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An update on the aetiology of orofacial clefts

Objective. To review recent data on the aetiology of cleft lip and palate.


Study selection. Literature and data on aetiology of cleft lip and palate using the following key words: ‘cleft lip’, ‘cleft palate’, ‘aetiology’, and ‘genetics’.

Data extraction. Relevant information and data were reviewed by the authors.

Data synthesis. Cleft lip and palate is one of the most common types of congenital malformation. The aetiology seems complex, but genetics plays a major role. Recently several genes causing syndromic cleft lip and palate have been discovered. Three of them—namely T-box transcription factor-22 (TBX22), poliovirus receptor like-1 (PVRL1), and interferon regulatory factor-6 (IRF6)—are responsible for causing X-linked cleft palate, cleft lip/palate-ectodermal dysplasia syndrome, and Van der Woude’s and popliteal pterygium syndromes, respectively; they are also implied in non-syndromic cleft lip and palate. The nature and function of these genes vary widely, illustrating high vulnerability within the craniofacial developmental pathways. The aetiological complexity of non-syndromic cleft lip and palate is also exemplified by the large number of candidate genes and loci.

Conclusions. The aetiology of non-syndromic cleft lip and palate is still largely unknown, but mutations in candidate genes have already been identified in a small proportion of cases of non-syndromic cleft lip and palate. Determining the relative risk of cleft lip and palate, on the basis of genetic background and environmental influence, including smoking, alcohol use, and dietary factors, will aid in genetic counselling and the development of future preventive measures.

Introduction

Cleft lip and palate (CLP) comprises a group of the most common congenital malformations. Immediately after birth, individuals with CLP have facial deformation, feeding problems, and frequent middle ear infection, the treatments of which require interventions from multiple disciplines. At the age of speech...
acquisition, speech therapy is often needed to correct problems resulting from muscular defects of the cleft. As the individual continues to grow, defects in tooth development and malocclusion require dental and sometimes surgical treatment. The lengthy series of treatment from birth to adulthood is a heavy burden for the patient, family, and society. Various efforts have been made to understand the aetiology of CLP so as to predict its occurrence and to prevent it from occurring in the future. In recent years, advances in genetics and molecular biology have begun to reveal the basis of craniofacial development, and a number of genes associated with CLP have been identified. Studies of the combined genetic and environmental cause of CLP are also growing. This article reviews the recently discovered genes involved in CLP, and provides an update on the aetiological factors underlying this common malformation.

**Embryology**

The lip and palatal regions are developed from the embryonic primary and secondary palates. The primary palate contributes to the lip, anterior tooth-bearing alveolus, and the anterior palate up to the incisal foramen. The secondary palate contributes to the remaining hard and soft palates. During the fourth week of gestation, neural crest cells from the anterior neural tube migrate to form the facial primordia, from which the nasal and the lateral maxillary processes fuse to form the primary palate. The secondary palate begins to form in the sixth week of gestation. Palatal shelves initially appear as two downward extensions from the inner side of the left and right maxilla along the lateral surface of the tongue. In the ninth week, the two palatal shelves undergo a rapid horizontal transformation, by moving over the tongue and fusing with each other and with the nasal septum. Failure in the fusion of primary or secondary palates leads to clefts of the respective areas. Thus, CLP appears commonly as cleft lip with or without cleft palate or isolated cleft palate.

Clinically, when CLP appears with other (usually two or more) malformations in recognisable patterns, it is classified as syndromic CLP (SCLP). If it appears as an isolated defect or if syndromes cannot be identified, the term non-syndromic CLP (NSCLP) is used. The number of CLP syndromes is large and still growing. A search of the Online Mendelian Inheritance in Man database using the term ‘cleft lip’ yielded more than 200 entries and a search using ‘cleft palate’ yielded close to 400.1 The distinction between NSCLP and SCLP, however, is sometimes not clear-cut. In families with SCLP, some affected members may present with only CLP, because of variable expression of the syndrome. On the other hand, more than 20% of patients with NSCLP were found to have associated congenital malformations in one study.2 Thus, some cases of SCLP and NSCLP might share a common aetiology.

