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The K526R substitution in viral protein PB2 enhances the effects of E627K on influenza virus replication

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Host-adaptive strategies, such as the E627K substitution in the PB2 protein, are critical for replication of avian influenza A viruses in mammalian hosts. Here we show that mutation PB2-K526R is present in some human H7N9 influenza isolates, in nearly 80% of H5N1 human isolates from Indonesia and, in conjunction with E627K, in almost all seasonal H3N2 viruses since 1970. Polymerase complexes containing PB2-526R derived from H7N9, H5N1 or H3N2 viruses exhibit increased polymerase activity. PB2-526R also enhances viral transcription and replication in cells. In comparison with viruses carrying 627K, H7N9 viruses carrying both 526R and 627K replicate more efficiently in mammalian (but not avian) cells and in mouse lung tissues, and cause greater body weight loss and mortality in infected mice. PB2-K526R interacts with nuclear export protein and our results suggest that it contributes to enhance replication for certain influenza virus subtypes, particularly in combination with 627K.
Influenza viruses utilize the viral polymerase complex, which is composed of PB1, PB2 and PA subunits, to replicate and transcribe the viral genome in the cell nucleus. Adaptation of viral polymerase is critical for efficient virus replication in a new host following cross-species transmission. Several adaptation markers in the polymerase have been identified among seasonal influenza viruses that are circulating in humans and avian influenza viruses, which cause sporadic human infections. The most well-characterized adaptation marker is PB2 E627K, which was found in a significant proportion of H5N1 human infections and also in one case of H7N7 infection. However, E627K is not exclusive to all influenza viruses that can replicate in humans.

Another adaption marker, PB2 590S/591R, was reported to enable replication fitness of swine origin 2009 H1N1 virus in humans. PB2 D701N and Q591K substitutions were also found to support H5N1 virus replication in mammalian hosts. The E627K substitution enhances H5N1 virus replication in the upper respiratory tract, where the temperature, at 33°C, is slightly lower than the core body temperature. PB2 627K and 627E/701N have been associated with enhanced transmission of influenza virus in mammalian hosts. A T271A substitution in PB2 was also described to enhance replication of 2009 H1N1 and H5N1 viruses in mammalian cells in vitro. PB2 has been reported to interact with host τ-importins; it is thought that differential binding of PB2 with importin-τ1 and -τ7 may regulate influenza virus polymerase activity. Details of how these genetic substitutions translate into adaptive mechanisms remain largely unknown.

While about 50% of H5N1 human isolates contain known PB2 adaptation markers, mainly E627K with some D701N and a few instances of 591K, as described above, the other half of human infections are not associated with known adaptation markers. Indonesia has experienced the highest number of human H5N1 infections, with 195 cases, 163 of which were fatal (http://www.who.int/influenza/human_animal_interface/EN_GIP_2014_0124CumulativeNumberH5N1cases.pdf). However, the majority (80%) of the H5N1 isolates from human cases in Indonesia do not harbour the PB2 627K or 701N markers. Apart from the markers described above, influenza viruses probably utilize uncharacterized adaptive substitutions in the PB2 polymerase subunit to support virus replication in human cells. In the spring of 2013, a novel avian H7N9 virus emerged that crossed the species barrier to infect humans in eastern China. More than 300 laboratory-confirmed cases of human infection, about 30% of which were fatal, have been identified since March 2013. While sporadic human infections caused by avian H5N1, H9N2 and H7N7 viruses have been recorded since 1997 (refs 21–23), no avian influenza virus has caused an outbreak of this magnitude since the 1918 pandemic. It is important to understand how this H7N9 virus gained the ability to infect humans so easily. Genetic characterization revealed that the H7N9 virus is a reassortant virus, possibly generated through multiple reassortment events to incorporate the haemagglutinin (HA) gene from an H7N3 virus, the neuraminidase gene originating from an H7N9 virus circulating in wild birds and other internal genes from H9N2 virus present in poultry. More than 300 laboratory-confirmed cases of human infection, about 30% of which were fatal, have been identified since March 2013. While sporadic human infections caused by avian H5N1, H9N2 and H7N7 viruses have been recorded since 1997 (refs 21–23), no avian influenza virus has caused an outbreak of this magnitude since the 1918 pandemic. It is important to understand how this H7N9 virus gained the ability to infect humans so easily. Genetic characterization revealed that the H7N9 virus is a reassortant virus, possibly generated through multiple reassortment events to incorporate the haemagglutinin (HA) gene from an H7N3 virus, the neuraminidase gene originating from an H7N9 virus circulating in wild birds and other internal genes from H9N2 virus present in poultry. More than 300 laboratory-confirmed cases of human infection, about 30% of which were fatal, have been identified since March 2013. While sporadic human infections caused by avian H5N1, H9N2 and H7N7 viruses have been recorded since 1997 (refs 21–23), no avian influenza virus has caused an outbreak of this magnitude since the 1918 pandemic. It is important to understand how this H7N9 virus gained the ability to infect humans so easily. Genetic characterization revealed that the H7N9 virus is a reassortant virus, possibly generated through multiple reassortment events to incorporate the haemagglutinin (HA) gene from an H7N3 virus, the neuraminidase gene originating from an H7N9 virus circulating in wild birds and other internal genes from H9N2 virus present in poultry. More than 300 laboratory-confirmed cases of human infection, about 30% of which were fatal, have been identified since March 2013. While sporadic human infections caused by avian H5N1, H9N2 and H7N7 viruses have been recorded since 1997 (refs 21–23), no avian influenza virus has caused an outbreak of this magnitude since the 1918 pandemic. It is important to understand how this H7N9 virus gained the ability to infect humans so easily. Genetic characterization revealed that the H7N9 virus is a reassortant virus, possibly generated through multiple reassortment events to incorporate the haemagglutinin (HA) gene from an H7N3 virus, the neuraminidase gene originating from an H7N9 virus circulating in wild birds and other internal genes from H9N2 virus present in poultry.

