<table>
<thead>
<tr>
<th><strong>Title</strong></th>
<th>Trichinella spiralis: specificity of ES antigens from pre-encysted larvae</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Author(s)</strong></td>
<td>Ko, RCC; Wong, TP</td>
</tr>
<tr>
<td><strong>Citation</strong></td>
<td>Journal of Helminthology, 1992, v. 66 n. 1, p. 38-44</td>
</tr>
<tr>
<td><strong>Issued Date</strong></td>
<td>1992</td>
</tr>
<tr>
<td><strong>URL</strong></td>
<td><a href="http://hdl.handle.net/10722/45434">http://hdl.handle.net/10722/45434</a></td>
</tr>
<tr>
<td><strong>Rights</strong></td>
<td>This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License.; Journal of Helminthology. Copyright © Cambridge University Press.</td>
</tr>
</tbody>
</table>
Trichinella spiralis: specificity of ES antigens from pre-encysted larvae

R. C. KO and T. P. WONG

Department of Zoology, University of Hong Kong, Hong Kong

ABSTRACT
Excretory/secretory (ES) antigens were obtained by culturing pre-encysted Trichinella spiralis larvae which were recovered from muscles of experimentally infected mice 14–15 days postinfection. Analyses of these antigens (PEL ES) with immunoblotting, SDS-PAGE and Triple Antibody ELISA showed that they yielded a low sensitivity and specificity when tested with antisera against the common nematodes of Chinese pigs. As compared to ES antigens from encysted larvae, PEL ES also contained more low molecular mass proteins.

KEY WORDS: Trichinella spiralis, pre-encysted larvae, ES antigens, specificity, Nematoda, immunology

INTRODUCTION
It is well known that excretory/secretory (ES) antigens are more specific than crude somatic antigens in the immunodiagnosis of many nematodes. GAMBLE et al. (1988) observed that ES antigens obtained from encysted muscle larvae of Trichinella spiralis after 24 h in culture were highly specific when they were applied to screen pigs for trichinellosis in Yugoslavia. Ko & Yeung (1989) noted a similar specificity when the antigens were used to detect antibodies in Chinese pigs. Antigenic epitopes unique to T. spiralis were found in proteins having molecular mass 45–53 kDa. Besides in vitro culture, the specific antigens were also successfully isolated from crude worm extracts by the rotating ampholine horizontal method (Ko & Yeung, 1991). However, Ko & Yeung (1989) discovered that ES products from adult worms yielded markedly weaker reactions when tested against homologous anti-pig sera using Triple Antibody ELISA and immunoblotting. False positive reactions with anti-Metastrongylus apri serum were also observed. Adult ES antigens consisted of epitopes different from those of muscle larvae.

Therefore, it appears that ES products released by T. spiralis during various stages of development exhibit different antigenicity. Presumably the first-stage larvae, which have an exceptional growth rate, will actively release ES materials soon after entering muscle cells. The larvae can grow from about 100 µm to 1 mm in length in less than three weeks. During the intracellular development of the worms, the muscle fibres will undergo re-organization to form “nurse cells”. Recent studies by Lee et al. (1991) and Despommier et al. (1990) have demonstrated the presence of ES antigens in some of the hypertrophic nuclei of these specialized cells. The authors suggested that the secreted antigens may play a role in the modulation of the host genomic expression.

The present study was undertaken to determine whether the ES antigens obtained from pre-encysted larvae (PEL ES) were antigenically different from those of the encysted worms (EL ES). The usefulness of PEL ES antigens in serodiagnosis was also examined.

Animals
The strain of T. spiralis found in Canada and has since been used as the protoscolex which can be placed in 0.8% NaCl and then placed in the subcutaneous tissue of mice. The larvae were collected and placed in 200 RPM and 4°C in a centrifuge for five minutes. The supernatant was checked under an inverted culture flask.

Positive control sera included those from six albino New Zealand rabbits, one of which was about 20 larvae/g of control. Hyperimmune (Trichuris suis, Metastrongylus apri) sera were procured from the weekly intake of incomplete adjuvants.

Antigens
Crude somatic antigens were obtained from the pre-encysted larvae as described by Ko & Yeung (1989) which were collected from pigs in Hong Kong, were also prepared for use. ES antigens were obtained by culturing T. spiralis in RPMI 1640 medium at 24 h in an incubator with an infrared lamp at 37°C. The culture solution was filter sterilized by a sample containing 100 µl of gel filtration and then pooled. The pooled samples were assayed using a complement fixation assay.