Recently discovered genes causing orofacial cleft syndromes

**T-box transcription factor-22**

X-linked cleft palate (CPX) is characterised by isolated cleft palate and ankyloglossia (tongue-tie); clinical expression of CPX is highly variable. High-arched palate, bifid uvula, or ankyloglossia could be the only presenting sign in affected males. Female carriers could be asymptomatic or they could express full features of CPX. The syndrome has been found in a number of large families who show inheritance in a Mendelian X-linked semidominant pattern. By using genetic linkage analysis, Stanier et al1 located the disease gene locus to chromosome Xq21. Braybrook et al4 performed extensive mutation analysis of candidate genes in the region and found mutations of the T-box transcription factor-22 gene (TBX22) in a large Icelandic family with CPX and in several smaller families from other countries. Animal experiments showed that expression of TBX22 was highly restricted to the palatal shelves just before their elevation to adopt a horizontal position, and at the base of the tongue corresponding to the frenulum, both of these expression patterns closely matched the clinical presentation of CPX. Involvement of TBX22 in NSCLP has recently been indicated from a genome-wide sibling-pair analyses in which the chromosome Xcen-q region, where TBX22 is located, showed promising multipoint logarithm of odds (LOD) scores.5 Mutation analysis of TBX22 in these patients could reveal whether the gene is involved in NSCLP as well.

**Poliovirus receptor like-1**

Cleft lip/palate ectodermal dysplasia syndrome (CLPED) is characterised by cleft lip with or without cleft palate, hidrotic ectodermal dysplasia, syndactyly, and occasionally mental retardation. Two other syndromes—the Zlotogora-Ogur syndrome and Margarita Island ectodermal dysplasia—are also classified as CLPED. The inheritance of CLPED appears to be autosomal recessive. Using positional cloning, Suzuki et al6 identified mutations of the poliovirus receptor like-1 gene (PVRL1) in CLPED families from Margarita Island, Israel, and Brazil.

The protein product of PVRL1 was initially identified as poliovirus receptor-related protein (PRR). Takahashi et al7 confirmed the function of PRR as a cell adhesion molecule, and they renamed it nectin-1. All three PVRL1 mutations found in families with CLPED resulted in truncations in nectin-1, thereby destroying the nectin-afadin-ponsin (NAP)—dependent cell-adhesion system. In animal experiments, PVRL1 was expressed at the medial edge epithelium of the palatal shelves and the skin surface epithelium—locations that corresponded to the clinical phenotypes of CLPED.8 Interestingly, heterozygous mutation of PVRL1 (W185X) was associated with NSCLP in northern Venezuela.8 Thus, a certain proportion of NSCLP cases can be explained by PVRL1 mutations.
**Interferon regulatory factor-6**

Van der Woude’s syndrome (VDWS) is the most common form of SCLP and accounts for 2% of all CLP cases.9 This syndrome is characterised by cleft lip with or without cleft palate, isolated cleft palate, pits or mucous cysts on the lower lip, and hypodontia. Popliteal pterygium syndrome (PPS) includes all the features of VDWS plus popliteal pterygium, synosthagia, distinct toe/nail abnormality, syndactylly, and genito-urinary malformations. Owing to their clinical similarities, VDWS and PPS were thought to be allelic—that is, caused by different mutations of the same gene.10 Clinical expressions of VDWS and PPS are also highly variable—for example, some family members with VDWS present only with hypodontia, and in PPS, popliteal pterygium is not always present.

The genetic locus for VDWS was localised to chromosome 1 back in 1990.11 Through linkage and chromosomal analysis, the critical area for VDWS was gradually narrowed to 1q32-q41. In 1999, PPS was also linked to the same region.12 In 2002, Kondo et al13 described a pair of monozygotic twins discordant for VDWS whose parents did not have the disorder. The VDWS in the affected twin was thought to arise from somatic mutation. Sequence analysis revealed a point mutation in the interferon regulatory factor-6 gene (IRF6), which is located within the VDWS critical region. Additional mutations of IRF6 were found in 45 unrelated families with VDWS, as well as in 13 families with PPS, thereby confirming a common genetic aetiology in both syndromes.

In animal experiments, IRF6 was expressed in tissues affected by both VDWS and PPS.13 The phenotypic heterogeneity of VDWS and PPS was shown to be due to different types of IRF6 mutation. In most cases of VDWS, IRF6 mutations produced a non-functional protein and haplo-insufficiency. The IRF6 mutations, however, were missense mutations that affected the DNA binding domain and caused a dominant-negative effect, which resulted in severe phenotypes. A partial or modifying role of IRF6 in NSCLP has been demonstrated in a study applying the transmission disequilibrium test, in which specific parental alleles at the VDWS locus were preferentially transmitted to the individuals with NSCLP.14