Results

PB2-K526R is present in three influenza virus subtypes. Examination of the sequences of H7N9 human isolates has revealed a variety of different mammalian adaptation markers present in PB2 genes. Besides the previously described 627K, 701N and 591K loci, we found a K526R genotype associated with either 627E/627K or 701D/701N (Supplementary Fig. 1). Subsequent analysis found three distinct major PB2 genotypes, 526R/627K, 526R/701N and 526R alone, to be present within some isolates obtained from our own study and in some reported H7N9 human cases identified since March 2013 in China (Table 1). The 526R substitution has not previously been described to be a mammalian adaptation marker for avian influenza viruses. However, screening of sequences in the GenBank influenza database revealed that K526R is present in some H5N1 human isolates, particularly those from human cases in Indonesia (Table 1). This may explain why the majority of human isolates from Indonesia do not carry the PB2-627K marker and the PB2-526R genotype immediately became the dominant circulating H3N2 in humans (Fig. 1; Table 1), which has continued to this day. Since H3N2 virus has been associated with humans for nearly 50 years, it seems that 526R may be a previously unidentified adaptation marker of the polymerase complex of influenza virus with the potential to become established in a viral strain, and may act alone, or in coordination with other markers, to facilitate viral replication in humans.

PB2-526R enhances polymerase activity alone and with 672K. The influenza A virus viral polymerase complex, which is composed of PB1, PB2 and PA subunits, is used to replicate and transcribe the viral genome in the cell nucleus. We tested whether the presence of 526R, either alone or in combination with other adaptation markers, altered the ribonucleoprotein (RNP) complex polymerase activity of H7N9 virus. We first compared polymerase activity using a minigenome assay, testing all PB2 genotypes identified from H7N9 viruses for which sequence data had been publicly released or obtained from our analyses (Table 1). As expected, PB2 carrying 627K or 701N substitutions exhibited higher polymerase activity than PB2 derived from an avian type virus at both 33 and 37 °C in human embryonic kidney 293T (HEK293T) cells (Fig. 2a). Polymerase activity of the PB2 genotype with 526R was 10-fold higher than that of the avian-PB2 genotype, while two genotypes of PB2 carrying either 526R/627K or 526R/701N showed moderately increased and also

This study analyzes influenza sequences available from public databases and also those arising from our analysis of H7N9 isolates from human cases to identify new adaptation marker(s) in the PB2 gene. Various PB2 adaptation markers are identified among the H7N9 human isolates, and a K526R substitution associated with other previously defined adaptive markers is detected in some human isolates. Further characterization of the PB2-526R genotype reveals that H7N9 virus carrying this marker replicates to a significantly higher titre in mammalian cells, and also in mice. Furthermore, we show that nearly 80% of H5N1 human isolates from Indonesia and almost all seasonal H3N2 influenza A viruses isolated since 1970 contain PB2-526R. Reverse mutation of 526R to 526K attenuates polymerase activity in H3N2 and Indonesia-subclade H5N1 viruses. Therefore, in addition to 627K, 701N, 590S/591R and 591K, 526R is another marker of mammalian adaption by avian influenza viruses, possibly functioning through interaction between viral PB2 and nuclear export protein (NEP) during virus replication.
Table 1 | PB2 proposed mammalian adaptation markers among human and avian H5N1, H9N2, H7N9 and seasonal H3N2 isolates.*

<table>
<thead>
<tr>
<th>PB2 genotype</th>
<th>All H5N1</th>
<th>Indonesia H5N1</th>
<th>H9N2</th>
<th>H7N9</th>
<th>H3N2</th>
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<tr>
<td></td>
<td>Human1 (n = 261)</td>
<td>Avian1 (n = 1,290)</td>
<td>Human2 (n = 70)</td>
<td>Avian2 (n = 48)</td>
<td>Human3 (n = 8)</td>
</tr>
<tr>
<td>Proposed human adaptation marker</td>
<td></td>
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<tr>
<td>526R</td>
<td>47 (18%)</td>
<td>20 (27%)</td>
<td>47 (56%)</td>
<td>13 (27%)</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>590S</td>
<td>8 (4%)</td>
<td>3 (8%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>5 (12%)</td>
</tr>
<tr>
<td>599K</td>
<td>3 (1%)</td>
<td>0 (0%)</td>
<td>2 (0%)</td>
<td>0 (0%)</td>
<td>5 (12%)</td>
</tr>
<tr>
<td>627K</td>
<td>114 (43%)</td>
<td>279 (35%)</td>
<td>1 (0%)</td>
<td>0 (0%)</td>
<td>4 (12%)</td>
</tr>
<tr>
<td>701N</td>
<td>14 (5%)</td>
<td>2 (0%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>8 (12%)</td>
</tr>
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</table>
| 526R/627K | 8 (3%) | 3 (6%) | 6 (0%) | 0 (0%) | 2 (0%) | 0 (0%) | 0 (0%) | 24 (0.3%) | 2 | 8 (4.85%)
| Avian genotype** | 67 (26%) | 94 (38%) | 14 (20%) | 35 (37%) | 1 (12%) | 413 (67%) | 15 (16%) | 44 (94%) | 0 | 14 (0.3%) |