After 24 h in culture, the ES antigens were harvested and assayed using a complement fixation assay. The assay was performed according to the manufacturer's instructions.
of ES antigens encysted larvae
KO and T. P. WONG
Hong Kong, Hong Kong

Trichinella spiralis larvae vs postinfection. Analyses by ELISA showed that the common nematodes of also contained more low specificity, Nematoda,

...specific than crude loses. GAMBLE et al. d muscle larvae of en they were applied 1989) noted a similar ies in Chinese pigs. ins having molecular antigens were also rotating ampholine) & YEUNG (1989) dly weaker reactions antibody ELISA and ylus api serum were different from those of spiralis during various mably the first-stage release ES materials about 100 µm to 1 mm development of the form “nurse cells”. et al. (1990) have hypertrophic nuclei of 1 antigens may play a her the ES antigens ically different from PEL ES antigens in

MATERIALS AND METHODS

Animals
The strain of T. spiralis used was originally isolated from a pig in Ontario, Canada and has since been maintained in both Wistar albino rats and ICR mice. Encysted larvae were recovered from mice 30 days postinfection by the standard pepsin digestion method. Pre-encysted larvae were recovered from the muscles of mice 14–15 days postinfection. The infected muscles were cut into fine pieces and then placed in 0.8% NaCl solution in a Baermann’s funnel at room temperature. A lamp was placed near the funnel stem to provide a heat gradient. After 4–5 h, the larvae were collected and they were concentrated by centrifugation for five minutes at 2000 RPM and 4°C in a refrigerated centrifuge (Beckman GPR, table top model) for five minutes. The worms were washed five times in sterile saline and then checked under an inverted microscope for viability before being transferred to culture flasks.

Positive control serum samples were obtained by orally infecting five weaned piglets and six albino New Zealand rabbits with 10,000 each. The animals were bled and killed on the same day after infection. The final worm burden of the pigs was about 20 larvae/g of muscle. Seven uninoculated piglets served as the negative control. Hyperimmune serum against the common parasites of Chinese pigs (Trichuris suis, Metastrongyulus api, Gnathostoma hispidum and cysticerci of Taenia solium) were produced in nine piglets and 11 rabbits by three intramuscular injections at weekly intervals using crude antigens and Freund’s complete and incomplete adjuvants.

Antigens
Crude somatic antigens of T. spiralis were prepared by sonication of muscle larvae as described by KO & YEUNG (1989). Crude antigens of heterologous species which were collected from pigs slaughtered in the Kennedy Town Abattoir, Hong Kong, were also prepared in a similar manner.

ES antigens were obtained by maintaining encysted and pre-encysted worms for 24 h in RPMI 1640 medium in an auto-flow microprocessor-controlled CO2 incubator with an infrared sensor (Nuaire). The maintenance methods basically followed those of KO & YEUNG (1989) and GAMBLE et al. (1983). For the culture of pre-encysted larvae, about 1,000 worms were maintained in each cantel neck 25 cm² culture flask (Costar). ES antigens were concentrated from the supernatant solution by a sample concentrator (Speed-Vac). The concentrated samples were desalted by gel filtration using Bio-Gel P6DG (Bio-Rad). Protein concentrations of the pooled samples were determined by a Protein Assay Kit (Bio-Rad) and the absorbance of the samples was read at 594 nm with a spectrophotometer (Milton Roy).

After 24 h in culture, usually less than 5% of the worms would appear inactivated or dead. Therefore, most antigens recovered from the culture medium were primarily ES products rather than somatic antigens released by dying worms (see GAMBLE et al., 1988).

Sodium dodecyl sulphate-polyacrylamide gel electrophoresis, immunoblotting and enzyme-linked immunosorbent assays
The details of these methods have been published elsewhere (KO & YEUNG, 1989). Sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) was performed according to LAEMMLI (1970) in a Mini-Protean II cell (Bio-Rad).
4 µg protein was electrophoresed in 10% gel at 100V for 1.5 h. The gels were stained with a silver staining kit (Pharmacia). A computer-controlled laser densitometer (Ultralab XL, LKB) was used to scan the stained gel. For molecular mass determinations, Dalton Mark VIII (Sigma) was used.

Enzyme-linked immunosorbent blotting (EITB) was performed on nitrocellulose paper (0-45 µm, Schleicher & Schuell), following the method of Towbin et al. (1979). A mini-transblot cell (Bio-Rad) was used and the free binding sites were saturated with 3% gelatin in TBS. The anti-rabbit IgG horse-radish peroxidase conjugate (affinity purified, Sigma) was diluted to 1:2000 (v/v). 4-chloro-1-naphthol was used as the substrate. Biotinylated SDS-PAGE standards (Bio-Rad) were used for molecular mass determinations.

