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<td>Chan, JFW; Zhang, J; Chan, CS; Yip, CY; Mak, WN; Zhu, H; Poon, KM; Tee, KM; Zhu, Z; Cai, J; Tsang, OL; Chik, KKH; Yin, F; Chan, KH; Kok, KH; Jin, D; Au Yeung, KHR; Yuen, KY</td>
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<tr>
<td><strong>Citation</strong></td>
<td>EBioMedicine, 2016, v. 14, p. 112-122</td>
</tr>
<tr>
<td><strong>Issued Date</strong></td>
<td>2016</td>
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<td><strong>URL</strong></td>
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Research Paper

Zika Virus Infection in Dexamethasone-immunosuppressed Mice
Demonstrating Disseminated Infection with Multi-organ Involvement Including Orchitis Effectively Treated by Recombinant Type I Interferons

Jasper Fuk-Woo Chan a,b,c,d,*, 1,2, Anna Jinxia Zhang b,1, Chris Chung-Sing Chan b, Cyril Chik-Yan Yip b, Winger Wing-Nga Mak b, Houshun Zhu b, Vincent Kwok-Man Poon b, Kah-Meng Tee b, Zheng Zhu b, Jian-Piao Cai b, Jessica Oi-Ling Tsang b, Kenn Ka-Heng Chik b, Feifei Yin e, Kwok-Hung Chan b, Kin-Hang Kok b,c, Dong-Yan Jin f, Rex Kwok-Him Au-Yeung g, Kwok-Yung Yuen a,b,c,d,h,⁎

⁎ These authors contributed equally to the study as co-corresponding authors.
* These authors contributed equally to the study as co-first authors.
1 These authors contributed equally to the study as co-first authors.
2 These authors contributed equally to the study as co-corresponding authors.

Abstract

Background: Disseminated or fatal Zika virus (ZIKV) infections were reported in immunosuppressed patients. Existing interferon-signaling/receptor-deficient mouse models may not be suitable for evaluating treatment effects of recombinant interferons.

Methods: We developed a novel mouse model for ZIKV infection by immunosuppressing BALB/c mice with dexamethasone.

Results: Dexamethasone-immunosuppressed male mice (6–8 weeks) developed disseminated infection as evidenced by the detection of ZIKV-NS1 protein expression and high viral loads in multiple organs. They had ≥10% weight loss and high clinical scores soon after dexamethasone withdrawal (10 dpi), which warranted euthanasia at 12 dpi. Viral loads in blood and most tissues at 5 dpi were significantly higher than those at 12 dpi (P < 0.05). Histological examination revealed prominent inflammatory infiltrates in multiple organs, and CD45+ and CD8+ inflammatory cells were seen in the testis. These findings suggested that clinical deterioration occurred during viral clearance by host immune response. Type I interferon treatments improved clinical outcome of mice (100% vs 0% survival).

Conclusions: Besides virus dissemination, inflammation of various tissues, especially orchitis, may be potential complications of ZIKV infection with significant implications on disease transmission and male fertility. Interferon treatment should be considered in patients at high risks for ZIKV-associated complications when the potential benefits outweigh the side effects of treatment.

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1. Introduction

Zika virus (ZIKV) is an emerging flavivirus that has been largely neglected for >60 years after its discovery due to its restricted geographical distribution and its presumed low clinical significance (Chan et al., 2016a). Since 2007, large-scale outbreaks of ZIKV infection have occurred in the Pacific islands, Latin America, and most recently, USA and Southeast Asia (Duffy et al., 2009; Musso and Gubler, 2016; Zhu et al., 2016). As of 27 October 2016, >70 countries/territories have
reported continuing mosquito-borne transmission of ZIKV (World Health Organization. Zika situation report. October 27, 2016). In addition to mosquito-borne transmission, sexual and transplacental transmissions of ZIKV have also been reported (Chan et al., 2016a; Musso et al., 2015a; Foy et al., 2011; Calvet et al., 2016). These non-vector-borne transmission routes render the control of the continuing epidemic more complicated.