*Full-length PB2 sequences of seasonal H3N2, human and avian H5N1, H9N2 and H7N9 influenza A viruses from NCBI Influenza Virus Resource Database, and human HPAI H5N1 from Influenza Sequence Database were analysed. Right column under each Human/Avian represents total number and percentage of isolates containing adaptation marker.

†Five human H5N2 isolates contain 526R/590S and one has 591K/701N.
‡Two human H5N1 isolates contain 526R/591K; two contain 590S/627K; two are 590S/701N and one contains both 627K and 701N markers.
§Two avian H5N1 isolates have 627K/701N and five contain 590S/627K.
∥One human H9N2 isolate has triple adaptation markers; 526R/590S/701N.
¶Three avian H7N9 isolates contain 526R/591K and signature one has 590S/627K.
#Three human H7N9 isolates contain 526R/701N genotypes.
** Avian genotype indicates isolates that lack any proposed human adaptation markers in the PB2.

We found that replacement of PB1 in the SG57 RNP complex with A/Puerto Rico/8/1934 (PR8) PB1 resulted in decreased polymerase activity in repeated assays, suggesting introduction of PB1 requires compatibility with other subunits in the H2N2 RNP. However, incorporation of PB1 from Aichi68-H3N2 into the SG57 backbone, which mimics the formation of the H3N2 RNP as it occurred in 1968, does not compromise RNP polymerase activity. Introduction of 526R into the SG57-PB2 (627K) backbone enhanced polymerase activity. Furthermore, the RNP containing Aichi68-PB1 and 526R/627K PB2 consistently exerts higher polymerase activity than either whole SG57 or SG57 with Aichi68-PB1 in repeated experiments (Supplementary Fig. 2). Further studies are needed to unveil the role of PB2-526R in compatibility with PB1. Since 526R is predominantly present among H5N1 human isolates from Indonesia, we examined the effect of PB2-526R on polymerase activity in this subclade of the H5N1 subtype. It is notable that altering 526R to 526K in the PB2 completely abolishes RNP polymerase activity in the minigenome assay for RNP derived from A/Indonesia/5/2005 (Fig. 2d). These results clearly show that 526R, alone or coupled with 627K, provide an adaptive advantage to the polymerase complexes of avian H7N9 and H5N1 influenza A viruses during mammalian cell infection and optimizes RNP fitness during H3N2 virus replication.

PB2-526R augments viral genome transcription and replication. To further understand whether the altered RNP polymerase activity and viral growth associated with 526R versions of PB2 is due to differences in the efficiency of virus genome transcription and replication, we examined levels of viral mRNA and vRNA in A549 cells infected with viruses containing different versions of PB2 at 6 h post infection. Quantitative reverse transcription-PCR (RT-PCR) analysis with mRNA- or vRNA-specific primers showed that A549 cells infected with viruses carrying either 627K, 701N or 526R PB2 have significantly higher levels of viral matrix protein (M1) and nucleoprotein (NP) mRNA and vRNA, as compared with cells infected with a virus containing avian PB2 (Fig. 3), suggesting these adaptation markers are critical for viral RNP transcription and replication in mammalian cells during the early phase of infection. Furthermore, addition of 526R to 627K or 701N provides an extra advantage in viral RNP transcription and replication efficiency. While the additional enhancement effect of 526R on 627K or 701N is not as strong as that seen when 627K is introduced into the avian PB2 genotype, the experiments consistently higher (two- to fivefold greater) polymerase activity than that associated with the single PB2 substitutions 627K or 701N. Notably, the enhanced effect of 526R in combination with 627K or 701N provides an extra advantage in viral RNP transcription and replication efficiency. While the additional enhancement effect of 526R on 627K or 701N is not as strong as that seen when 627K is introduced into the avian PB2 genotype, the experiments...
were repeated four times, and consistently demonstrated that 526R exerts a positive effect on virus replication and transcription, either alone or when combined with 627K or 701N. It appears that the effect of 526R on viral genome transcription and replication is more apparent in the early hours of infection (Supplementary Fig. 3). These results suggest that 526R enhances the effects of 627K and 701N adaptations, resulting in optimal transcription and replication of influenza virus RNP in mammalian cells.

**526R synergizes with other PB2 markers in H7N9 replication.** To further characterize the phenotype of viruses carrying 526R PB2, reverse genetic (RG) versions of viruses from human isolates were constructed. These viruses were then tested for their ability to replicate in mammalian cells.

