Hong Kong, Pokfulam Road 102, Hong Kong

Tel: +852-22553994, Fax: +852-28162863, E-mail: mfyuen@hkucc.hku.hk

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translation into viral proteins. Among the many viral proteins synthesized, three related viral proteins, sharing an identical 149 amino acid sequence, make up the HBcrAg. These include the HBV core antigen (HBcAg)—structural component of the viral capsid, HBV e antigen (HBeAg)—the N-terminal processed product of the precore protein, and a truncated 22 kDa precore protein (p22Cr)—also processed product of precore protein with additional protein processing at both the N- and C-terminals (Fig. 1).^{11,12} Serum HBcrAg can be detected and quantified with the chemiluminescence method.¹¹ In the following sections, the clinical relevance of HBcrAg will be discussed.

2. Profile in natural history of CHB

The profile of serum HBcrAg in different disease phases of CHB has been characterized by two studies involving Asian and European patients with genotype A-D HBV infection.^{13,14} In both studies, the serum HBcrAg level was significantly higher in HBeAg-positive patients compared to HBeAg-negative patients, owing to the fact that HBeAg production (one of the components of HBcrAg) was diminished after HBeAg seroconversion. More importantly, serum HBcrAg may be able to differentiate between chronic infection and chronic hepatitis. For HBeAg-positive patients, chronic infection was associated with higher HBcrAg than chronic hepatitis (8.54 and 7.92 log U/mL, respectively, $p < 0.001$). For HBeAg-negative patients, chronic infection was associated with lower HBcrAg than chronic hepatitis (2.60 and 4.92 log U/mL, respectively, $p < 0.001$).¹⁴ For HBsAg-negative patients, that is, those achieved functional cure, serum HBcrAg was only detected in 21% with a median level of 2.7 log U/mL.^{14,15}

Serum HBcrAg correlated well with serum HBV DNA (r , 0.69

to 0.87; $p < 0.001$),¹⁶⁻¹⁹ serum HBsAg titre ($r = 0.703$, $p < 0.001$),¹⁴ intrahepatic total DNA (r , 0.664 to 0.70; $p < 0.001$)^{18,19} and intrahepatic cccDNA ($r = 0.664$, $p < 0.001$) (Table 1).¹⁸

3. Profile in patients undergoing antiviral treatment

Although NAs are potent inhibitors of viral replication with marked HBV DNA suppression, the ongoing inhibitory effects of NA on intrahepatic viral reservoir, as reflected by progressive decline of other viral markers, is more modest. In a study of 222 patients on entecavir (ETV) for 7 years, serum HBcrAg declined at a rate of 0.244 log kU/mL/year, which was more impressive than the decline in serum HBsAg titre of 0.107 log IU/mL/year.²⁰ Almost one-third (32%) even achieved undetectable serum HBcrAg at year 7.²⁰ Serum HBcrAg showed persisted linear correlation with intrahepatic cccDNA, even in treatment-experienced patients ($r = 0.692$, $p < 0.001$).²¹ The decline in serum HBcrAg observed in patients on long term NA therapy correlated modestly with decline in the intrahepatic cccDNA ($r = 0.419$, $p = 0.005$).²²

Less data exists for the kinetics of HBcrAg in patients treated with PEG-IFN. One study ($n = 58$) reported significant decline of serum HBcrAg from 8.042 log U/mL at baseline to 5.301 log U/mL at 24 weeks after completion of a 48-week course of PEG-IFN.²³ Due to the small number of patients and short follow-up duration, the effects of a finite duration PEG-IFN on the profile of serum HBcrAg in the long term remains elusive.

The kinetics of HBcrAg using novel anti-HBV compounds has been described. ARC-520 is a RNA interference agent which directly targets cccDNA-derived transcription and blocks downstream viral replicatory steps. After a single dose of intravenous ARC-520, serum HBcrAg reduced by 1.4 log kU/mL at 85 days of dosing.²⁴ ARC-520 initially entered phase 2 clinical trial, but

