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Differential cytokine expression in EBV positive peripheral T cell lymphomas

J W Y Ho, R H S Liang, G Srivastava

Abstract

Aim—To investigate whether specific cytokines are secreted locally at the tumour site in Epstein-Barr virus (EBV) positive peripheral T cell lymphoma (PTCL).

Methods—An RNase protection assay system was used to study the differential expression of 21 cytokines in parallel in eight cases of EBV positive non-nasal PTCL, and compared with 11 EBV negative non-nasal PTCLs and three EBV positive nasal natural killer (NK) cell lymphomas.

Results—Among the eight EBV positive cases, interferon γ (IFN-γ), lymphotoxin β (LTβ), interleukin 10 (IL-10), tumour necrosis factor α (TNF-α), transforming growth factor β1 (TGF-β1), and IL-1 receptor α (IL-1Ra) were frequently detectable. IL-15, IL-6, IL-4, IL-1β, TGF-β1, and IL-9 were sporadically detectable. Of the frequently detectable cytokines, IFN-γ and LTβ were commonly detected in the EBV negative cases. For cases with > 50% EBV encoded small non-polyadenylated RNA (EBER) positive cells, IL-10, TNF-α, and TGF-β1 were detected in three of three cases, and IL-1Ra in two of three cases. For cases with < 20% EBER positive cases, IL-10 was detected in three of five cases, TNF-α in two of four cases, but TGF-β1 and IL-1Ra were not detected. Interestingly, IL-6 was detected in two of three cases with > 50% EBER positive cells, but only in one of five cases with < 20% EBER positive cells. For comparison, IL-9 was detected in two of five cases with > 50% EBER positive cells, but not at all in cases with < 20% EBER positive cells.

In primary nasal T/natural killer (NK) cell lymphomas, Epstein-Barr virus (EBV) is associated with 100% of cases,1,2 the virus exists as a single clonal episome, and is found in almost all tumour cells.3 In non-nasal peripheral T cell lymphoma (PTCL), EBV is detected in only 52% of cases,3 the virus might be monoclonal or biclonal, and is found only in a proportion of atypical cells. EBV infection and cytokine expression are closely associated in B cells,4 and we speculate that specific cytokines are secreted locally in the tumour environment of EBV positive PTCLs.

In vitro studies on EBV infection suggest that viral survival in lymphoblastoid cells can be maintained largely by cytokine mediated autocrine loops.5 In vivo studies have shown that various cytokine genes can be upregulated in a number of EBV associated malignancies.6–8 Many cytokines are suspected to contribute towards the histological characteristics and clinical behaviour of tumours.6–8 Moreover, large numbers and variable amounts of cytokines are found in various malignant lymphomas.10–14

From the literature, it is clear that the expression of many cytokines is altered in peripheral T cell lymphomas. To date, however, the differential expression of a panel of cytokines in EBV positive peripheral T cell lymphomas has not been reported. Therefore, it is not clear whether EBV infection is associated with upregulation of cytokines in the microenvironment of PTCLs. We have examined cytokine expression in EBV positive non-nasal peripheral T cell lymphomas, compared with EBV negative non-nasal PTCLs, as well as with nasal NK cell lymphoma. The results obtained should give an insight into the factors influencing viral survival in the tumour environment of peripheral T cell lymphomas.

Table 1 Peripheral T cell lymphoma (PTCL) and natural killer cell lymphoma (NL)

<table>
<thead>
<tr>
<th>PTCL</th>
<th>EBV positive</th>
<th>EBV negative</th>
<th>NL EBV positive</th>
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<tbody>
<tr>
<td>Frequently detectable cytokines</td>
<td>IFN-γ 8/8, LTβ 7/7, IL-10 6/8, TNF-α 5/7, TGF-β1 4/7, IL-1Ra 4/8</td>
<td>10/11, 9/10, 5/11, 5/10, 9/10, 9/11</td>
<td>3/3, 2/2, 3/3, 2/2, 0/2, 2/2</td>
</tr>
<tr>
<td>Sporadically detectable cytokines</td>
<td>IL-15 3/7, IL-6 3/8, IL-4 3/8, IL-10 2/8, TGF-β1 0/7, IL-9 0/7</td>
<td>3/10, 5/10, 1/11, 4/10, 4/10, 2/10</td>
<td>3/3, 3/3, 0/3, 0/2, 0/2, 0/3</td>
</tr>
</tbody>
</table>

Frequently detectable cytokines are detected in 50% or more cases of Epstein-Barr virus (EBV) positive PTCLs. Sporadically detectable cytokines are detected in less than 50% of cases of EBV positive PTCLs. IFN-γ, interferon γ; IL, interleukin; IL-1Ra, IL-1 receptor α; LTβ, lymphotoxin β; TGF-β1, transforming growth factor β1; TNF, tumour necrosis factor.
Materials and methods