Figure 1 | Phylogenetic tree of H3N2 virus PB2. The PB2 tree was constructed using the neighbour-joining method with the Poisson model of amino-acid substitution; the reliability of the phylogenetic tree was determined by bootstrap analysis using 1,000 replicates. All sequences were obtained from Genbank. Viruses in red represent vaccine strains recommended by the World Health Organization.
carrying the 627K, 701N, 526R/627K, 526R/701N and solely 526R PB2 genotypes were constructed in the A/Zhejiang/DTID-ZJU01/2013 backbone. Comparison of the growth kinetics of these viruses with a similarly constructed RG version of an avian H7N9 strain (A/Chicken/Zhejiang/DTID-ZJU01/2013) revealed that viruses carrying any of the adaptation markers, including the novel 526R, replicated to a 1–2 log higher titre than the avian H7N9 virus in Madin-Darby canine kidney (MDCK) cells.
other H7N9 viruses that contain different PB2 adaptation markers were able to replicate to various levels in both HEK293T and A549 cells, with the 526R/627K version of the virus replicating to the highest levels (Fig. 4b,c). In contrast to the results obtained in MDCK, HEK293T and A549 cells, there is no replication advantage in avian cells (DF-1) for viruses containing either single or coupled 526R/627K PB2 mammalian adaptation markers, with the 526R/627K genotype being inhibited (Fig. 4d).

To exclude the possibility of underestimation of the growth of avian PB2 virus in the MDCK cell-based plaque assay, we also tested the titres of avian PB2 and avian 526R PB2 viruses by assessing the 50% egg infectious dose, with the results indicating that the plaque assay reflects the trend of virus titres obtained in the growth kinetics experiments. These results further confirmed that the 526R-526R substitution may enhance replication of H7N9 viruses in mammalian cells, but not in avian cells.

PB2-526R enhances H7N9 virus replication in mice. To further examine the effect of PB2-526R on virus replication, RG versions of H7N9 viruses representing the different PB2 genotypes identified in human isolates were tested in a mouse infection model. We first estimated the 50% mouse lethal dose (MLD50) of H7N9 viruses containing different genotypes of PB2. Our results showed that inclusion of K526R, alone or coupled with 627K or 701N, increased mortality in the infected mice (Fig. 5a). To confirm that the K526R substitution caused increased virus replication in vivo, mice were infected with a sub-lethal (2.25 × 10^4 plaque-forming units (PFU), based on pre-experimental data obtained using either 627K or 701N PB2 genotype viruses) dose of one of the different versions of H7N9 virus and were observed, and changes in body weight recorded over the following 14 days. Viruses carrying 526R or 627K substitutions caused moderate body weight loss (Fig. 5b,d), while RG avian H7N9 viruses or viruses with 701N alone did not cause any apparent loss of body weight in infected mice, as compared with the phosphate-buffered saline (PBS) control or avian-type PB2 virus-infected groups (Fig. 5c,d). Consistent with the results of the RNP polymerase activity and growth kinetics assays, the 526R/627K PB2 genotype virus caused the greatest body weight loss in mice, which peaked at around 25% on day 8 post infection (Fig. 5b). To examine virus replication ability in mouse lung tissues, three mice from each of the PB2-526R, 526R/627K, 627K, 526R/701N, 701N, avian-PB2, avian-PB2-526R and pure avian H7N9 (avian RG) infection groups were euthanized on day 3 post infection and lung tissues collected for virus titration. Consistent with the body weight loss result, viruses carrying 526R/627K replicated to the highest titres in the lungs of mice, about 1.5 to 2 logs higher than that in the lungs of mice infected with H7N9 virus carrying avian type PB2 and significantly higher than virus with 627K alone (Fig. 5b,d). While 701N PB2 virus caused no apparent body weight loss in mice, it replicated to a reasonably high titre in the lungs of infected mice and its replication ability was enhanced by the addition of 526R (Fig. 5c). H7N9 viruses carrying 526R PB2 also exhibited enhanced replication (about 10-fold) in the lung tissues of infected mice (Fig. 5d). Comparison of virus titres from mice infected with 526R/627K, 526R/701N or 526R viruses mirrored the results of the RNP activity and growth kinetics analyses, confirming that 526R/627K provides the most optimal PB2 genotype for replication of H7N9 virus in mammalian cells. These results confirm that the PB2-526R adaptation optimizes the in vivo replication ability of H7N9 viruses in mammalian host species.

PB2-526R confers a growth advantage in H3N2 virus. As described above, the H3N2 virus obtained K526R around 1970.
and the PB2-526R genotype immediately became the dominant circulating H3N2 in humans, persisting to the current day (Fig. 1). We have shown that addition of 526R to the PB2 of A/Hong Kong/1/1968 H3N2 virus enhances RNP polymerase activity, while back-mutation of 526R to 526K in the PB2 of A/Guangdong/ST798/2008 decreased RNP polymerase activity (Fig. 2b,c). To test whether gaining 526R has provided an additional growth advantage to the H3N2 virus, we constructed RG versions of the H3N2 virus and a variant with a 526K back-mutation in the A/Guangdong/ST798/2008 backbone. We adopted an assay that was previously used for measuring a minor mutation in the A/Guangdong/ST798/2008 backbone. We obtained RNP polymerase activity, while back-mutation of 526R to 526K in the PB2 of A/Guangdong/ST798/2008 decreased RNP polymerase activity (Fig. 2b,c). To test whether gaining 526R has provided an additional growth advantage to the H3N2 virus, we constructed RG versions of the H3N2 virus and a variant with a 526K back-mutation in the A/Guangdong/ST798/2008 backbone. We adopted an assay that was previously used for measuring a minor mutation in the A/Guangdong/ST798/2008 backbone. We obtained RG versions of H7N9 viruses containing avian-PB2, avian-PB2-526R, PB2-526R/627K, PB2-627K, PB2-526R/701N or PB2-701N, along with the remaining seven gene segments of A/Zeijing/DTD-ZJU01/2013, were used to infect (a) MDCK cells at an MOI of 0.001 and (b) HEK293T cells and (c) A549 cells at an MOI of 0.01. The cells were cultured at 37°C and supernatants collected at 24, 48 and 72 h post infection and subjected to plaque assays in MDCK cells to determine virus titres. A subset of reverse genetic versions of H7N9 viruses containing avian-PB2, avian-PB2-526R, PB2-526R/627K or PB2-627K or rescued reverse genetic A/Chicken/Zeijing/DTD-ZJU01/2013 (Avian RG), were used to infect DF-1 cells at an MOI of 0.01 and cells were then cultured at 39°C. Supernatants were collected at 24, 48 and 72 h post infection and subjected to plaque assays in MDCK cells to determine virus titres. The values displayed are the log10 means ± s.d. from three separate experiments. Statistical significance was analysed by one-way analysis of variance, corrected by the Bonferroni post-test: ***P < 0.001 and **P < 0.01.