**Candidate genes or loci for non-syndromic cleft lip and palate**

**Transforming growth factor-alpha**

In 1989, Ardinger et al15 showed in a case-control study that transforming growth factor-alpha (TGFA) was associated with NSCLP. A number of follow-up studies in different populations provided mixed results. Machida et al16 sequenced the TGFA gene in a group of NSCLP patients and found five mutations that could be aetiological to orofacial clefts. The combined effect of TGFA mutation and environmental influence in NSCLP has been analysed by several groups of researchers. The rare TGFA variant (Taql C2 allele) and maternal smoking could increase the risk of cleft palate by 6 to 8 times17 and of cleft lip with or without cleft palate by 2 times.18 If multivitamins were not consumed during the first trimester of pregnancy and the baby is carrying the TGFA Taql C2 allele, the relative risk for cleft lip with or without cleft palate increased by 3 to 8 times.19

**Drosophila msh home box homolog-1**

Mice lacking functional Drosophila msh homeo box homolog-1 (MSX1) develop a cleft of the secondary palate and tooth agenesis.20 In humans, MSX1 mutation was first shown to cause an autosomal dominant form of tooth agenesis.21 Subsequently, van den Boogaard et al22 described a family with a common pattern of tooth agenesis plus a mixture of cleft lip with or without cleft palate and cleft palate. Direct sequencing of MSX1 revealed a disease-causing mutation. Recently, a large-scale sequence analysis of MSX1 performed on 917 CLP patients identified mutations in 16 patients with cleft lip with or without cleft palate, or cleft palate alone, providing evidence that this gene could be involved in both forms of cleft.23 The authors estimated that MSX1 mutations contributed to 2% of all NSCLP cases. A recent study showed that the combined genetic background of rare variants of TGFA and MSX1 could increase the risk of cleft palate by up to 9.7 times, demonstrating the significance of gene-gene interaction in the aetiology of NSCLP.24

**5,10-Methylenetetrahydrofolate reductase**

The association between folic acid deficiency and neural tube defects has been well established. 5,10-Methylenetetrahydrofolate reductase (MTHFR) is the enzyme responsible for catalysing the conversion of 5,10-methylenetetrahydrofolate into 5-methyl-tetrahydrofolate in the folate metabolism pathway. The MTHFR C677T single-nucleotide polymorphism (SNP) is thermally labile and considered a risk factor of neural tube defects.25 In NSCLP, the MTHFR C677T genotype in the mother conferred a risk of CLP in offspring that was increased by 4.6 times.26 In periconceptional folic acid deficiency, the MTHFR thermally labile variant could lead to a risk of CLP that was increased by 10 times.27

**Transforming growth factor beta-3**

Mice lacking functional gene encoding transforming growth factor beta-3 (TGFβ3) displayed cleft palate because of defective adhesion of opposing palatal shelves.28 In humans, TGFβ3 was associated with NSCLP in different populations.24,29 A newly discovered SNP of TGFβ3 (IVS5+104 A>G) increased the risk of CLP by up to 16 times in a Korean population.30

**Other genes and loci**

The special AT-rich sequence-binding protein-2 gene (SATB2), located at chromosome 2q32-33, was disturbed in two unrelated patients with cleft palate.31 Involvement of...
this gene in palatogenesis was confirmed by expression analysis. Although an initial search for SATB2 mutations in 70 patients with cleft palate was negative, the gene could still be responsible for other CLP cases.

Markers from chromosome 4q21 have been linked to familial NSCLP.\(^{32}\) The acyl-coenzyme A desaturase-4 gene (ACOD4) from chromosome 4q21 was recently found disrupted in a family with cleft lip.\(^{33}\) Mutation analysis of ACOD4 in families with 4q-linked CLP or other patients with CLP might reveal more mutations.

Yoshiura et al\(^{34}\) reported on a family with CLP in three generations; all affected members had a balanced translocation at chromosome 19q13. Breakpoint cloning revealed a novel gene termed ‘cleft lip and palate–associated transmembrane protein-1’ (CLPTM1). Eight rare variants of CLPTM1 were found in 74 patients with NSCLP, but none was significantly associated with cleft lip or palate. The authors concluded that CLPTM1 was not a major contributor to CLP. On the other hand, the same region of chromosome 19q13 has been implicated in NSCLP through linkage\(^{35}\) and transmission disequilibrium studies.\(^{36}\) Thus, CLPTM1 or other genes in this locus could still be associated with NSCLP.