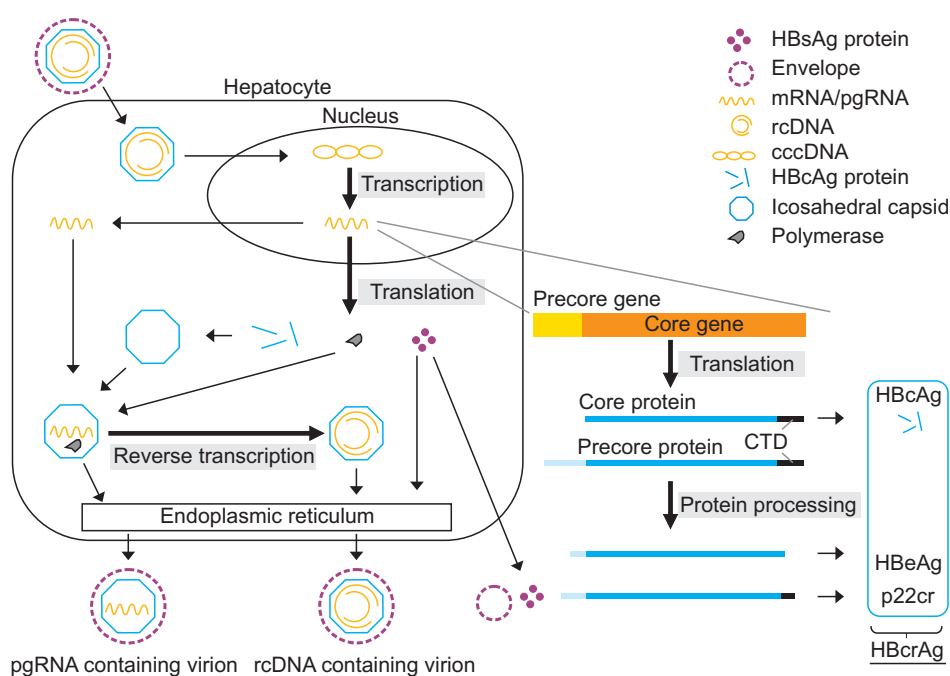


Fig. 1. Schematic illustration of the hepatitis B virus (HBV) replication cycle and production of viral markers. HBsAg, HBV surface antigen; mRNA, messenger RNA; pgRNA, pregenomic RNA; rcDNA, relaxed circular DNA; cccDNA, covalently closed circular DNA; HBcAg, HBV core antigen; CTD, C-terminal domain; HBeAg, HBV e antigen; p22cr, truncated 22 kDa precore protein; HBcrAg, hepatitis B core-related antigen.

Table 1. Correlation between Serum HBcrAg and HBV RNA with Respect to Intrahepatic cccDNA

	Treatment naïve		Treatment experienced	
	r	p-value	r	p-value
HBcrAg	0.664–0.70	<0.001 ^{18,19}	0.692	<0.001 ²¹
HBV RNA	HBeAg-positive: 0.39	0.002 ⁴⁵	0.08	0.55 ⁴⁴
	HBeAg-negative: 0.1	0.654 ⁴⁵		

HBcrAg, hepatitis B core-related antigen; HBV, hepatitis B virus; cccDNA, covalently closed circular DNA; HBeAg, hepatitis B e antigen.

was subsequently discontinued due to deaths observed in non-human primates receiving excessive doses. Nevertheless, other RNA interference compounds are being actively developed, and the findings from the ARC-520 study showed that serum HBcrAg may be a suitable marker for monitoring of the treatment effects.

4. Potential clinical applications

Previous studies have demonstrated a potential role for HBcrAg in determining risk for HCC development. Higher serum HBcrAg was shown to be associated with HCC development in both treatment-naïve and treatment-experienced patients. In 1,031 treatment-naïve patients where 78 developed HCC at a median of 10.7 years, serum HBcrAg >2.9 log U/mL was associated with 5-fold increased risk of HCC.²⁵ In 76 patients treated with NA who had undetectable serum HBV DNA, post-treatment serum HBcrAg of >3.89 log U/mL was associated with 3-fold increased risk of HCC.²⁶

The risk of HBV reactivation after immunosuppressive therapy may also be predicted by serum HBcrAg in patients with occult hepatitis B infection. In 124 HBsAg-/anti-HBc+ patients with undetectable serum HBV DNA who received rituximab or underwent allogeneic hematopoietic stem cell transplantation, cumulative rate of HBV reactivation was 40.4% at 2-year. Serum HBcrAg at baseline was detectable in 43 patients (34.7%) and was associated with almost 3-fold increased risk of HBV reactivation compared to patients without detectable serum HBcrAg.²⁷