PB2-526R optimizes interaction between PB2 and NEP. The molecular mechanism underlying host adaptation of influenza A virus through PB2 mutation remains largely undescribed.

Figure 4 | Growth kinetics of H7N9 viruses carrying different PB2 variants in mammalian and avian cells. Reverse genetic versions of H7N9 viruses containing avian-PB2, avian-PB2-526R, PB2-526R/627K, PB2-627K, PB2-526R/701N or PB2-701N, along with the remaining seven gene segments of A/Zeijing/DTD-ZJU01/2013, were used to infect (a) MDCK cells at an MOI of 0.001 and (b) HEK293T cells and (c) A549 cells at an MOI of 0.01. The cells were cultured at 37°C and supernatants collected at 24, 48 and 72 h post infection and subjected to plaque assays in MDCK cells to determine virus titres. A subset of reverse genetic versions of H7N9 viruses containing avian-PB2, avian-PB2-526R, PB2-526R/627K or PB2-627K or rescued reverse genetic A/Chicken/Zeijing/DTD-ZJU01/2013 (Avian RG), were used to infect DF-1 cells at an MOI of 0.01 and cells were then cultured at 39°C. Supernatants were collected at 24, 48 and 72 h post infection and subjected to plaque assays in MDCK cells to determine virus titres. The values displayed are the log10 means ± s.d. from three separate experiments. Statistical significance was analysed by one-way analysis of variance, corrected by the Bonferroni post-test: ***P < 0.001 and **P < 0.01. PFU, plaque-forming units; h.p.i., hours post infection.
Figure 5 | Infection and replication of H7N9 viruses with different PB2 genotypes in mice. (a) Groups of mice were infected with 25 μl inoculums containing 10^4, 10^5 or 10^6 PFU of reverse genetic versions of viruses, as specified. Mice were observed daily for changes in body weight for 14 days (day 0 to day 14). Animals that lost >25% of their pre-infection weight were euthanized, in accordance with our institutional animal ethics guidelines. The MLD50 values were calculated by the method described by Reed and Muench56. To test virus replication in mice, groups of nine BALB/c mice were each inoculated intranasally with 2.25 × 10^4 PFU of H7N9 recombinant virus containing PB2-526R/627K, PB2-627K, PB2-526R/701N, PB2-701N, avian-PB2, avian-PB2-526R or with a wholly reverse genetic version of avian H7N9 virus (avian RG). Results compare the mean percentage weight loss or gain for groups of mice infected with 627K or 526R/627K viruses or administered PBS (control) (b), 701N or 526R/701N viruses or PBS control (c) and avian RG, avian-PB2 or avian-PB2-526R viruses or PBS control (d); error bars represent s.d. The fractional number labels within the graphs indicate the number of surviving mice in each of the groups at the end of the experiment. Three mice from each of the groups infected with viruses carrying 627K or 526R/627K (a, right panel), 701N or 526R/701N (b, right panel) or with RG avian H7N9 virus, avian-PB2 or PB2-526R in the backbone of a human isolate (A/Zhejiang/DTID-ZJU01/2013) were killed at 72 h post infection. Lung tissues were removed and homogenated for estimation of virus replication by plaque assay. The values displayed represent the log_{10} mean titres ± s.d. Statistical significance was calculated by one-way analysis of variance, corrected by the Bonferroni post-test: **P < 0.01 and *P < 0.05. d.p.i., days post infection.