Chromosome 6p23 has been indicated in NSCLP by linkage studies.\(^{5,37}\) Chromosomal aberrations involving 6p23 in patients with CLP have also been reported.\(^{38}\) It is therefore likely that a yet unidentified CLP gene exists at chromosome 6p23. A summary of the above genes and loci implied in NSCLP, with chromosomal locations and available evidence, is shown in the Table.

### Environmental factors

#### Smoking

The relationship between maternal smoking and CLP is not strong, but it is significant. Several studies have consistently yielded a relative risk of about 1.3 to 1.5.\(^{19}\) When maternal smoking was considered together with certain genetic background, the combined effect was more significant. Furthermore, van Rooij et al\(^{40}\) found that maternal glutathione s-transferase \( \theta -1 \) (GSTT1) genotype, combined with smoking, could significantly increase the risk of CLP (odds ratio=4.9). And Beaty et al\(^{41}\) reported that maternal smoking and infant MSX1 genotypes contributed to an elevated risk for CLP by 7.16 times.

#### Alcohol use

Heavy maternal drinking, apart from causing foetal alcohol syndrome, increases the risk of CLP. Munger et al\(^{42}\) showed that maternal drinking posted an increased risk from 1.5 to 4.7 times for CLP in a dose-dependent manner. The results were supported by Shaw and Lammer\(^{43}\) that mothers who consumed more than five drinks per occasion had a 3.4 times the risk of CLP developing in the offspring. Low-level alcohol consumption, however, did not seem to increase the risk of orofacial clefts.\(^{44}\) The link between alcohol consumption and genotypes on the risk of CLP has yet to be shown.

#### Use of folic acid and multivitamins

Shaw et al\(^{45}\) reported that if vitamin supplements were not taken during early pregnancy, the risk for CLP could be tripled. Folic acid deficiency with the background of the TGFA TaqI C2 genotype was also found to increase the risk of CLP.\(^{46}\) In addition, defective maternal vitamin-dependent homocysteine metabolism is a risk factor for CLP in offspring; in a case-control study, mothers of patients with CLP had a significantly higher homocysteine level, lower level of whole-blood vitamin B6, and higher rate of hyperhomocysteinemia.\(^{47}\) The role of folic acid supplementation in the prevention of CLP has been investigated in several studies. It seems that low-dose folic acid supplementation by fortifying cereal grain products could not protect against CLP.\(^{48}\) Only a very high dose of supplementary folic acid (10 mg/d) could reduce the risk of CLP significantly (65%).\(^{49}\)
Discussion

The complexity and heterogeneity of CLP, as shown by its extensive involvement in craniofacial syndromes and the number of anticipated candidate genes, could be anticipated from the long developmental duration of the primary and secondary palates. Disruption of the coordinated migration and fusion of various facial processes by genetic, environmental, or combined factors at any timepoint could lead to CLP. Genes, such as those encoding transcription factors (eg TBX22, MSX1), growth factors (eg TGFA, TGFb3), and adhesion molecules (eg PVRL1), have all been implicated in the aetiology of CLP. In the search of genes causing syndromic CLP, current methods of positional cloning or positional candidate approaches could be applied to families or patients. With the completion of the human genome sequence, gene discovery is accelerated by the availability of target sequences. Close collaboration between clinicians and scientists is still essential, as illustrated by the example of VDWS.

The future search for genes in NSCLP will not be straightforward. Mutations in SCLP genes or other candidate genes so far could only be found in a handful of NSCLP cases or families. Promising NSCLP gene candidates, such as MSXI, was expected to cause up to 2% of NSCLP cases.23 The proportion was remarkably similar to that of VDWS in all CLP cases. Therefore, MSXI mutations may represent a new syndrome, which has variable penetrance for the CLP and tooth agenesis phenotypes.

Instead of directly searching for disease-causing mutations in NSCLP, studies exploring the relative risk imparted by candidate genes and gene-environmental interactions are becoming popular approaches, for several reasons. Firstly, the amount of sequence data, such as SNPs, on candidate genes is constantly increasing. These sequence data not only allow more CLP-associated mutations to be identified, but functional correlation may also be attributed to some of these subtle variations. Secondly, high-throughput genotyping such as DNA microarray analyses is now more readily available, so that a large number of candidate genes and SNPs could be tested simultaneously. In the near future, the relative contribution from these candidate CLP genes could be integrated into a genetic test for the weighed risk of CLP. Such data could provide additional information on prospective parents in genetic counselling. Preventive measures, including dietary supplementation or lifestyle modification, could then be prescribed accordingly.

References


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