ZIKV was not considered as an important human pathogen in the past as most infected adult patients were asymptomatic or developed a self-limiting acute febrile illness which resolved within 1–2 weeks (Chan et al., 2016a; Duffy et al., 2009). However, it has been recently recognized that infected mothers may transmit the virus transplacentally, leading to congenital malformations, including microcephaly, cerebral malformations, ophthalmological and hearing defects, and arthrogryposis (Chan et al., 2016a; Mlakar et al., 2016; de Paula et al., 2016; Leal et al., 2016). Some infected adults may also develop severe neurological complications, such as Guillain–Barré syndrome, meningoencephalitis, and myelitis (Cao-Lormeau et al., 2016; Carteaux et al., 2016; Mecharles et al., 2016). Moreover, ZIKV-related fatalities have been increasingly recognized. Most of the patients with fatal infection had underlying medical conditions and some were markedly immunosuppressed, including a patient with systemic lupus erythematosus and rheumatoid arthritis who was on corticosteroid therapy and died of disseminated infection with detectable ZIKV RNA in blood, brain, spleen, liver, kidney, lung, and heart obtained at postmortem examination (Pan American Health Organization/World Health Organization (PAHO/WHO), 2015; Sarmiento-Ospina et al., 2016; Arzuza-Ortega et al., 2016).

A number of animal models have been developed for studying the pathogenesis and evaluating countermeasures for ZIKV infection. Rhesus macaques with subcutaneous ZIKV inoculation develop mild clinical signs that resemble the self-limiting illness in most infected immunocompetent adults (Dudley et al., 2016). This non-human primate model provides a robust platform for the evaluation of vaccines and countermeasures for ZIKV infection. Moreover, expertise and facilities for working with non-human primates are not available in most research laboratories. Wild-type adult BALB/c mice are not susceptible to intraperitoneal ZIKV inoculation (Lazear et al., 2016; Aziz et al., 2016; Arzuza-Ortega et al., 2016). Male and female BALB/c mice, 6–8 weeks old, were obtained from the Laboratory Animal Unit of The University of Hong Kong. The mice were kept in biosafety level-2 housing and given access to standard pellet food and water ad libitum. Virus inoculation experiments were performed in a biosafety level-2 animal facility according to the standard operating procedures approved by the Committee on the Use of Live Animals in Teaching and Research of The University of Hong Kong as we previously described (Zheng et al., 2008). Approval was obtained from the Committee on the Use of Live Animals in Teaching and Research of The University of Hong Kong. Male and female BALB/c mice, 6–8 weeks old, were obtained from the Laboratory Animal Unit of The University of Hong Kong. The mice were kept in biosafety level-2 housing and given access to standard pellet food and water ad libitum. Virus inoculation experiments were performed in a biosafety level-2 animal facility according to the standard operating procedures approved by the Committee on the Use of Live Animals in Teaching and Research of The University of Hong Kong as described previously (Zhang et al., 2014). The mice were randomly divided into 11 groups and given different regimens of virus inoculation, dexamethasone, and recombinant interferon treatment (Table 1). Phosphate-buffered saline (PBS) was used to dilute the virus stocks to the desired concentration, and inocula were back-titrated to verify the dose given. On the day of virus inoculation, a dose of the virus equivalent to 6 × 10⁶ TCID₅₀ (3.24 × 10⁶ plaque forming units) in 200 μl of PBS was inoculated via the intraperitoneal route into mice under ketamine (100 mg/kg) and xylazine (10 mg/kg) anesthesia. Mice in the negative-control groups (groups 5 to 8) were injected with the same volume of PBS. Mice were monitored three times each day for clinical signs of disease and a numerical score was assigned at each observation as previously described (Dowall et al., 2016; Graham et al., 2015). Their body weight and survival were monitored for 14 days post-inoculation (dpi) or until euthanasia. Three mice in each group (except groups 7 and 8 which included mock-infected control mice without dexamethasone immunosuppression and group 9 which included ZIKV-inoculated, dexamethasone-immunosuppressed mice without dexamethasone withdrawal) were sacrificed at 5 dpi for virological, histological, and immunohistochemistry analyses. The remaining mice were sacrificed at 14 dpi or euthanized when there was a 20% weight loss or 10% weight loss with ≥1 clinical sign (Dowall et al., 2016). Samples of brain, testis/epididymis (male), prostate (male), ovary/uterus (female), kidney, urinary bladder, spleen, liver, pancreas, intestine, heart, lung, and salivary gland were collected at necropsy. The specimens were separated into two parts, one immediately fixed in 10% PBS-buffered formalin, the other immediately frozen at −80 °C until further experiments. Blood samples were also collected for RNA extraction and real-time PCR analysis.