The viral NEP has been suggested to be involved in influenza virus host adaptation through its action as a regulator to switch virus genome transcription and replication37,38. Expression of NEP is regulated during virus replication, and increased expression of NEP brought about through disruption of splicing control is deleterious to virus replication37,39. We confirmed the interaction between PB2 and NEP in a co-immunoprecipitation assay using NEP and different versions of PB2 derived from H7N9 virus (Fig. 7a). To test the impact of co-expression of NEP on RNP polymerase activity, we co-expressed RNP containing various PB2 genotypes with increasing amounts of H7N9 virus NEP in HEK293T cells. Our result showed that increased levels of NEP exert an inhibitory effect and that this effect is much more profound for RNP with avian type PB2, but less so for RNP complexes containing PB2-526R, PB2-701N and PB2-627K. It is notable that when 526R is coupled with 627K in PB2, the resulting RNP complex is able to tolerate much higher levels of NEP (Fig. 7b). A similar inhibitory effect on RNP polymerase activity by avian type NEP has been observed for H5N1 viruses37. No adaptive substitutions have been observed in the NEP of H7N9 virus. Analysis of NEP expression in MDCK and A549 cells infected with H7N9 viruses carrying various PB2 markers confirmed that NEP is not expressed in the earliest hours of infection and that PB-526R-bearing viruses express higher levels of NEP than their counterparts that lack this substitution (Fig. 7c). Since PB2 interacts with both NP and PB1, we examined the PB2–NP interaction, and found that 526R enhances the interaction between PB2 and NP in the context of the RNP (Supplementary Fig. 5). While further studies are needed to reveal the molecular details of how PB2 and NEP interact with each other and host factors, it seems possible that the K526R-adaptive substitution enhances the interaction between PB2, the other RNP subunits and NEP during virus replication.

Discussion
Historically, very few influenza viruses have become established in mammalian hosts, namely the H1N1, H2N2 and H3N2 subtypes. Host adaptation requires the virus to overcome restriction barriers and replicate efficiently in a new host. To overcome host barriers, viruses must gain specificity for binding to cell receptors, adapt to use host machinery for genome replication, transcription and protein synthesis in cells and evade
restriction by elements of the host innate immune system during virus infection. The influenza virus replication complex is composed of three polymerase subunits (PB1, PB2 and PA) plus NP. Characterization of host adaptation markers among human isolates is important for recognizing potential for cross-species transmission in avian influenza A viruses. Adaptation of avian influenza A viruses in mammals has been extensively studied and the most well-characterized adaptation markers are located in the PB2 polymerase subunit. The PB2 E627K substitution is recognized as a dominant adaptation marker in the majority of human-adapted influenza A viruses, facilitating replication in mammalian cells, however, the 590S/591R motif has previously been characterized in the majority (~80%) of H5N1 human isolates from Indonesia.

Human infections with H7N9 virus emerged in China in March 2013 (refs 19,20,31). While efficient human to human transmission was not observed and a serological study found no evidence of circulation of H7N9 virus in the general population, sporadic inter-human transmissions were observed and laboratory transmission studies indicate that, unlike the H5N1 virus, this virus may be able to be transmitted between ferrets. Genetic analysis showed that the H7N9 virus is currently still of the avian genotype. In addition to the Q226L substitution in the HA, which may provide the virus with some ability to bind to human type receptors, what other distinct properties has this virus gained to make it different from other avian influenza viruses? This study found that isolates from human H7N9 cases carried one or more adaptation markers in the PB2 polymerase gene (Table 1). No similar association was found among avian or environmental H7N9 isolates, suggesting these adaptive changes occurred after virus replication in human cells. A distinct substitution, K526R, was found among some of the human isolates, in conjunction with 627K or 701N markers. The K526R substitution has not been recognized as a mammalian adaptation marker previously. However, sequence surveillance found that while 526R is not commonly present in human H5N1 isolates, 526R was found among some of the human isolates, in conjunction with 627K or 701N markers. It seems likely that different genetic lineages of avian influenza viruses evolve by using different adaptive pathways to enable their replication in humans. For example, the 2009 H1N1 virus adapted by using a PB2 590S/591R motif that synergized with the function of 627K to gain the ability to replicate in humans. A similar adaptive strategy was found among H9N2 human isolates with 7 out of 10 human isolates containing the 590S/591R motif.

Figure 6 | Comparison of the growth rate of H3N2 A/Guangdong/ST798/2008 strains carrying either 526R or 526K PB2 in mixed cultures. (a) MDCK cells or (b) A549 cells were infected at an MOI of 0.001 with a mixture (1:1) of wild-type (WT) (PB2-526R) and variant (PB2 R526K) A/Guangdong/ST798/2008 and sequentially passaged four times. At each passage viral RNA was isolated from cell culture supernatants at 48 h post infection, and the PB2 gene amplified by RT-PCR and sequenced. The genetic code for WT PB2-526R is AGA; variant PB2 526K is AAA. The DNA sequence chromatograms shown are from one of the three MDCK (a) and one of the four A549 (b) experiments in which 526R outgrew 526K. Experiments were repeated four times.