The Triple Antibody ELISA was performed according to Ko & Yeung (1989) and Gamble et al. (1983). Chequer-board titrations were used to determine the optimal conditions of the assay. Flat bottom polystyrene plates (Linbro, Flow Lab) were coated overnight at 4°C with antigens (0.1 µg PEL ES, 0.2 µg EL ES, 0.4 µg crude antigens in 100 µl/well) diluted in coating buffer (0.1 M carbonate-bicarbonate buffer, pH 9.6). Coated plates were washed three times with 0.9% NaCl and 0.05% Tween 20 using an automatic washer (Ultrawash, Dynatech) before the addition of the test serum (100 µl/well). After incubation for 2 h at 37°C and then washing, rabbit anti-porcine IgG (1:30 000 for crude and ES, 1:25 000 for PEL ES) was added. After a further incubation at room temperature for 2 h, the plates were washed before the addition of 100 µl goat anti-rabbit IgG horse-radish peroxidase conjugate (Sigma) (1:30000 for crude and ES, 1:20000 for PEL ES). The plates were incubated overnight at 4°C. They were washed once more before the addition of ortho-phenylenediamine (OPD). The absorbance was read by an automatic ELISA reader (MR 710, Dynatech) at 490 nm (test filter) and 410 nm (reference filter). Absorbance values greater than 5 × the mean OD of the normal pool were considered as positive.

RESULTS

SDS-PAGE

Using a non-gradient gel, the SDS-PAGE profiles of the PEL ES, EL ES and crude antigens were shown to be different (Fig. 1). The PEL ES antigens were consisted of at least 20 bands; the molecular mass of the prominent bands were at 15–20 kDa. The bands at 45, 47 and 53 kDa were weakly stained. However, these bands which contained the epitope specific to T. spiralis were distinctive in the EL ES antigens which were also consisted of more proteins with larger molecular mass.

EITB

EITB was carried out with crude antigens and ES antigens from both pre-encysted and encysted worms. Various antiserum against common parasites of Chinese pigs were used as developing sera (Fig. 2).

Lanes 2–7 show the blotting results of PEL ES. No bands were observed when anti- G. hispidum (lane 6) and M. apri (lane 7) sera were used as developing sera. Weakly stained bands were observed against the following antiserum: four at 15, 18, 45 and 56 kDa against anti-T. spiralis serum (lane 3), one at 15 kDa against anti-T. suis (lane 4) and one at 56 kDa against anti-C. cellulosae (lane 5).

Lanes 8–19 show the blotting results of ES antigens from encysted worms and crude worm extracts. Intensively stained bands were present at 45, 47 and 53 kDa when anti-T. spiralis and 10). Three faint from encysted larva were shown when c.

FIG. 1. SDS-PAGE el (PEL) and encysted larv kDa). Proteins with spe marker.

ELISA

Sera samples, w and 30 postinfectior and PEL ES antig +ve/OD –ve ratio antigens, 5–9, 12: 1

Fig. 3 compares against pig anti-T. 2 excess condition. F maximum at 12 µg/v the maximum at 1

Fig. 4 compares against various pig produced a substa
or 1.5 h. The gels were
puter-controlled laser
ined gel. For molecular
ed.
formed on nitro-
method of Towner et
free binding sites were
horse-radish peroxidase
2000 (v/v). 4-chloro-1-
E standards (Bio-Rad)
to Ko & Yeung (1989)
used to determine the
ates (Linbro, Flow Lab)
S, 0.2 µg EL ES, 0.4 µg
fer (0.1 M carbonate-
three times with 0.9%-
(Ultrawash, Dynatech)
cubation for 2 h at 37°C
ide and ES, 1:25 000 for
temperature for 2 h, the
rabbit IgG horse-radish
, 1:20 000 for PEL ES).
shed once more before
rance was read by an
(test filter) and 410 nm
mean OD of the normal

![SDS-PAGE electrophoresis using non-gradient gel to compare ES antigens from pre-encysted (PEL) and encysted larvae (EL).](image)

**FIG. 1.** SDS-PAGE electrophoresis using non-gradient gel to compare ES antigens from pre-encysted (PEL) and encysted larvae (EL). PEL antigens were dominated by low molecular mass proteins (15–20 kDa). Proteins with specific epitopes (45, 47, 53 kDa) were prominent in EL ES. M=molecular mass marker.

when anti- *T. spiralis* serum was used to blot both EL ES and crude antigens (lane 8 and 10). Three faint bands at 59, 41, 33 kDa were observed in lane 14 ES antigens from encysted larvae and anti-*T. suis* were used. However, at least 10 strong bands were shown when crude antigens were blotted with the same antiserum (lane 18). The crude antigens also cross-reacted with anti- *G. spinigerum* and *C. cellulosae* serum (lanes 16, 17).

