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<tr>
<td>Citation</td>
<td>Chemical Science, 2016, v. 7 n. 3, p. 2094-2099</td>
</tr>
<tr>
<td>Issued Date</td>
<td>2016</td>
</tr>
<tr>
<td>URL</td>
<td><a href="http://hdl.handle.net/10722/231635">http://hdl.handle.net/10722/231635</a></td>
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HKOCI-3: a fluorescent hypochlorous acid probe for live-cell and in vivo imaging and quantitative application in flow cytometry and a 96-well microplate assay†

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Ultra-selective and ultra-sensitive probes for hypochlorous acid (HOCl), one of the most poorly understood reactive oxygen species (ROS), are urgently needed to unravel the HOCl functions in important biological processes such as development and innate immunity. Based on a selective oxidative O-dearylation reaction of 2,6-dichlorophenol toward HOCl over other reactive oxygen species, we have developed a novel fluorescent probe HKOCI-3 for HOCl detection with ultra-selectivity, ultra-sensitivity and a rapid turn-on response. The functional robustness of HKOCI-3 for endogenous HOCl detection and imaging has been thoroughly scrutinized in multiple types of phagocytes and in vivo imaging of live intact zebrafish embryos. Furthermore, HKOCI-3 has been successfully applied to the detection of endogenous HOCl by a 96-well microplate assay and flow cytometry. Therefore, HKOCI-3 holds great promise as a versatile molecular tool that enables innovative investigation of HOCl biology and ROS-related diseases in multiple detection modalities.

Results and discussion

Design and synthesis of the fluorescent probe HKOCI-3

A widely used fluorescent probe HPF, reported by Nagano and coworkers,12 shows a strong response toward ’OH and ONOO−,
but not HOCl. As part of our continued efforts in developing fluorescent probes for the detection of specific reactive oxygen species, we noticed that, by incorporating ortho halogen substituents, the selectivity of the resulting phenols toward HOCl over other reactive oxygen/nitrogen species can be dramatically improved. Therefore, we designed a new fluorescent probe HKOCl-3 (Scheme 1) by incorporating two ortho chlorine substituents into HPF. Since the introduction of two chlorine atoms (2,6-dichlorophenol moiety) lowers the pKa of the phenol by more than 3 orders of magnitude (from 10.0 to 6.79), the election-rich phenoxide form of the phenol by more than 3 orders of magnitude (from 10.0 to 6.79), the election-rich phenoxide form of HKOCl-3 will be dominant under physiological conditions (pH = 7.4), resulting in a great enhancement of its nucleophilicity and reactivity toward HOCl. In addition, the phenoxide form of HKOCl-3 can quench the fluorescence by a photoinduced electron transfer (PeT) process more efficiently, affording lower basal fluorescence. Herein, we report that HKOCl-3 is indeed an excellent fluorescent probe for HOCl detection with ultra-selectivity, ultra-sensitivity and a rapid turn-on response in live cells and in vivo.

As shown in Scheme 2, the probe HKOCl-3 was readily synthesized from diarylether 4 (obtained by Ullmann coupling of 3-methoxyphenol and aryl iodide 2, followed by demethylation) and benzoic acid 5 (prepared from fluorescein) in a good yield.

**Reactivity and selectivity of HKOCl-3 for HOCl**

For chemical characterization, we first investigated the spectroscopic properties of the fluorescent probe HKOCl-3 (10 μM) in potassium phosphate buffer (0.1 M, pH 7.4, 0.1% DMF) at 25 °C. HKOCl-3 showed an obvious absorption peak at 455 nm (ε = 2.3 × 10^4 M⁻¹ cm⁻¹; Fig. S1†) and as expected, its fluorescence is completely quenched (Φ = 0.001). Upon exposure to 1 equiv. HOCl, a >358-fold enhancement in the fluorescence intensity was observed, demonstrating the ultra-sensitivity of this probe in aqueous solution (Fig. 1a). A linear relationship of the fluorescence intensity of HKOCl-3 at 527 nm with the concentrations of HOCl (0–10 μM) was observed (Fig. 1b). Moreover, the detection limit of HKOCl-3 was calculated to be as low as 0.33 nM (3σ/k). This again confirms the ultrasensitivity of HKOCl-3, compared with the previously reported HOCl probes. In particular, the reaction of HKOCl-3 (10 μM) with HOCl caused a dramatic time-dependent fluorescence increase, which was complete within 1 min (Fig. 1c), suggesting a remarkably fast reaction between the probe and HOCl.