2.2. Animal Model and Viral Challenge

Might provide a Brandy Russell and Barbara Johnson, Centers for Disease Control and Prevention, USA. The virus was amplified by three additional passages in Vero cells (ATCC) in minimum essential medium (MEM) supplemented with 1% fetal calf serum and 100 units/ml penicillin plus 100 μg/ml streptomycin to make working stocks of the virus. For virus titration, aliquots of ZIKV were applied on confluent Vero cells in 96-well plates for 50% tissue culture infectious dose (TCID₅₀) assay as we previously described with slight modifications (Zhou et al., 2014). Briefly, serial 10-fold dilutions of ZIKV were inoculated in a Vero cell monolayer in quadruplicate and cultured in penicillin/streptomycin-supplemented MEM. The plates were observed for cytopathic effect for 5 days. Viral titer was calculated with the Reed and Munch endpoint method. One TCID₅₀ was interpreted as the amount of virus that causes cytopathic effect in 50% of inoculated wells.

2.3. Histopathology and Immunohistochemistry

Paraffin-embedded tissues were cut into 4–6 μm sections, mounted on slides, and stained with hematoxylin and eosin (H&E) for light microscopy examination as we previously described (Zheng et al., 2008).
For immunohistochemical staining of ZIKV-NS1 antigen, mouse antiserum against ZIKV-NS1 protein prepared as we previously described was used as primary antibody (Chan et al., 2016b). De-paraffinized and rehydrated tissue sections were treated with Antigen Unmasking Solution according to manufacturer’s instructions (Vector Laboratories Inc., Burlingame, CA, USA) and then stained with Mouse on Mouse Polymer IHC kit (Abcam, Cambridge, United Kingdom). The primary antibody mouse anti-ZIKV-NS1 antiserum (1:1000 dilution with 1% BSA/PBS) was incubated at 4 °C overnight. This was followed by Mouse on Mouse HRP polymer kit (Abcam) with horseradish peroxidase-conjugated secondary antibody for 15 min. Color development was performed using 3,3′-diaminobenzidine (DAB) (Vector Laboratories, Burlingame, CA, USA). For immunohistochemical staining of CD45 and CD8, the sections were incubated at 4 °C for overnight with primary antibody (rabbit anti-mouse CD45, or rat anti-mouse CD8α (Abcam) after antigen unmasking and blocking. This was then followed by incubation with biotin-conjugated goat anti-rabbit IgG or goat anti-rat IgG (Calbiochem, Darmstadt, Germany) for 30 min at room temperature. Streptavidin/peroxidase complex reagent (Vector Laboratories) was then added and incubated at room temperature for 30 min. Color development was done with DAB (Vector Laboratories). All tissue sections were examined microscopically by two pathologists in an operator-blinded manner. Images were captured with Nikon80i imaging system equipped with Spot-advance computer software.