Levels of HBcrAg may also potentially predict the likelihood of achieving partial cure in patients on antiviral therapy, as defined by a sustained off-therapy virological control. These patients may be able to discontinue NA with close monitoring for virological breakthroughs. Different markers, including HBsAg titre, have been reported to show predictive ability for post-NA cessation virological relapse.²⁸ The role of serum HBcrAg in this special population was also investigated in 45 HBeAg-negative patients, showing that a high end-of-therapy serum HBcrAg >3.7 log IU/mL was associated with 3.7-fold risk of virological relapse within one year of NA cessation.²⁹

From the above studies, it is clear that serum HBcrAg levels would carry multiple implications depending on the context of measurement. Extra care should be taken when interpreting

the results, taking into account the HBsAg status, the phase of disease, the duration of antiviral treatment and the indication of measurement.³⁰

HEPATITIS B VIRUS RNA

1. Overview

Earlier concepts regarding HBV replication did not acknowledge the presence of HBV RNA outside hepatocytes, as completion of rcDNA synthesis, the so-called “mature genome,” was considered to be necessary for envelopment and exit of virions to the blood stream.^{31,32} In recent studies, HBV RNA have been shown to be detectable in serum of patients who are either treatment-naïve or treatment-experienced, challenging the concept that only rcDNA-containing virions can be secreted. Indeed, the serum HBV RNA is present in the form of virion containing primarily full-length pregenomic RNA (pgRNA), which is encapsidated by Hbc protein, rather than preC mRNA that bears the same length of 3.5 kb, nor is it associated with exosomes (Fig. 1).³³ Most studies measure serum HBV RNA using standardized multistep procedures. In simple terms, these include serum HBV nucleic acid extraction, DNA digestion, reverse transcription of RNA (targeting specific HBV gene regions) to complementary DNA, followed by real-time polymerase chain reaction.³⁴⁻⁴¹ In the following sections, the clinical relevance of circulating HBV RNA will be discussed.

2. Profile in natural history of CHB

In general, HBV RNA is present in treatment naïve CHB patients in lower levels compared to HBV DNA by approximately 1–2 log₁₀ copies/mL.⁴² In a study of 11 patients, serum HBV DNA and HBV RNA were 7.87 and 6.31 log₁₀ copies/mL, respectively (p=0.007).³³ In another study characterizing HBV RNA in the natural history of CHB in a larger population of 102 untreated patients, serum HBV RNA was found to differ significantly between the disease phases, a phenomenon similar to HBcrAg. The highest levels were observed in HBeAg-positive chronic infection, followed by HBeAg-positive chronic hepatitis, HBeAg-negative chronic hepatitis, and lowest in HBeAg-negative chronic infection (6.78, 5.73, 4.52, and 2.96 log₁₀ copies/mL, respectively; p<0.001 for inter-group comparisons and for trend).⁴³

Serum HBV RNA demonstrated different degrees of correlation with known viral markers in treatment naïve patients. Regarding serum HBV DNA, there was strong linear correlation between these two viral nucleic acids ($r=0.928$, $p<0.001$).⁴³ Serum HBV RNA level also showed good linear correlation with serum HBsAg titre ($r=0.67$, $p<0.001$).⁴⁴ In comparison, serum HBV RNA showed only modest linear correlations with intrahepatic cccDNA in HBeAg-positive patients ($r=0.39$, $p=0.002$ ⁴⁵ or $r=0.25$, $p=0.02$ ⁴⁴) and no significant correlation in HBeAg-negative patients ($r=0.10$ $p=0.654$) (Table 1).⁴⁵