Interestingly, upon treatment with HOCl, HKOCl-3 underwent a bathochromic shift in the absorption peak (from 455 nm to 499 nm, Fig. S1, ESI†) and fluorescence emission peak (Fig. 1a and S2, ESI†). This indicates that the oxidation of HKOCl-3 produced fluorescent products, such as fluorescein and its mono- or di-chlorinated derivatives (Scheme 1), which have been confirmed by ESI-MS analysis (ESI†). Moreover, the fluorescence signal remained unchanged for 30 min (Fig. S3, ESI†), indicating the striking chemostability of the fluorescent products toward the highly reactive HOCl in the reaction mixture.

Next, the selectivity of HKOCl-3 was examined by measuring its fluorescence response upon treatment with various analytes.
in potassium phosphate buffer (0.1 M, pH 7.4). As shown in Fig. 1d and Table S1 (ESI†), 1 equiv. of HOCl gave a >358-fold enhancement in fluorescence intensity, while 10 equiv. (100 μM) of other biologically relevant reactive oxygen/nitrogen species (H2O2, O2·-, ROO·, TBHP, 'NO and O2−) only triggered a negligible fluorescence increase. Specifically, the probe exhibited a >83-fold higher increase in fluorescence intensity toward HOCl over other potentially interfering highly reactive oxygen species (‘OH and ONOO−). Collectively, these results demonstrate the ultra-selectivity of HKOCl-3 for HOCl.

The biocompatibility of HKOCl-3 was then examined. HKOCl-3 exhibits an excellent stability toward pH changes (3.0–10.8, Fig. S4, ESI†) and its fluorescence response toward HOCl was significant across a wide range of pH values (4–10) relevant to cellular processes (Fig. S5, ESI†). The interference from common coexisting biological substances (Na+, Ca2+, Mg2+, Zn2+, glutathione, etc.) was found to be minimal (Fig. S6, ESI†). Collectively, these results suggest that the probe HKOCl-3 could be used to detect HOCl reliably in complex cellular milieus.

### Evaluation of HKOCl-3 for endogenous HOCl detection in live cells and in vivo

As a first step to establish the biological applications of HKOCl-3, the cytotoxicity of HKOCl-3 was assessed in RAW264.7 mouse macrophages. HKOCl-3 was found to be virtually nontoxic when used up to 20 μM after 24 h incubation (Fig. S7, ESI†). Then we examined the performance of this probe in confocal imaging of endogenous HOCl in different types of activated phagocytes: RAW264.7 mouse macrophages, BV-2 mouse microglia, THP-1 human monocyctic macrophages and primary human polymorphonuclear neutrophils (PMNs). To induce endogenous HOCl, different types of phagocytes were co-incubated with HKOCl-3 (1 μM) and the PKC activator PMA (phorbol myristate acetate: 500 ng mL−1) for 30 min, followed by confocal imaging (Fig. 2, S8 and S9, ESI†). As expected, unstimulated phagocytes loaded with HKOCl-3 showed barely detectable background fluorescence signals (Fig. 2). In the presence of PMA, the fluorescence signal was significantly enhanced in all four types of cells. This result suggests that endogenous HOCl production can be robustly visualized with HKOCl-3 in activated phagocytes.

The 96-well microplate fluorescence measurement is an indispensable platform for the development of low-cost, high-throughput screening assays, and is preferred over flow cytometry for its ability to preserve the physical integrity of live adherent cells during tests. However, the high background noise of 96-well microplate assays prohibits the successful application of numerous fluorescent probes, despite their excellent performance in chemical systems. To test the applicability of HKOCl-3 in this platform, RAW264.7 mouse macrophages were co-incubated with HKOCl-3 (2 μM) and various concentrations of PMA (0–1000 ng mL−1) for 30 min before the fluorescence measurement. We found that HKOCl-3 gave a dose-dependent response toward the PMA stimulation in RAW264.7 cells in a sensitive manner (Fig. 3a), which is in agreement with our confocal imaging results (Fig. S10, ESI†).