2.4. Viral Load Studies

Total nucleic acid (TNA) was extracted from the blood and necropsied tissues using EZ1 Virus Mini Kit v2.0 and QIAasympnomy DSP Virus/Pathogen Mini Kit (QIAGEN, Hilden, Germany), respectively, as we previously described (Zheng et al., 2008; Chan et al., 2016b; Chan et al., 2015a). ZIKV envelope gene was measured by using Quantinova Probe RT-PCR Kit (QIAGEN) in LightCycler 96 Real-Time PCR System (Roche Diagnostics, Basel, Switzerland). 5 μl of purified TNA was amplified in a 20 μl-reaction containing 10 μl of 2× Quantinova SYBR Green RT-PCR Mix (QIAGEN) in LightCycler 96 Real-Time PCR System. 5 μl of purified TNA was amplified in a 20 μl-reaction containing 10 μl of 2× Quantinova SYBR Green RT-PCR Master Mix, 0.2 μl QN SYBR Green RT-mix, 0.5 μM forward primer (5′-ACGCGAGCTCATCCTATTTG-3′) and 0.5 μM reverse primer (5′-CAAGAGAAGGTGTTCGAAGAAGG-3′) for the β-actin gene. Reactions were incubated at 50 °C for 10 min, followed by 95 °C for 2 min, and then thermal cycled for 50 cycles (95 °C for 5 s, 55 °C for 30 s). Internal control (β-actin) gene was measured by using QuantiNova SYBR Green RT-PCR Kit (QIAGEN) in LightCycler 96 Real-Time PCR System. 5 μl of purified TNA was amplified in a 20 μl-reaction containing 10 μl of 2× Quantinova SYBR Green RT-PCR Master Mix, 0.2 μl QN SYBR Green RT-mix, 0.5 μM forward primer (5′-ACGCGAGCTCATCCTATTTG-3′) and 0.5 μM reverse primer (5′-CAAGAGAAGGTGTTCGAAGAAGG-3′) for the β-actin gene. Reactions were incubated at 50 °C for 10 min, followed by 95 °C for 2 min, and then thermal cycled for 50 cycles (95 °C for 5 s, 55 °C for 30 s). A series of 10-fold dilutions equivalent to 1 × 10⁰ to 1 × 10⁶ copies/reaction mixture were prepared to generate standard curves and run in parallel with the test samples.

2.5. Statistical Analysis

All data were analyzed with GraphPad Prism software (GraphPad Software, Inc). Kaplan-Meier survival curves were analyzed by the log rank test, and weight losses were compared using two-way ANOVA. Student’s t-test was used to determine significant differences in virus titers, and Tukey–Kramer post hoc tests were used to discern differences among individual treatment groups as previously reported (Aliota et al., 2016; Rossi et al., 2016). P-values < 0.05 were considered statistically significant.

3. Results

3.1. Dexamethasone-immunosuppressed Mice Developed Disseminated ZIKV Infection

To establish a novel mouse model for ZIKV infection, we compared the clinical, histological, and virological findings of male (group 1) and female (group 2) mice with dexamethasone immunosuppression and ZIKV inoculation with those of the appropriate controls (groups 3 to 8) (Table 1). In terms of the clinical parameters, the dexamethasone-immunosuppressed mice developed mild (~5%) weight loss (Fig. 1A) and no mortality at 5 dpi (Fig. 1B and C). The weight losses of the dexamethasone-immunosuppressed mice with ZIKV inoculation (groups 1 and 2)
were consistently more significant than those of their comparators, including the ZIKV-inoculated mice without dexamethasone immunosuppression (groups 3 and 4) and mock-infected mice without dexamethasone immunosuppression (groups 7 and 8) starting at 1 dpi (P < 0.05). Minimal histological changes and inflammatory infiltrates were seen in the tissues of the male and female mice with dexamethasone immunosuppression and ZIKV inoculation (groups 1 and 2). On the other hand, ZIKV-NS1 protein expression was detected by immunohistochemical staining in most tissues of these mice, but not in dexamethasone-immunosuppressed mice with mock infection (groups 5 and 6), suggesting that the viral protein expression was specific and not related to dexamethasone effects (Fig. 2). The dexamethasone-immunosuppressed mice with ZIKV inoculation (groups 1 and 2) also had high mean viral loads in blood and most tissues at 5 dpi, especially in the testis/epididymis, ovary/uterus, prostate, spleen, and pancreas (Figs. 3A and B and S1A and B). These findings at 5 dpi were suggestive of disseminated but non-lethal ZIKV infection involving different organs with minimal inflammatory response due to dexamethasone immunosuppression.

3.2. Clinical Deterioration with Multi-organ Inflammatory Cell Infiltrates Occurred in the Mice after Dexamethasone Withdrawal