3. Profile in patients undergoing approved antiviral treatment

In contrast to untreated patients, a relatively higher serum HBV RNA level with respect to HBV DNA level was observed in patients treated with NAs.⁴² This can be explained by the fact that HBV pgRNA is only produced from cccDNA by transcription. Any secreted pgRNA-containing virions originate from partially or un-transcribed pgRNA, which is paradoxically increased when reverse transcriptase and DNA polymerase activities are inhibited by NA.³³ Therefore, it is not surprising that serum HBV RNA could still be detected even in patients who had undetectable serum HBV DNA after NA treatment. In a cross-sectional study involving 47 patients treated with ETV for a median duration of 3 years with undetectable serum HBV DNA, serum HBV RNA was still detected in 35 (74.47%) at a median level of 3.02 (range, 2.33 to 4.80) \log_{10} copies/mL. There was significant correlation between serum HBV RNA and serum HBsAg titre ($r=0.665$, $p<0.001$), intrahepatic HBV RNA level ($r=0.725$, $p<0.001$) as well as histological disease severity in terms of grading of necro-inflammation ($r=0.665$, $p<0.001$) and staging. However, intrahepatic cccDNA showed no correlation with serum HBV RNA after antiviral therapy (Table 1).⁴⁴

Serum HBV RNA might also be useful in predicting treatment response in patients receiving NA monotherapy, and in NA-induced HBeAg seroconversion. A low on-treatment serum HBV RNA at week 12 of lamivudine or ETV was associated with shorter interval to achieve serum HBV DNA undetectability ($n=5$).⁴⁰ Among 50 HBeAg-positive patients who received NA for a mean duration of 19 months, 15 patients who achieved HBeAg seroconversion had significantly greater decline in serum HBV full-length RNA from baseline by 1 \log_{10} and 1.8 \log_{10} at 3 months and 6 months, respectively ($p<0.001$ for months 3 and 6), compared to 35 patients without HBeAg seroconversion.³⁹

The profile of serum HBV RNA in patients receiving combination PEG-IFN and NA therapy was also reported. Similar to NA monotherapy, serum HBV RNA was relatively higher compared to serum HBV DNA. This was demonstrated in a study of 23 patients (13 HBeAg-positive and 10 HBeAg-negative) who were treated with 48 weeks combination PEG-IFN alfa-2a with adefovir. Moreover, the decline in HBV RNA levels in HBeAg-

positive patients was greater for those on combination therapy from 30 weeks onwards compared to patients on NA monotherapy. In both HBeAg-positive and HBeAg-negative patients, responders to combination therapy had significantly lower serum HBV RNA levels compared to non-responders from 30 weeks onwards, and baseline HBV RNA was independently associated with treatment response (odds ratio=0.44, $p=0.019$).³⁵

4. Profile in patients undergoing novel anti-HBV drug trials

Owing to the different mechanisms of action against HBV in novel therapies, traditional viral markers may not be sufficient to reflect the mechanistic effects of these novel agents. Novel markers including serum HBV RNA therefore emerge as an important marker to monitor their antiviral effects. The profile of serum HBV RNA has been characterized in two classes of novel agents: core protein allosteric modulators (CpAM) and retinoic acid-inducible gene 1/nucleotide-binding oligomerization domain-containing protein 2 (RIG-I/NOD2) agonist.

For CpAM, serum HBV RNA was measured in 73 HBeAg-positive treatment-naïve patients who were treated with NVR3-778, PEG-IFN or NVR3-778 + PEG-IFN. After 28 days of therapy, all patients treated with 600 mg BD or 1,000 mg BD NVR3-778 monotherapy had serum HBV RNA decline by $>0.5 \log_{10}$ IU/mL, and all patients treated with combination NVR3-778 + PEG-IFN had serum HBV RNA decline by $>1.0 \log_{10}$ copies/mL. The biggest drop of HBV RNA from baseline was observed in patients treated with combination NVR3-778 + PEG-IFN ($-2.06 \log_{10}$ copies/mL for cohort taking 600 mg BD NVR3-778), followed by NVR3-778 monotherapy ($-1.42 \log_{10}$ copies/mL), and PEG-IFN monotherapy ($-0.89 \log_{10}$ copies/mL). There was high linear correlation between serum HBV RNA and DNA levels ($r=0.91$, $p<0.0001$).⁴⁶ The decline in HBV RNA with NVR3-778 is not surprising. CpAM inhibits the formation of normal icosahedral capsids, which is responsible for the encapsidation of pgRNA, a prerequisite for subsequent excretion into the circulation as RNA-containing virions.