Table 2  Summary of mRNA expression of EBER by ISH; LMP1 and EBNA2 by RT–PCR; and the detectable cytokines by RNase protection assay in Epstein-Barr virus (EBV) positive peripheral T cell lymphomas (PTCLs) of different subtypes

<table>
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<th>Subtype</th>
<th>AIL</th>
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<th>MF</th>
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<th>AIL</th>
<th>AIL</th>
<th>LEL</th>
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<tbody>
<tr>
<td>EBER</td>
<td>&gt;50%#</td>
<td>&gt;50%*</td>
<td>&lt;50%</td>
<td>&lt;50%</td>
<td>&lt;50%</td>
<td>&lt;50%</td>
<td>&lt;50%</td>
<td>&lt;50%</td>
</tr>
<tr>
<td>LMP1</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
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<tr>
<td>EBNA2</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>IFN-γ</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
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<td>–</td>
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<tr>
<td>IL-10</td>
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<td>+</td>
<td>+</td>
<td>+</td>
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<td>–</td>
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<td>TNF-α</td>
<td>+</td>
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<tr>
<td>IL-1Ra</td>
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<td>+</td>
<td>+</td>
<td>–</td>
<td>–</td>
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<tr>
<td>IL-15</td>
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<td>–</td>
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<td>IL-4</td>
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<td>–</td>
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<tr>
<td>IL-1β</td>
<td>–</td>
<td>–</td>
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<td>–</td>
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# EBER was positive in >50% of the atypical cells; *EBER was positive in <20% of the atypical cells.

AIL, angioimmunoblastic T cell lymphoma; EBER, EBV encoded small non-polyadenylated RNAs; EBNA2, EBV nuclear antigen 2; IFN-γ, interferon γ; IL, interleukin; IL-1Ra, IL-1 receptor 1; ISH, in situ hybridisation; LEL, lymphoepithelioid lymphoma; LT, lymphotoxin β; MF, mycosis fungoides; ND, not determined; PL, pleomorphic medium size and large T cell lymphoma; RT–PCR, reverse transcription–polymerase chain reaction; TGF-β, transforming growth factor β1; TNF-α, tumour necrosis factor α.

Figure 1  Detection of Epstein-Barr virus (EBV) encoded small non-polyadenylated RNA (EBER) 1 and 2 by in situ hybridisation (ISH) using a fluorescein isothiocyanate (FITC) labelled probe. (A) EBER was positive in >50% of atypical cells including blast like cells. (B) EBER was only positive in a subpopulation (up to 20%) of atypical cells including blast like cells. (C) LMP1 protein was positive in isolated cells. (D) Interleukin 10 (IL-10) protein was positive in many cells showing cytoplasmic staining.
was carried out as described previously. The primers and probes used have also been described previously.

Three template sets were used for RPA (Pharmingen, San Diego, USA). Each set contains DNA templates that can be used for the T7 polymerase directed synthesis of 32P labelled antisense RNA probes that can in turn hybridise with target human mRNA encoding IL-5, IL-4, IL-10, IL-14, IL-15, IL-9, IL-2, IL-13, and interferon γ (IFN-γ) (hCK-1); IL-12p35, IL-12p40, IL-10, IL-1α, IL-1β, IL-1 receptor α (IL-1Ra), IL-6, and IFN-γ (hCK-2); TNF-β, lymphotoxin β (LTβ), tumour necrosis factor α (TNF-α), IFN-γ, IFN-β, transforming growth factor β3 (TGF-β3), TGF-β2, and TGF-β1 (hCK-3). These assays were carried out essentially according to the manufacturer’s instructions.

Results

Table 1 summarises the expression of the cytokines. Table 2 summarises the histological subtypes, expression of EBV, and cytokine gene transcripts for the eight EBV positive cases. Among the eight EBV positive cases, three cases showed > 50% EBER positive cells (fig

Figure 2  Epstein-Barr virus (EBV) nuclear antigen 2 (EBNA2) and EBV latent membrane protein 1 (LMP1) mRNA expression analysis for the eight EBV positive peripheral T cell lymphomas (PTCLs) by reverse transcription–polymerase chain reaction (RT–PCR) followed by Southern blot hybridisation. Whereas EBNA2 mRNA was detected in cases 3 and 8 only, LMP1 mRNA was detected in all cases. The B95.8 cell line was used as a positive control.