To investigate the possible effects of immune reconstitution in the dexamethasone-immunosuppressed mice, dexamethasone was stopped after 9 dpi. This led to prominent weight loss and increased symptoms in the dexamethasone-immunosuppressed mice (groups 1 and 2). The most prominent body weight loss was observed in the male mice with dexamethasone immunosuppression and ZIKV inoculation (group 1), with all 6 mice having weight loss of ≥10% at 12 dpi (Fig. 1A). All of the female mice with dexamethasone immunosuppression and ZIKV inoculation (group 2) also had progressive weight loss and 4/6 (66.7%) of them had ≥10% weight loss at 14 dpi. In contrast, none of the mice with dexamethasone immunosuppression from 3 days before to 13 days post-infection (group 9) developed abrupt weight loss between 10 dpi and 14 dpi (Fig. 1A). The weight loss of mice in groups 1 and 2 became consistently more than those of their comparators in the other control groups (groups 3 to 8), including those in the dexamethasone-immunosuppressed mice with mock infection (groups 5 and 6) since 10 dpi (P < 0.05). Together, these findings suggested that the combination of immune reconstitution after dexamethasone withdrawal and disseminated virus infection were responsible for the abrupt clinical deterioration. All of the mice in groups 1 and 2 developed rapid breathing, lethargy, and/or ruffled fur since 11 dpi (group 1) or 12 dpi (group 2), shortly after dexamethasone was stopped (Fig. 1B). The reasons for the earlier onset of weight loss and symptoms in the male mice were not fully understood, but might be related to the higher cumulative dose of dexamethasone because of their baseline body weights and/or possible effects of androgen on virus replication (Tian et al., 2012). Based on the predefined criteria, all 6 (100%) male and 4/6 (66.7%) female mice were euthanized at 12 dpi and 14 dpi, respectively (Fig. 1C). Comparatively, all the mice in the other control groups (groups 3 to 9) had either gained weight or had <5% weight loss with spontaneous recovery at 14 dpi (Fig. 1A), remained asymptomatic (Fig. 1B), and survived through the study period (Fig. 1C).

At euthanasia (12–14 dpi), H&E staining of the necropsied tissues of these mice showed prominent acute inflammatory reactions with predominantly lymphocytic infiltrates. The most prominent inflammatory changes were seen in the brain (cortical parenchymal and perivascular lymphocytic infiltrates) (Fig. 4A to C), kidney (acute tubulitis and interstitial inflammation) (Fig. 4D to F), and testis (necrotic and hemorrhagic seminiferous tubules with marked lymphocytic infiltration in the perimeter of the tubules and the interstitium) (Fig. 5A to D). ZIKV-NS1
protein expression was still visible, but to a lesser degree, in the immunohistochemical staining of the testis/epididymis, ovary/uterus, kidney, spleen, small intestine, pancreas, and salivary gland of the dexamethasone-immunosuppressed mice with ZIKV inoculation at 12–14 dpi compared with 5 dpi. In contrast, no inflammatory reaction and viral protein expression were seen in any organ of the control mice with ZIKV inoculation alone (groups 3 and 4) or dexamethasone immunosuppression alone (groups 5 and 6) (Fig. 5C and D). These findings confirmed that mice with ZIKV inoculation but no dexamethasone immunosuppression were not susceptible to infection as previously reported, and that the histological changes in the model mice (groups 1 and 2) were unrelated to dexamethasone-induced effects such as drug-induced testicular toxicity (Lazear et al., 2016; Dowall et al., 2016; Aliota et al., 2016; Rossi et al., 2016). Overall, these findings were suggestive of multi-organ inflammation upon immune reconstitution with partial viral clearance in the mice after withdrawal of dexamethasone immunosuppression.