For RIG-I/NOD2 agonist, serum HBV RNA was measured in 20 patients who were treated with SB 9200 (Inarigivir soproxil) for 12 weeks at different doses followed by 12 weeks of TDF. At week 12 of therapy, compared to placebo, significant decline in serum HBV RNA from baseline was observed only in 11 HBeAg-negative patients but not in the nine HBeAg-positive patients. In addition, the serum HBV RNA decline observed in HBeAg-negative patients was dose-dependent: $-1.84 \log_{10}$ and $-3.15 \log_{10}$ for SB 9200 (25 mg) and SB 9200 (50 mg), respectively. Of the 11 HBeAg-negative patients, nine had undetectable serum HBV RNA at week 24. In contrast, only two out of the nine HBeAg-positive patients had undetectable serum HBV RNA at the same time point.⁴⁷ RIG-I and NOD2 are host pattern recognition receptors which are viral sensor proteins that induces interferon-mediated antiviral immune responses in virus-infected cells. In addition, RIG-1 binding to pgRNA suppresses

its encapsidation, leading to a reduction in RNA-containing virion secretion.⁴⁸

Since novel anti-HBV agents act on different steps from NAs in the viral replication cycle, serum HBV DNA is no longer sufficient to assess treatment response for these newer compounds. From the above examples, serum HBV RNA might act as an additional useful marker for monitoring the direct antiviral effects of these novel therapies.

5. Potential clinical applications

The need to identify patients with high risk of post-NA cessation relapse has been discussed above. Apart from serum HBcrAg, serum HBV RNA has also been shown to predict virological relapse after NA cessation. In a study of 36 patients who stopped NA after a mean treatment duration of 36 weeks, 19 (52.8%) developed post-NA cessation viral rebound at 24 weeks after treatment discontinuation. On-therapy HBV DNA + RNA titres decreased rapidly in the non-rebound group compared to that in the rebound group. More specifically, on-therapy high serum HBV DNA + HBV RNA titres at 3 months was significantly associated with viral rebound (odds ratio=9.474, $p=0.043$).⁴¹ In another study of 33 patients who had taken >3 years of NA with undetectable serum HBV DNA, 21 still had detectable serum HBV RNA at the time of treatment cessation. At 24 weeks after NA cessation, viral rebound occurred in all 21 patients (100%), compared to only three (25%) of the 12 patients with undetectable serum HBV RNA ($p=0.001$).³³ The presence of pgRNA virion in the serum may reflect the ongoing

transcriptional activity of cccDNA and signifies lower chance of sustained off-therapy virological suppression.

Serum RNA has also been studied in patients after acute HBV infection. Currently, no accurate marker exists to predict the persistence of HBV after acute infection. It often relies on clinical history and repeated serological monitoring of HBsAg and HBV DNA. Serum HBV RNA was characterized in three blood donor samples with positive IgM anti-HBc (consistent with acute HBV infection). Serum pgRNA was detectable throughout the viraemic phase, being 2.33 to 2.58 log lower than serum HBV DNA in two patients who had resolved HBV infection, compared to 1.85 log lower than serum HBV DNA in the remaining patient who had persistent HBsAg seropositivity for >6 months. However, no significant differences could be observed in terms of correlations of RNA and DNA between the two patients who had resolved HBV infection compared to the patient who evolved into chronic HBV carrier.⁴² The role of serum HBV RNA in prediction of HBV chronicity after acute exposure remains to be determined.

CONCLUSION AND FUTURE DIRECTIONS

Both serum HBcrAg and HBV RNA are emerging markers for diagnosis, monitoring and prognostication in patients with CHB. Table 2 summarizes the potential clinical applications of HBcrAg and HBV RNA in CHB infection. To date, serum HBcrAg has been shown to be the best surrogate marker for intrahepatic cccDNA. It is increasingly studied as a potential

Table 2. Potential Clinical Application of HBcrAg and HBV RNA in Chronic Hepatitis B Infection

Area of interest	HBcrAg	HBV RNA
Natural history		
Differentiate disease phases	+	+
Predicts spontaneous HBeAg seroconversion	+	No data
Predicts spontaneous HBsAg seroclearance	(-)*	No data
Antiviral treatment: PEG-IFN or NA		
Predict treatment-induced HBeAg seroconversion	+	+
Predict post-NA cessation flare	+	+
Clinical trials of new antiviral agents		
Dynamic change in siRNA	+	No data
Dynamic change in CpAM	No data	+
Dynamic change in RIG-I + NOD2 agonist	No data	+
Special populations		
Predict HCC development	+	No data
Predict reactivation of HBV under immunosuppression	+	No data
Profile in acute infection	No data	(-) [†]

HBcrAg, hepatitis B core-related antigen; HBV, hepatitis B virus; HBeAg, hepatitis B virus e antigen; HBsAg, hepatitis B virus surface antigen; PEG-IFN, pegylated interferon; NA, nucleos(t)ide analogues; siRNA, short interfering RNA; CpAM, core protein allosteric modulators; RIG-I, retinoic acid-inducible gene I; NOD2, nucleotide-binding oligomerization domain-containing protein 2; HCC, hepatocellular carcinoma.