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The 526R PB2 mutation is rarely found among avian isolates and it was logical to presume that it may be an uncharacterized marker for human adaptation. Importantly, we found that the H3N2 virus has also gained the PB2-K526R substitution, in addition to 627K; this occurred around 1970, based on available sequence data, and this genotype has become prevalent in humans since then (Table 1; Fig. 1). It may be speculated that the introduction of an avian PB1 into the H2N2 virus in the 1968 reassortment event might have necessitated a requirement for rebalancing of RNP fitness in the H3N2 virus, with the combination of 526R/627K providing optimal efficiency for RNP polymerases during H3N2 virus infections in humans. Indeed, our results showed that PB2-526R/627K elicits higher RNP polymerase activity when combined with the PB1 from 1968 reassortant H3N2 virus than does PB2-627K (Supplementary Fig. 2). Introduction of 526R or 526K substitutions into A/Hong Kong/1/1968 (526K PB2) and A/Guangdong/ST798/2008 (526R PB2) PB2 proteins led to enhanced and reduced RNP polymerase activity, respectively, in minigenome assays (Fig. 2). Furthermore, the A/Guangdong/ST798/2008 version of H3N2 virus carrying 526R PB2 completely outgrows the 526K variant after only four passages in infected MDCK and A549 cells (Fig. 6). It remains to be tested whether the particularly high seasonal activity of H3N2 virus in humans, compared with H1N1, may be due to its more optimal replication properties and if gaining K526R has contributed to the fitness of H3N2 virus. Avian H5N1 virus has caused >600 human infections in over a dozen countries since 1997, and various adaptation markers in PB2 were observed among human isolates (Table 1). It was suggested that the effect of 627K on H5N1 virus may be dependent on the virus lineage. Interestingly, PB2 carrying 526R, but not 627K, is commonly found among H5N1 human isolates from Indonesia, which has the highest number of human H5N1 cases and the highest mortality rate to date. We showed that mutation of PB2-526R to 526K completely abolished polymerase activity for RNP complexes derived from one of the human isolates, A/Indonesia/5/2005 (Fig. 2d). Taken together, these results support the notion that K526R is a new adaptation marker for H7N9 and H5N1-(Indonesia) avian influenza A virus in humans and also provides an extra advantage for H3N2 virus replication. Protein structure data for the C-terminal domain of PB2, which contains the 267 and 701 residues, suggests that either 627K or 701N may enhance PB2 exposure to allow better interaction with elements of the host machinery such as importins, which facilitate nuclear import during virus replication.
Infection\textsuperscript{50,51}. There is no structural data available for analysing the domain that covers the region containing residue 526. How this region may enhance the fitness of PB2, or whether it interacts with residues 627 and 701 or other polymerase subunits, remains to be determined. Recent studies suggest that NEP has a role in regulating influenza A virus genome transcription and replication, independent to its nuclear export function\textsuperscript{37}. This study provides preliminary evidence that gaining the 526R substitution in the PB2, in addition to other adaptation markers, could enhance RNP polymerase activity through coordination with NEP expression for more efficient viral transcription and genome replication during influenza A virus infection.

Further studies are necessary to determine whether viruses that have gained dual 526R/627K or 526R/701N substitutions may transmit more efficiently in humans, or in animal models. Full evaluation of the transmission and replication abilities of H7N9 and other avian influenza A viruses, such as H5N1- (Indonesia), bearing K526R PB2 will provide valuable information for understanding the adaptation process and establish the importance of this new marker in surveillance of zoonotic influenza A viruses.

**Methods**

**Viruses.** All influenza viruses used in this study were rescued using the RGs technique to ensure the absence of contamination from other microbes. Viruses were propagated in embryonated chicken eggs (9–11 days old) for 3 days (P1), using the standard protocol approved by the Committee on the Use of Live Animals in Teaching & Research (CULATR) of the University of Hong Kong.

**Viral genome characterization and RGs.** The H7N9 viruses from laboratory-confirmed human cases have been characterized in our previous studies\textsuperscript{20,52}. The sequences of strains of H7N9 and H3N2 viruses used in this study have been deposited in Genbank (Accession codes AGU02226 to AGU02234, KJ633805, and KJ73719 to KJ73726). Eight segments from the A/Zhejiang/DTID-ZJU01/2013 strains were cloned into a pHW2000 plasmid system\textsuperscript{33,54}, to be used as backbones. RG versions of both A/Zhejiang/DTID-ZJU01/2013 (avian, ZJ/1, PB2-701N) and A/Chicken/Zhejiang/DTID-ZJU01/2013 (avian RG) were constructed. The ZJ/1 backbone was used to rescue recombinant viruses with various PB2 genes containing different adaptation markers. Minigenomes from the H3N2 strains A/Hong Kong/1/68 and A/Guangdong/ST798/2008, the H5N1 strain A/Indonesia/5/2005 and RG versions of A/Guangdong/ST798/2008 were cloned similarly and used to rescue virus together with 526R or 526R PB2 genes. Additional PB2 variants were generated using the QuikChange mutagenesis kit (Agilent) and also rescued using the ZJ/1 backbone. Full-length PB2 sequences of human and avian H5N1, H9N2, H7N7 and H7N9 viruses from the National Center for Biotechnology Information (NCBI) Influenza Virus Resource Database, together with all influenza A H7N9 human infections.