3.3. Treatment with Recombinant Type I Interferons was Associated with Better Clinical Outcome in the Mice

We next evaluated the effects of recombinant type I interferon treatment in our mouse model. We used male mice as they had earlier onset of weight loss and clinical symptoms requiring necropsy at 12 dpi. The mice were treated with pegylated interferon-α2b (PegIntron®, Merck & Co., Inc., Whitehouse Station, NJ, USA) 1920 IU/dose every 96 h subcutaneously at 1 dpi, 5 dpi, and 9 dpi (group 10) or interferon-β1b (Betaferon®, Bayer Schering Pharma AG, Berlin, Germany) 160,000 IU/dose every 48 h intraperitoneally at 1 dpi, 3 dpi, 5 dpi, 7 dpi, and 9 dpi (group 11). As shown in Fig. 6A, the mice treated with pegylated interferon-α2b (group 10) or interferon-β1b (group 11) had ~10% weight loss with spontaneous recovery at 14 dpi. The weight loss of the untreated group became significantly more than those of the mice treated with either interferon-α2 or interferon-β1b starting at 10 dpi (P < 0.05). All...
of these mice remained asymptomatic and survived through the study period (Fig. 6B and C). None of their tissues showed prominent inflammatory reactions in H&E staining at 5 dpi or 14 dpi. ZIKV-NS1 protein expression was only rarely seen in the immunohistochemical staining of the testis/epididymis, kidney, spleen, small intestine, lung, and pancreas collected at 5 dpi, and testis, epididymis, kidney, and spleen at 14 dpi. They had reduced mean viral loads in blood and all the tissues (\(12-4\log_{10}\text{copies}/10^6\beta\)-actin) as compared with those of the untreated mice at 5 dpi and 14 dpi (Fig. 7A and B). The reductions were most significant in the tissues with high viral loads, such as the spleen, testis, pancreas, and prostate (\(P < 0.05\)). Overall, these findings suggested that early use of systemic recombinant type I interferons improved the clinical, histological, and virological parameters of mice with disseminated ZIKV infection.

4. Discussion

The full spectrum of clinical manifestations and complications of ZIKV infection remains incompletely understood as of today. The previous assumption that ZIKV infection is an entirely self-limiting disease without severe or long-lasting sequelae has been overturned by the increasing recognition of congenital malformations, neurological complications, immune-mediated thrombocytopenia, and even fatality in some immunosuppressed patients (Pan American Health Organization/
World Health Organization (PAHO/WHO), 2015; Sarmiento-Ospina et al., 2016; Arzuza-Ortega et al., 2016; Duijster et al., 2016). Notably, patients with severe non-pregnancy-related complications of ZIKV often deteriorated suddenly after an initially mild disease phase as the viral load began to decrease (Cao-Lormeau et al., 2016; Mecharles et al., 2016; Sarmiento-Ospina et al., 2016). This has led us to hypothesize that, like many other flavivirus infections, including yellow fever, dengue, and West Nile virus infection, the host immune response may also play a role in these ZIKV-associated complications, especially during the viral clearance phase by the host immune system (Quaresma et al., 2013; Screaton et al., 2015; Wang et al., 2003). In this study, we characterized a novel and readily available mouse model for severe ZIKV infection which attempts to provide an alternative venue for studying the host immune response of and evaluating countermeasures for ZIKV infection. The findings in our study have important implications on the pathogenesis, potential complications, and treatment of ZIKV infection.

The dexamethasone-immunosuppressed mice with ZIKV inoculation in our study developed disseminated infection with viremia and multi-organ involvement, including the brain, urogenital tract, intestine, liver, spleen, pancreas, heart, lung, and salivary gland as evident by ZIKV-NS1 protein expression on immunohistochemical staining and/or detectable viral load in these tissues. Immunohistochemistry staining of the testis confirmed the presence of inflammatory cell infiltration (pan-leukocyte marker CD45 +) with predominantly CD8+ T lymphocytes. Clinically, the male mice developed earlier onset of disease than the female mice, with ≥10% weight loss and ≥1 clinical sign, which warranted euthanasia at 12 dpi. Their weight loss, clinical scores, and histological evidence of inflammatory reactions were most severe soon after dexamethasone withdrawal, when viral loads had already decreased by about 2–5 log_{10} copies/10^6 /β-actin. Overall, these findings suggested that, like the other related flaviviruses, the host immune response might have led to marked clinical deterioration in the face of disseminated ZIKV infection at the time when immune-mediated clearance of the virus began. Our findings provided an additional explanation for the pathogenesis of fatal ZIKV infection, which has been proposed to be related to uncontrolled virus dissemination in previously described mouse models utilizing types I/II interferon-signaling/-receptor-deficient mice that were unable to mount a robust host innate immune response.