Figure 3  Multiprobe RNase protection assay for the detection of cytokines using hCK-1, hCK-2, and hCK-3 probe sets. Lane 1 shows the unprotected probe sets. Lane 2 shows the corresponding RNase protected probes after hybridisation with the total RNA (2 µg; provided by PharMingen) as positive control. Lane 3 shows the corresponding RNase protected probes after hybridisation with total RNA (2 µg) from a case of Epstein-Barr virus (EBV) positive angioimmunoblastic T cell lymphoma (AIL). The negative control using yeast tRNA (2 µg) is not shown. Note that each probe band (lane 1) migrates more slowly than its protected bands (lanes 2 and 3); this is the result of flanking sequences in the probe that are not protected by mRNA. Because of the differences in the expression of different cytokines, the exposure time for each lane varied, resulting also in variations in the intensity of the housekeeping gene L32. The undigested probe sets were also loaded (typically 1000–2000 counts/minute/lane) as markers. IFN, interferon; IL, interleukin; IL-1Ra, IL-1 receptor; LTβ, lymphotoxin β; TGF, transforming growth factor; TNF, tumour necrosis factor.
Table 3  Expression of LMP1, EBNA2, and IL-10 (as measured by immunohistochemistry) in Epstein-Barr virus (EBV) positive peripheral T cell lymphomas

<table>
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<tbody>
<tr>
<td>LMP1</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
<td>+</td>
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<tr>
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<td>+</td>
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</table>

AIL, angioimmunoblastic T cell lymphoma; EBNA2, EBV nuclear antigen 2; IL-10, interleukin 10; LEL, lymphoepithelioid lymphoma; MF, mycosis fungoides; PL, pleomorphic medium size and large T cell lymphoma.

1A) and five cases showed < 20% EBER positive cells (fig 1B). The three nasal cases showed EBER positive cells in more than 70% of the atypical cells. All eight cases showed LMP1 mRNA expression, and two cases showed EBNA2 mRNA expression (fig 2). However, using immunohistochemical staining, LMP1 was detected only in isolated cells (fig 1C), and EBNA2 was not detected at all.

Figure 3 illustrates the results obtained from the RPA system. Each set of RPA analysis requires a minimum of 5 µg of total RNA, so that each analysis could only be repeated once because of the limited amounts of tissue available for our study. The x ray film was given maximum exposure to avoid underdetection.

Our aim was to identify the presence or absence of the cytokines in the tumour environment. Overall, 12 of 21 cytokines showed detectable levels of expression. Nine of 21 cytokines, namely IL-5, IL-14, IL-2, IL-12p35, IL-12p40, IL-1α, IFN-γ, TGF-β3, and TGF-β2 were not detected by RPA; either their expression level was below the detection limit or they were not expressed.

NON-NASAL PTCL
Frequently detectable cytokines
As shown in Tables 1 and 2, irrespective of the presence of EBV, IFN-γ and LTβ were detectable in almost all cases. In cases with > 50% EBER positive cells, IL-10, TNF-α, and TGF-β1 were uniformly detectable (three of three cases), and IL-1Ra was frequently detectable (two of three cases). In cases with < 20% EBER positive cells, only IL-10 was preferentially detectable (three of five cases). Overall, IL-10 and TNF-α were marginally more frequently detected in EBV positive cases compared with EBV negative cases. At the cellular level, the IL-10 protein was detected by immunohistochemistry in the cytoplasm of many cells (fig 1D). Table 3 compares the production of the IL-10 protein with the EBV proteins, namely LMP1 and EBNA2.

Sporadically detectable cytokines
These cytokines are listed in table 1. However, in cases with > 50% EBER positive cells, IL-6 was frequently detectable (two of three cases). In contrast, in cases with < 20% EBER positive cells, IL-6 was rarely detectable (table 2). In EBV negative cases, most of these cytokines were detected sporadically (table 1).

NK CELL LYMPHOMA
IFN-γ, LTβ, IL-10, TNF-α, IL-1Ra, IL-15, and IL-6 were detectable in every tested case of EBV positive NK cell lymphoma. In PTCL cases with > 50% EBER positive cells, TGF-β1 was not detected at all, whereas IL-6 and IL-1Ra were detected in all cases.

Discussion
The study of cytokine expression by means of RPA indicated that cytokines can be differentiated into two sets: those cytokines that are associated directly with the presence of EBV and those that are not.