*Profile of HBcrAg reported in patients with HBsAg seroclearance, but no further data on predictive power; [†]Only data on 2 patients with acute HBV infection was reported (see text).

predictor for HCC development, post-NA cessation relapse, and HBV reactivation in patients undergoing immunosuppressive therapy. More validation studies are needed to identify specific cutoff values for each clinical outcome. Serum HBV RNA exists as pgRNA-containing virions and reflects upstream viral replication activities. The most relevant application of serum HBV RNA appears to be as a monitoring tool for treatment effect in patients receiving novel anti-HBV therapies. Meanwhile, the specific methods and technical details of serum RNA detection vary widely between different studies and standardization of such is urgently needed. Moreover, whether serum HBV RNA has a role in patients with occult hepatitis B infection, as well as in prediction of HBsAg seroclearance, HCC development and HBV reactivation in immunosuppressed patients remains to be illustrated by future studies.

CONFLICTS OF INTEREST

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REFERENCES

1. Yuen MF, Chen DS, Dusheiko GM, et al. Hepatitis B virus infection. *Nat Rev Dis Primers* 2018;4:18035.
2. Lai CL, Yuen MF. The natural history and treatment of chronic hepatitis B: a critical evaluation of standard treatment criteria and end points. *Ann Intern Med* 2007;147:58-61.
3. European Association for the Study of the Liver. EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection. *J Hepatol* 2017;67:370-398.
4. Zoulim F. New insight on hepatitis B virus persistence from the study of intrahepatic viral cccDNA. *J Hepatol* 2005;42:302-308.
5. Mason AL, Xu L, Guo L, Kuhns M, Perrillo RP. Molecular basis for persistent hepatitis B virus infection in the liver after clearance of serum hepatitis B surface antigen. *Hepatology* 1998;27:1736-1742.
6. Yuen MF, Wong DK, Fung J, et al. HBsAg seroclearance in chronic hepatitis B in Asian patients: replicative level and risk of hepatocellular carcinoma. *Gastroenterology* 2008;135:1192-1199.
7. Fung J, Cheung KS, Wong DK, et al. Long-term outcomes and predictive scores for hepatocellular carcinoma and hepatitis B surface antigen seroclearance after hepatitis B e-antigen seroclearance. *Hepatology* 2018;68:462-472.
8. Kim GA, Lim YS, An J, et al. HBsAg seroclearance after nucleoside analogue therapy in patients with chronic hepatitis B: clinical outcomes and durability. *Gut* 2014;63:1325-1332.
9. Liu J, Yang HI, Lee MH, et al. Incidence and determinants of spontaneous hepatitis B surface antigen seroclearance: a community-based follow-up study. *Gastroenterology* 2010;139:474-482.
10. Durantel D, Zoulim F. New antiviral targets for innovative treatment concepts for hepatitis B virus and hepatitis delta virus. *J Hepatol* 2016;64:S117-S131.
11. Kimura T, Ohno N, Terada N, et al. Hepatitis B virus DNA-negative Dane particles lack core protein but contain a 22-kDa precore protein without C-terminal arginine-rich domain. *J Biol Chem* 2005;280:21713-21719.
12. Kimura T, Rokuhara A, Sakamoto Y, et al. Sensitive enzyme immunoassay for hepatitis B virus core-related antigens and their correlation to virus load. *J Clin Microbiol* 2002;40:439-445.
13. Maasoumy B, Wiegand SB, Jaroszewicz J, et al. Hepatitis B core-related antigen (HBcrAg) levels in the natural history of hepatitis B virus infection in a large European cohort predominantly infected with genotypes A and D. *Clin Microbiol Infect* 2015;21:606.
14. Seto WK, Wong DK, Fung J, et al. Linearized hepatitis B surface antigen and hepatitis B core-related antigen in the natural history of chronic hepatitis B. *Clin Microbiol Infect* 2014;20:1173-1180.
15. Seto WK, Tanaka Y, Wong DK, et al. Evidence of serologic activity in chronic hepatitis B after surface antigen (HBsAg) seroclearance documented by conventional HBsAg assay. *Hepatol Int* 2012;7:98-105.
16. Rokuhara A, Sun X, Tanaka E, et al. Hepatitis B virus core and core-related antigen quantitation in Chinese patients with chronic genotype B and C hepatitis B virus infection. *J Gastroenterol Hepatol* 2005;20:1726-1730.
17. Rokuhara A, Tanaka E, Matsumoto A, et al. Clinical evaluation of a new enzyme immunoassay for hepatitis B virus core-related antigen; a marker distinct from viral DNA for monitoring lamivudine treatment. *J Viral Hepat* 2003;10:324-330.
18. Wong DK, Tanaka Y, Lai CL, Mizokami M, Fung J, Yuen MF. Hepatitis B virus core-related antigens as markers for monitoring chronic hepatitis B infection. *J Clin Microbiol* 2007;45:3942-3947.
19. Wong DK, Seto WK, Cheung KS, et al. Hepatitis B virus core-related antigen as a surrogate marker for covalently closed circular DNA. *Liver Int* 2017;37:995-1001.
20. Lam YF, Seto WK, Wong D, et al. Seven-year treatment outcome of entecavir in a real-world cohort: effects on clinical parameters, HBsAg and HBcrAg levels. *Clin Transl Gastroenterol* 2017;8:e125.
21. Suzuki F, Miyakoshi H, Kobayashi M, Kumada H. Correlation between serum hepatitis B virus core-related antigen and intrahepatic covalently closed circular DNA in chronic hepatitis B patients. *J Med Virol* 2009;81:27-33.
22. Lai CL, Wong D, Ip P, et al. Reduction of covalently closed circular DNA with long-term nucleos(t)ide analogue treatment in chronic hepatitis B. *J Hepatol* 2017;66:275-281.
23. Ma H, Yang RF, Li XH, Jin Q, Wei L. HBcrAg identifies patients failing to achieve HBeAg seroconversion treated with pegylated