**Figure 7 | Effect of PB2-526R in interaction between PB2 and NEP.** (a) Co-immunoprecipitation of NEP with different versions of PB2 polymerase derived from H7N9 virus. Enhanced GFP (eGFP)-NEP and FLAG-PB2 were co-expressed in HEK293T cells. Whole-cell extracts were used for immunoprecipitation with an anti-GFP antibody and analysed by western blot analysis, using specific antibodies against eGFP and FLAG. The experiment was repeated three times. (b) Effect of different levels of NEP on RNP polymerase activity. RNP complexes containing different versions of PB2 were co-transfected with increasing amounts of NEP (H7N9) expression vector or empty vector, together with the firefly luciferase reporter pYH-Luci and a Renilla luciferase reporter (internal control). Luciferase activity was measured at 24 h post transfection, following incubation at 37 °C, and the effect of different amounts of NEP on the polymerase activity of RNP with and without PB2-526R was compared. Data represent mean luciferase activity from three separate experiments, calculated after normalization with Renilla luciferase activity, ± s.d. ‘—’ represents blank control; RNP without the PB2 gene. Statistical significance was analysed by the Student’s t-test: ***P<0.001, **P<0.01. (c) Western blot analysis of NEP expression in MDCK and A549 cells infected with H7N9 viruses carrying different versions of the PB2 segment. Confluent MDCK or A549 cells were infected with various reverse genetic H7N9 viruses, at an MOI of 1. Cells were collected at 5 and 7 h post infection and whole-cell lysates were analysed for NEP and NP expression by western blot using antibodies specific for NEP and NP, respectively. Blotting with antibody against β-tubulin, which served as a loading control, was also performed. Numbers indicate the ratio of NEP expression between viruses carrying 526R in the PB2 and those without 526R. The experiment was repeated twice. Full-size western blots are provided in Supplementary Fig. 6.
isolates in the Global Initiative on Sharing All Influenza Data (GISAID) database (http://gisaid.org) were analysed for prevalence of the K526R substitution.

**Minigene reporter assays.** Luciferase activity-based minigene reporter assays were performed as described previously53. RNP complexes composed of PA, PB1, PB2 and NP derived from A/Zhejiang/DTID-ZJU01/2013 and cloned into L-(tosylamido-2-phenyl) ethyl chloromethyl ketone (TAPI)-containing expression plasmid (pRL-construct) were transfected into HEK293T cells. After 24 h post transfection, RNP transfection activity was normalized against pRL-TK activity. To test the effect of NEP on RNP activity, experiments were performed as previously described57. The coding region of the RNP complex (PB1, PB2, PA and NP) were cloned separately into the pCI plasmid. HEK293T cells were transfected with the pCI-based RNP expression plasmids, the RNP expression plasmid and the two luciferase reporter plasmids, and RNP activity measured at 24 h post transfection.

**Growth kinetics of virus in cells.** Confluent MDCK cells were infected with RG viruses at a multiplicity of infection (MOI) of 0.001. The viral inoculums were removed after 1 h of adsorption at 37°C and replaced with minimal essential media (MEM) containing 10% FBS. Infected cells were incubated with 5% CO2. Culture supernatants were collected at different time points post infection and viral titres determined by the plaque assay in MDCK cells. Confluent DF-1 cells were infected with RG viruses at an MOI of 0.01. The viral inoculums were removed after 1 h adsorption at 39°C, then cells were washed with MEM and infected with MEM containing 5% CO2. Culture supernatants were collected at different time points post infection and viral titres determined by the standard 50% tissue culture infectious dose assay in MDCK cells.

**Replication in mice and lethal dose determination.** Female BALB/c mice, aged 4–6 weeks, were obtained from the Laboratory Animal Unit, the University of Hong Kong. For MDCK determination, groups of four or six mice were anesthetized with isoflurane (Halocarbon Laboratory), and intranasally inoculated with 25 μl of 10-fold serial dilutions of RG virus in PBS, the dose range being 103–105 PFU. Body weight and survival were monitored daily for 14 days after infection. The MDLD50 of RG viruses were calculated by the method of Reed and Muench58. To test virus replication in mice, groups of six or nine mice were anesthetized with isoflurane, and inoculated intranasally with 2.25 × 104–106 PFU. Body weight and survival were monitored daily for 14 days after infection.

**Immunoprecipitation assay.** To study the interaction between NEP and PB2, HEK293T cells were transfected with green fluorescent protein (GFP)-tagged NEP expression plasmid and FLAG-tagged PB2 expression plasmid. After 48 h, cells were lysed in cell lysis buffer (Tris-HCl 50 mM, 150 mM NaCl, 1% TritonX, pH 7.4). Cell debris was cleared by centrifugation, and 1 μl of GFP antibody (Invitrogen) was added to the cell lysate, which was then incubated for 3 h at 4°C. Thirty microliters of protein A sepharose beads (GE Health) were then added to the lysate, followed by incubation for 1 h at 4°C. The beads were washed with cell lysis buffer and finally boiled in SDS buffer at 95°C. Proteins were resolved with SDS-polyacrylamide gel electrophoresis and detected by western blot using anti-FLAG (dilution 1:5,000) (Sigma) antibodies. To study the interaction between NP and PB2 in RNP complexes, pCX plasmids expressing PB1, PA, NP and FLAG-tagged PB2 were co-transfected into HEK293T cells. The immunoprecipitation steps were performed as described above, except that a laboratory-made polyclonal NP antibody (dilution 1:3,000) was used.

**Statistical analysis.** Statistical analysis was carried out using GraphPad Prism V5 software (GraphPad Software Inc.).

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Author contributions
W.S. and P.W. performed the research, analysed and interpreted the results and wrote the manuscript. B.W.-Y.M., S.-Y.L., X.H., W.-L.W., M.Z., X.W., S.Y. and Y.C. performed animal experiments, virus isolation and analysed data. L.L. and K.-Y.Y. co-directed the study. H.C. directed the study, analysed and interpreted results and wrote the manuscript.

Additional information
Accession codes: Nucleotide sequences for the H7N9 and H3N2 viruses used in this study have been deposited in the NCBI GenBank database with accession codes AGU02226 to AGU02234, KJ633805, and KJ473719 to KJ473726.
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