**ELISA**

Sera samples, which were taken from experimentally infected mice on days 14 and 30 postinfection, were tested by Triple Antibody ELISA against crude, EL ES and PEL ES antigens. PEL ES antigens yielded negative reactions. The OD +ve/OD –ve ratios were as follows: for crude antigens, 4.6, 8.7; for EL ES antigens, 5.9, 12; for PEL ES antigens, 0.49, 0.79.

Fig. 3 compares the sensitivity of the different antigen when they were tested against pig anti-*T. spiralis* serum. Both the conjugates and substrates were under excess condition. For PEL ES antigens, the increase in OD values reaches the maximum at 12 µg/ml whereas for EL ES and crude antigens, the increase reaches the maximum at 1 µg/ml and 12 µg/ml respectively.

Fig. 4 compares the specificity of the different antigens when they were tested against various pig antisera. PEL ES antigens exhibited a low specificity. They produced a substantially low OD+ve/OD–ve ratio when tested against the
homologous anti-\textit{T. spiralis} serum; the ratio was below the 5× mean OD−ve of control samples, i.e. a false negative reaction. However, they yielded a false positive reaction with anti-\textit{M. apri} serum. As compared to EL ES antigens, PEL ES antigens yielded higher OD+ve/OD−ve ratios against anti-\textit{G. hispidum} and \textit{C. cellulosae} sera.

FIG. 2. Enzyme-linked immunotransfer blot of crude and ES antigens from pre-encysted (PEL) and encysted larvae (EL), using homologous and heterologous rabbit antisera as developing serum. Molecular mass marker: lanes 1 and 20. PEL ES antigens: lanes 2-7. EL ES antigens: lanes 8-9 and 11-14. Crude antigens: lanes 10 and 15-19. The following antisera were used: normal pig serum (N), \textit{Trichinella spiralis} (Ts), \textit{Trichuris suis} (Tr), \textit{Cysticercus cellulosae} (C), \textit{Gnathostoma hispidum} (G), \textit{Metastrongylus apri} (M). Arrows indicate positions of weakly stained bands.

FIG. 3. Comparison of worm extract (CWE) at 2.0 OD, using pig anti-\textit{Trichinella spiralis} antisera.

FIG. 4. The OD+ve/OD−ve ratio when crude and ES anti-\textit{Gnathostoma hispidum} sera are used.

Data in the pre- and encysted larva sera low molecular stained bands were not found in any sera, homologous and heterologous antisera productions. For anti-\textit{M. apri} serum.
DISCUSSION

Data in the present study demonstrate that the ES products from pre-encysted and encysted larvae are antigenically different. The former, which are dominated by low molecular mass proteins, have a low sensitivity and specificity. Only weakly stained bands were obtained in the blotting with anti-\textit{T. spiralis} serum. These antigens also invariably yielded low OD positive to negative ratios when tested against homologous antiserum by the Triple Antibody ELISA. The lack of specificity of the antigens was indicated by the strong false positive reaction against anti-\textit{M. apri} serum. EITB also showed some cross-reactivities with anti-\textit{T. suis} and
C. cellulosae sera. Therefore, the PEL ES antigens are unsuitable as serodiagnostic antigens. The lack of specificity is probably due to the fact that the immunodominant 45–53 kDa proteins of the EL ES antigens are only present in small quantities in PEL ES antigens. Gamble & Graham (1984) reported that the antigenic epitope unique to T. spiralis was recognized by an IgM monoclonal antibody (7C2C3). However, our studies have shown that the epitope could also be recognized by two IgG, as well as IgM monoclonal antibodies (unpublished data).

The antigenic profile of PEL ES products seems to resemble that of ES antigens obtained from adult worms as reported in an earlier study (Ko & Yeung, 1989). The adult worm ES antigens were found to yield weaker reactions in Triple Antibody ELISA. They were rich in low molecular mass proteins and lacked the specific 45–53 kDa bands (as demonstrated by EITB). A strong false positive reaction with anti-M. apri serum was also observed. But, as compared to PEL ES antigens, those from the adult worms contained more high molecular mass proteins. Therefore, it appears that the composition of the ES products released by T. spiralis will vary according to the stage of worm development in the host.

The specific proteins are probably secreted by the stichosome of this trichurid nematode in substantial quantities during the active phase of encystment. The transformation of the striated muscle fibre of the mammalian host into a nurse cell with hypertrophic nuclei is completed by 20–30 days postinfection. In a recent study using immunocytochemical methods and monoclonal antibodies as probes, we observed that some hypertrophic nuclei of nurse cells at 30 days postinfection were stained positively for ES antigens. It was suggested that only some molecules in the ES pool can enter the nuclei or different monoclonal antibodies may have different affinities to the subunits of the glycoproteins which can enter the nuclei (Lee et al., 1991).

ACKNOWLEDGEMENT
This project was generously supported by the Croucher Foundation, Hong Kong.

REFERENCES

Accepted 11th September, 1991.