Our mouse model is also useful for studying ZIKV’s tissue tropism and potential complications of severe ZIKV infection. In addition to the reported findings of detectable virus particles and/or RNA in the brain, spinal cord, kidney, spleen, liver, testis, ovary, heart, lung, muscle, and blood of types I/II interferon-signaling-/receptor-deficient mice with ZIKV infection, our study identified intestine, pancreas, and salivary gland as other possible tissues and anatomical sites for virus infection (Dick, 1952; Lazear et al., 2016; Dowall et al., 2016; Allota et al., 2016; Rossi et al., 2016). This tissue tropism of ZIKV in our mouse model concurs with the in-vitro observation that ZIKV efficiently replicates in diverse cell types of neuronal, testicular, prostatic, renal, intestinal, hepatic, and placental origin (Chan et al., 2016b; Brault et al., 2016; Hughes et al., 2016). Such degree of virus dissemination and multi-organ involvement is also compatible with the clinical findings in patients with severe and/or fatal ZIKV infection, in whom viral particles and/or RNA were detected in multiple organs at post-mortem examination (Pan American Health Organization/World Health Organization (PAHO/WHO), 2015; Sarmiento-Ospina et al., 2016; Arzuza-Ortega et al., 2016). While inflammatory neurological complications, such as Guillain-Barré syndrome, meningoencephalitis, and myelitis, have been recently reported in patients with ZIKV infection, inflammatory disorders of the other non-neuronal tissues were not well recognized (Cao-Lormeau et al., 2016; Carteaux et al., 2016; Mecharles et al., 2016). Our findings showed that inflammation could be observed in multiple organs including the testis, kidney, spleen, liver, intestine, pancreas, lung, and salivary gland outside the nervous system.

Among these non-neuronal tissues, the inflammatory reactions were most prominent in the testis of our model mice. Some patients with ZIKV infection reported hematospermia, pelvic pain, and dysuria with detectable viral particles and/or RNA in their semen (Chan et al., 2016a; Musso et al., 2015a; Foy et al., 2011). Histological evidence of orchitis has not been reported due to the difficulty in obtaining the patients’ testicular tissues for histological examination. Previous mouse
models for ZIKV infection utilizing types I/II interferon-signaling-/receptor-deficient mice have also showed that viral particles and RNA could be detected in the mice’s testes, but histological analysis were not reported (Lazear et al., 2016). The markedly necrotic and hemorrhagic seminiferous tubules observed in our mice are highly alarming as orchitis may have long-term effects on fertility. These changes were not accountable by dexamethasone-induced testicular toxicity, as they were morphologically different from the latter, and were not present in any of the testes of the control mice with dexamethasone treatment and mock infection (Khorsandi et al., 2013). During revision of this

Fig. 5. Representative histological findings and immunohistochemistry staining of the testis of dexamethasone-immunosuppressed BALB/c mice with ZIKV inoculation and dexamethasone-immunosuppressed mock-infected control mice. Sections (H&E) of the testis of a dexamethasone-immunosuppressed mouse with ZIKV inoculation (A and C) and mock infection (B and D). (A, original magnification ×100) and (C, original magnification ×400): necrotic and hemorrhagic seminiferous tubules associated with marked lymphocytic infiltration (arrows) in the perimeter of the tubules and the interstitium. (B, original magnification ×100) and (D, original magnification ×400): normal architecture consisting of seminiferous tubules with layers of cells in normal maturation stages. Immunohistochemistry staining of CD45 (E and F) and CD8 (G and H) antigens showing CD45+ and CD8+ cells (arrows) in the testis of dexamethasone-immunosuppressed mice with ZIKV inoculation (E and G) but not the dexamethasone-immunosuppressed mice with mock infection (F and H) (E to H, original magnification ×400).
work, similar findings were reported in the testes of C57BL/6 mice treated with anti-Ifna1 blocking monoclonal antibody and inoculated with ZIKV (Govero et al., 2016). Clinical studies to confirm the presence of orchitis and to assess the fertility of convalescent male patients should be conducted to ascertain the long-term consequences of ZIKV infection regarding reproductive and hormonal derangements. Inflammation of other organs, such as acute tubulitis, interstitial nephritis, sialadenitis, hepatitis, enteritis, and acute pancreatitis have been reported in patients with ZIKV or other flavivirus infections (Chan et al., 2016a; Sarmiento-Ospina et al., 2016; Arzuza-Ortega et al., 2016; Dujister et al., 2016; Bonaldo et al., 2016; Gourinat et al., 2015; Musso et al., 2015b; Mercado et al., 2016; Bhagat et al., 2012; Torres et al., 2000; Macnamara, 1954; Chatterjee et al., 2014). These potential complications may be increasingly recognized as the ZIKV epidemic continues to expand into developed countries with a large ageing and immunosuppressed population.