- interferon Alfa-2b. *Chin Med J (Engl)* 2016;129:2212-2219.
24. Yuen MF, Chan HLY, Liu KS, et al. Differential reductions in viral antigens expressed from CCCDNAs integrated DNA in treatment naïve HBeAg positive and negative patients with chronic HBV after RNA interference therapy with ARC-520. *J Hepatol* 2016;64:S390-S391.
 25. Tada T, Kumada T, Toyoda H, et al. HBcrAg predicts hepatocellular carcinoma development: an analysis using time-dependent receiver operating characteristics. *J Hepatol* 2016;65:48-56.
 26. Cheung KS, Seto WK, Wong DK, Lai CL, Yuen MF. Relationship between HBsAg, HBcrAg and hepatocellular carcinoma in patients with undetectable HBV DNA under nucleos(t)ide therapy. *J Viral Hepat* 2017;24:654-661.
 27. Seto WK, Wong DK, Chan TS, et al. Association of hepatitis B core-related antigen with hepatitis B virus reactivation in occult viral carriers undergoing high-risk immunosuppressive therapy. *Am J Gastroenterol* 2016;111:1788-1795.
 28. Lee HA, Seo YS, Park SW, et al. Hepatitis B surface antigen titer is a good indicator of durable viral response after entecavir off-treatment for chronic hepatitis B. *Clin Mol Hepatol* 2016;22:382-389.
 29. Jung KS, Park JY, Chon YE, et al. Clinical outcomes and predictors for relapse after cessation of oral antiviral treatment in chronic hepatitis B patients. *J Gastroenterol* 2016;51:830-839.
 30. Mak LY, Wong DK, Cheung KS, Seto WK, Lai CL, Yuen MF. Review article: hepatitis B core-related antigen (HBcrAg): an emerging marker for chronic hepatitis B virus infection. *Aliment Pharmacol Ther* 2018;47:43-54.
 31. Gerelisaikhan T, Tavis JE, Bruss V. Hepatitis B virus nucleocapsid envelopment does not occur without genomic DNA synthesis. *J Virol* 1996;70:4269-4274.
 32. Perlman D, Hu J. Duck hepatitis B virus virion secretion requires a double-stranded DNA genome. *J Virol* 2003;77:2287-2294.
 33. Wang J, Shen T, Huang X, et al. Serum hepatitis B virus RNA is encapsidated pregenome RNA that may be associated with persistence of viral infection and rebound. *J Hepatol* 2016;65:700-710.
 34. Rokuhara A, Matsumoto A, Tanaka E, et al. Hepatitis B virus RNA is measurable in serum and can be a new marker for monitoring lamivudine therapy. *J Gastroenterol* 2006;41:785-790.
 35. Jansen L, Kootstra NA, van Dort KA, Takkenberg RB, Reesink HW, Zaaijer HL. Hepatitis B virus pregenomic RNA is present in virions in plasma and is associated with a response to pegylated interferon alfa-2a and nucleos(t)ide analogues. *J Infect Dis* 2016;213:224-232.
 36. Hatakeyama T, Noguchi C, Hiraga N, et al. Serum HBV RNA is a predictor of early emergence of the YMDD mutant in patients treated with lamivudine. *Hepatology* 2007;45:1179-1186.