Irrespective of EBV infection, IFN-γ and LTβ were commonly detectable in all cases. Together with most of the sporadically detectable cytokines, namely IL-15, IL-4, IL-1β, TNF-β, and IL-9 (table 1), these cytokines do not correlate specifically with EBV infection. Instead, the expression of many of these cytokines correlated well with the tumour entities.12 21–23

TGF-β1 was detectable in all three cases with > 50% EBER positive cells. This potent immunosuppressive agent has been detected before in the tumour cells of PTCLs,24 but no reference was made to the presence of EBV. Its absence in other EBV positive PTCLs and all three cases of EBV positive NK cell lymphoma suggest that this cytokine might not play a direct and primary role in EBV infection. However, it might have a cooperative role in PTCLs that harbour high numbers of EBV.25–26 Similarly, IL-1Ra and IL-6 were detectable in two of three cases with > 50% EBER positive cells. Unlike TGF-β1, they were detectable in every cases of EBV positive NK cell lymphoma tested. Such a correlation suggests that these two cytokines are associated only indirectly with both T and NK cell tumours that harbour high numbers of EBV.27

Others have reported IL-10 expression in PTCLs to be subtype specific rather than associated with EBV;28 our result did not show the same trend. Among the six frequently detectable cytokines, IL-10 and TNF-α showed marginal preferential expression in the eight EBV positive PTCLs. They were also detected in all cases of EBV positive NK cell lymphoma. These two cytokines did not appear to be lymphoma subtype specific.

IL-10 is a pleiotropic cytokine. It is a potent immunosuppressor of cytotoxic T cells and a costimulator of B cell growth. Presented to NK cells, IL-10 is found to mediate IL-2 induced NK production of IFN-γ.29 IL-10 has been associated with many EBV associated malignancies, such as Hodgkin’s disease and Burkitt’s lymphoma, in which it is thought to mediate immunosuppression in the microenvironment.7–8

EBV gene transcript studies of the eight cases revealed that only isolated cells were LMP1 positive, irrespective of the percentage of EBER infected cells, indicating mixed latency I and II EBV positive cells. The detection of EBNA2 at the mRNA level, but not at the protein level, suggests that there might be rare latency III cells in the tumour cell population of the two cases. Therefore, the latency pattern appears to be similar to that of EBV positive NK cell lymphoma, the EBV gene expression pattern of which has been studied.
In contrast to the isolated detection of LMP1 positive cells, immunostaining revealed, at the cellular level, that many cells stained positive for IL-10, irrespective of the percentage of EBER positive cells. The percentage of IL-10 positive cells did not appear to correlate with the percentage of EBER positive cells. In vitro studies on B cell lines suggest that IL-10 expression might be transactivated by LMP1, but this has not been studied in T cell lines or T cell tumours; therefore, the effect of EBV proteins and IL-10 on each other in T cells is not clear. We suspect that IL-10 was already present in the tumour environment upon viral infection, but further studies are needed to clarify whether or not EBV had a sustaining or enhancing effect on IL-10 expression. Moreover, our results indicate that there is no correlation between IL-10 and EBNAA2 expression. However, this also needs to be clarified further in future in vitro studies.

Comparing the two immunosuppressive agents, TGF-β1 was detected more frequently in EBV negative cases, whereas IL-10 was detected more often in EBV positive cases. Interestingly, in cases with >50% EBER positive cells, both IL-10 and TGF-β1 were expressed. In contrast, in the highly EBV associated NK cell lymphomas, IL-10 was detectable, but TGF-β1 was not (table 1). Thus, EBV infection correlates with IL-10 expression both in non-nasal PTCL and NK cell lymphoma, and local immunosuppression in EBV positive cases might be the result of IL-10 rather than TGF-β1 expression. The coexpression of TGF-β1 and IL-10 is unique in cases with >50% EBER positive cells, so their effect on immunosuppression in those cases will require further study.

TNF-α is also a pleiotropic cytokine, and it can be produced by B and T cells, as well as NK cells. EBV infection of B cells promotes its expression, and it has been detected in EBV positive B immunoblasts in AILD cases. Recent studies have shown that EBV positive T cells also upregulate TNF-α expression. Further, EBV selectively upregulated TNF-α expression over IFN-γ and IL-1α. TNF-α and TGF-β are known to be functionally redundant cytokines in many aspects. Interestingly, our study shows that in the EBV negative cases, the frequency of detection of TNF-α and TGF-β was similar. In EBV positive cases, however, there was a preferential upregulation of TNF-α and downregulation of TGF-β (table 1). In comparison, TNF-α is also detected selectively in EBV positive NK cell lymphoma. This is interesting because in normal NK cells, upregulation of TNF-α induced apoptosis.

In summary, among the 21 cytokines studied, we noted a trend for the preferential expression of IL-10 and TNF-α in EBV positive non-nasal PTCL. This finding correlates strongly with a similar survey on EBV immortalized B cell lines. Thus, in EBV positive non-nasal PTCL, the strongest candidates associated directly with EBV infection are IL-10 and TNF-α. Furthermore, at the cellular level, as shown by protein staining, the detection of more IL-10 positive cells compared with EBER and LMP1 suggests that such cytokine environments might enhance EBV infection and contribute towards tumorigenesis.

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