Finally, our mouse model also provided a novel avenue for the evaluation of anti-ZIKV treatment. Type I interferons have broad-spectrum antiviral activities including those against ZIKV, but type I interferon-signaling-/receptor-deficient mice were not suitable for evaluation of the effects of recombinant type I interferons. We therefore evaluated the antiviral effects of two commercially available preparations of recombinant type I interferons in this new mouse model (Hamel et al., 2015; Zumla et al., 2016; Chan et al., 2013; Chan et al., 2015b). We showed that the early use of either drug was associated with improved clinical outcome with no fatality (100% fatality in untreated mice), markedly decreased inflammatory response after dexamethasone withdrawal, and reduced viral loads in various tissues of the mice as compared to those of the untreated mice. The viral load reductions were especially significant in the early phase of the disease (5 dpi), when the mice were on dexamethasone. These findings suggested that the early use of recombinant interferons might help to control viral replication during the initial phase of infection, and prevent the subsequent development of severe complications related to an exaggerated immune response in the presence of high viral loads as seen in the untreated mice. It is important to further confirm these results in our mouse model using different ZIKV strains and in clinical trials because ZIKV antagonizes mouse STAT2 less efficiently than human STAT2, and thus may be more susceptible to type I interferons in mice (Grant et al., 2016). While most patients with mild ZIKV infection may not require systemic interferon treatment, clinical trials should be considered to evaluate the benefits of the early use of interferon treatment in patients at risk of developing severe ZIKV-associated complications, such as those with underlying comorbidities (Sarmiento-Ospina et al., 2016). The increased risk of fetal loss and low birth weight associated with interferon therapy in the first trimester of pregnancy may be outweighed by the risk of congenital malformations due to ZIKV infection. The optimal timing of treatment commencement should be further investigated as late commencement of interferon treatment may be useless or deleterious and should be avoided (Solomon et al., 2003).

In summary, this novel mouse model is useful for investigating host immune response-associated damage of and evaluating countermeasures for ZIKV infection. Inflammation of different visceral organs may be important complications of ZIKV that should be further studied in infected humans. Long-term monitoring of the testicular function of ZIKV-infected male patients should be considered. Clinical trials should be
considered for evaluating the effects of recombinant interferon treatments in patients at high risk for ZIKV-associated complications when the potential benefits may outweigh the side effects of treatment.

Supplementary data to this article can be found online at http://dx.doi.org/10.1016/j.ebiom.2016.11.017.

Author Contribution

JFWC, AJZ, CCSC, and KYY designed the study. AJZ, CCSC, HZ, and VKMP performed infections. CCYY, JOLT, KKHC, and KHC prepared virus stocks and viral titrations. AJZ, WWNM, VKMP, and RKHAY prepared histology and immunohistochemistry slides. CCYY, KMT, ZZ, and JPC performed viral load studies. JFWC, AJZ, RKHAY, VKMP, and KYY acquired images at the microscope, analyzed, and quantified the data. FY, KHK, DYJ provided technical assistance and edited the manuscript. JFW, AJZ, and KYY wrote the manuscript.

Conflict of Interest Statement

Jasper F.W. Chan has received travel grants from Pfizer Corporation Hong Kong and Astellas Pharma Hong Kong Corporation Limited, and was an invited speaker for Gilead Sciences Hong Kong Limited. The other authors declared no conflict of interests. The funding sources had no role in study design, data collection, analysis or interpretation or writing of the report. The corresponding authors had full access to all the data in the study and had final responsibility for the decision to submit for publication.

Acknowledgements & Funding

We are grateful to Can Li, Andrew Chak-Yiu Lee, and Shuofeng Yuan for their facilitation of the study. This work was partly supported by the donations of Larry Chi-Kin Yung, and Hui Hoy and Chow Sin Lan Charity Fund Limited; and funding from the Consultancy Service for Enhancing
Laboratory Surveillance of Emerging Infectious Diseases of the Department of Health, Hong Kong Special Administrative Region; and the Committee for Research and Conference Grant, The University of Hong Kong; and the Collaborative Innovation Center for Diagnosis and Treatment of Infectious Diseases, the Ministry of Education of China.

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