 37. Kurosaki M, Tsuchiya K, Nakanishi H, Itakura J, Izumi N. Serum HBV RNA as a possible marker of HBV replication in the liver during nucleos(t)ide analogue therapy. *J Gastroenterol* 2013;48:777-778.
 38. Su Q, Wang SF, Chang TE, et al. Circulating hepatitis B virus nucleic acids in chronic infection: representation of differently polyadenylated viral transcripts during progression to nonreplicative stages. *Clin Cancer Res* 2001;7:2005-2015.
 39. van Bömmel F, Bartens A, Mysickova A, et al. Serum hepatitis B virus RNA levels as an early predictor of hepatitis B envelope antigen seroconversion during treatment with polymerase inhibitors. *Hepatology* 2015;61:66-76.
 40. Huang YW, Takahashi S, Tsuge M, et al. On-treatment low serum HBV RNA level predicts initial virological response in chronic hepatitis B patients receiving nucleoside analogue therapy. *Antivir Ther* 2015;20:369-375.
 41. Tsuge M, Murakami E, Imamura M, et al. Serum HBV RNA and HBeAg are useful markers for the safe discontinuation of nucleoside analogue treatments in chronic hepatitis B patients. *J Gastroenterol* 2013;48:1188-1204.
 42. Butler EK, Gersch J, McNamara A, et al. HBV serum DNA and RNA levels in nucleos(t)ide analogue-treated or untreated patients during chronic and acute infection. *Hepatology* 2018;68:2016-2117.
 43. Wang J, Yu Y, Li G, et al. Natural history of serum HBV-RNA in chronic HBV infection. *J Viral Hepat* 2018;25:1038-1047.
 44. Gao Y, Li Y, Meng Q, et al. Serum hepatitis B virus DNA, RNA, and HBsAg: which correlated better with intrahepatic covalently closed circular DNA before and after nucleos(t)ide analogue treatment? *J Clin Microbiol* 2017;55:2972-2982.
 45. Wang J, Du M, Huang H, et al. Reply to: "Serum HBV pgRNA as a clinical marker for cccDNA activity": consistent loss of serum HBV RNA might predict the "para-functional cure" of chronic hepatitis B. *J Hepatol* 2017;66:462-463.
 46. Yuen MF, Kim DJ, Weilert F, et al. NVR3-778, a first-in-class HBV core inhibitor, alone and in combination with Peg-interferon (Peg-IFN), in treatment-naïve HBeAg-positive patients: early reductions in HBV DNA and HBeAg. *J Hepatol* 2016;64:S210-S211.
 47. Yuen MF, Coffin CS, Elkhatab SG, et al. SB 9200 an oral selective immunomodulator is safe and efficacious in treatment-naïve, non-cirrhotic HBV patients: results from cohort 1 of the ACHIEVE trial. *Hepatology* 2017;66:22-23A.
 48. Sato S, Li K, Kameyama T, et al. The RNA sensor RIG-I dually functions as an innate sensor and direct antiviral factor for hepatitis B virus. *Immunity* 2015;42:123-132.