Next, the selectivity of HKOCl-3 toward HOCl was thoroughly scrutinized by examining the effects of a series of enzyme inhibitors. RAW264.7 cells were co-incubated with HKOCl-3 (2 μM) and PMA (500 ng mL−1) in the presence or absence of one of the following inhibitors: the PKC inhibitors G6983, G6976 and Ro32-0432, the NOX (NADPH oxidase) inhibitor DPI (diphenyleneiodonium chloride), and the MPO (myeloperoxidase) inhibitor ABAH (aminobenzoic acid hydrazide). The observation that the fluorescence signal was potently and dose-dependently reduced by these inhibitors confirms the selectivity of HKOCl-3 toward HOCl detection, as PKC and NOX are known to be upstream of MPO in the pathway for endogenous HOCl production in activated RAW264.7 cells (Fig. 3b, ESI†).

The 96-well microplate platform has also been applied to evaluate the HOCl scavenging efficiency of five organosulfur compounds, namely: α-lipoic acid (which participates in aerobic metabolism), NAC ([N-acetyl cysteine], γ-methionine, γ-cysteine, and γ-cystine (Fig. 4 and S11, ESI†)). Except for γ-cystine, which is a dimeric form of oxidized γ-cysteine with a very poor aqueous solubility (200 μM in HBSS), all scavengers in the mM range were able to remove the HOCl generated in cells, with the order of scavenging efficiency as follows: γ-cysteine < γ-methionine < α-lipoic acid < NAC. Therefore, for the first time, we quantitatively and directly evaluated the HOCl removal effects of these scavengers, which support the findings generated in cell-free systems or by indirect measurement. To the best of our knowledge, HKOCl-3 is the first small-molecule probe that can be successfully applied to a 96-well microplate assay and can screen for molecules that scavenge HOCl or modulate its production.

Flow cytometry has become a definitive quantitative cellular analysis technique that rapidly integrates information about
the multifaceted characteristics of cell populations (e.g. 10^3 cells per second). Based on the superb probe performance seen in confocal imaging and the 96-well microplate assay, we decided to explore the application of HKOCl-3 in flow cytometry. RAW264.7 cells were co-incubated with HKOCl-3 (2 μM) and PMA (500 ng mL^-1) in the presence or absence of the NOX inhibitor DPI (100 nM) for 30 min, followed by flow cytometry analysis (Fig. 5). In the presence of PMA, the geometric mean of the fluorescence intensity was significantly elevated. The appearance of two populations of cells upon the PMA challenge is an interesting and highly reproducible phenomenon in this assay, which could be explained by the distinct excitability or responsiveness of cell subpopulations (possibly under different cellular redox status) toward PMA treatment. In addition, DPI blunted this fluorescence increase to a great extent. This result demonstrates that endogenous HOCl production in RAW264.7 cells can be readily detected in flow cytometry by using HKOCl-3 as a fluorescent probe.

For decades, MPO-related oxidative damage in diverse inflammatory, vascular and neurodegenerative diseases has been reported in numerous studies, which vividly illustrates the functional relevance of HOCl in cardiovascular and nervous systems and the dire consequences of its dysregulated production. Despite the well-documented pathological roles of HOCl in tissue damage, virtually nothing is known of HOCl function in normal tissues, such as those in developing fetuses. Strikingly, in zebrafish, an excellent in vivo model for studying the
established by visualizing endogenous HOCl in multiple living phagocytes in an oxidative burst model and in the in vivo imaging of live zebrafish embryos at different developmental stages. In addition, HKOCI-3 has shown superior sensitivity in the quantitative detection of endogenous HOCl in flow cytometry and a 96-well microplate assay. Collectively, our probe is expected to offer exciting glimpses into the unknown roles of HOCl in cellular processes and to help accelerate discoveries of drugs for the treatment of human diseases.

Acknowledgements

We thank Prof. Anskar Yu-Hung Leung, Dr Jing Guo, Dr Alvin Chun-Hang Ma and Yuhan Guo for technical assistance, and the HKU Li Ka Shing Faculty of Medicine Faculty Core Facility and Zebrafish Core Facility, and the HKU School of Biological Sciences Central Facilities Laboratory for support in confocal microscopy. This work was supported by The University of Hong Kong, the University Development Fund, and Hong Kong Research Grants Council under General Research Fund Scheme (HKU 706410P and HKU 17305714).

Notes and references

A comparison of the performance of recently published fluorescent probes for HOCl imaging in terms of selectivity and sensitivity can be found in Table S2 of ESI.