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<th><strong>Title</strong></th>
<th>Materials and Methods for Prevention and Treatment of Viral Infections</th>
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<tr>
<td><strong>Inventor(s)</strong></td>
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Lau et al.

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ABSTRACT

The subject invention provides a novel and advantageous method for preventing and treating viral infection. Specifically exemplified herein are therapeutic uses of forsythoside A and jacaranone, compounds isolated from traditional Chinese medicinal material such as Fructus Forsythiae (Lian Qiao). Also provided is use of a Yin Qiao San composition for preventing and treating viral infection.
Milled powder of *Fructus Forsythiae*

- Suspended in 10x milli-Q H$_2$O
- Boiled for 2 hrs

Repeated twice

Residue
- Showed anti-viral effect

Water extract
- Evaporated to dryness
- Redissolved in MeOH

MeOH extract
- Showed anti-viral effect
- Purified by HPLC

Fr. 1  Fr. 2  ...  Fr. 4  Fr. 5  ...  Fr. 13

Fr. 1  Fr. 3  Fr. 4  ...  Fr. 6

Inhibition of COX-2
- Further purified by HPLC

Fr. 1  Fr. 2  Compound A1  Fr. 3

Fr. 1  Fr. 2  Compound D1

Inhibition of COX-2

FIG. 1
Table of contents:

- **FIG. 6A**
  - Compound D1

- **FIG. 6B**
  - Compound A1
FIG. 8A

Macrophage-H1N1

3.5E+04  2.9E+04  3.6E+03
3.0E+04  2.5E+04  
2.0E+04  1.5E+04  
1.0E+04  
5.0E+03
5.0E+03
0.0E+00

DMSO  Compound A1 (100ug/ml)

FIG. 8B

Macrophage-H9N2

9.1E+02  1.4E+02
8.0E+02
7.0E+02
6.0E+02
5.0E+02
4.0E+02
3.0E+02
2.0E+02
1.0E+02
0.0E+00

DMSO  Compound A1 (100ug/ml)

FIG. 8C

Macrophage-H3N2

6.1E+03  5.4E+02
5.0E+03
4.0E+03
3.0E+03
2.0E+03
1.0E+03
0.0E+00

DMSO  Compound A1 (100ug/ml)
FIG. 9A
**FIG. 9B**

![Graph](image)

Percentage of cell surrounding structures (%)

<table>
<thead>
<tr>
<th></th>
<th>DMSO</th>
<th>Compound A1 (100ug/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>8h</td>
<td></td>
<td></td>
</tr>
<tr>
<td>10h</td>
<td></td>
<td></td>
</tr>
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</table>

**FIG. 9C**

![Image](image)

h.p.i. M 2 4 9 24

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<tr>
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<th>DMSO</th>
<th>Compound A1</th>
</tr>
</thead>
<tbody>
<tr>
<td>M1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Actin</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

0.13 1.18 1.20 1.70 0.04 0.19 0.11 0.39
H9N2/G1

FIG. 11A

H9N2/G1

FIG. 11B
FIG. 12

FIG. 13A
FIG. 13B

FIG. 13C
FIG. 14

IL-1beta

FIG. 15A
**FIG. 15B**
MATERIALS AND METHODS FOR PREVENTION AND TREATMENT OF VIRAL INFECTIONS

CROSS-REFERENCE TO A RELATED APPLICATION

[0001] This application claims the benefit of U.S. provisional application Ser. No. 61/291,073 filed Dec. 30, 2009, which is incorporated herein by reference in its entirety.

BACKGROUND OF THE INVENTION

[0002] Viral infections are responsible for many acute and chronic life-threatening diseases.

[0003] It is estimated that about 33.4 million people are living with human immunodeficiency virus (HIV) worldwide. In addition, an estimated 2 billion people have been infected with hepatitis B virus, and 600,000 people die each year due to the acute or chronic consequences of the infection.

[0004] Influenza is an acute and one of the most widely spread viral infections worldwide.

[0005] Major influenza A pandemics include the Asian flu pandemic in 1957 (H2N2), the Hong Kong flu pandemic in 1968 (H3N2), the re-emergence of H1N1 (Russian flu) in 1970, the H5N1 bird flu in 1997 and 2003, and the most recent outbreak of the swine flu (H1N1) in April 2009. As of November 2009, worldwide more than 207 countries and territories or communities have reported laboratory confirmed cases of pandemic influenza H1N1, including at least 8768 deaths. In addition, the avian influenza H5N1 virus still presents a significant health concern. According to the World Health Organization, H5N1 results in a high mortality rate of more than 60% as per reported cases.

[0006] Despite extensive efforts, the development of effective anti-viral drugs has largely been empirical. There is no cure for HIV. Also, there is no specific treatment for acute hepatitis, although anti-viral treatments may improve the conditions of some patients with chronic hepatitis infection. In addition, almost all circulating strains of seasonal influenza A viruses are resistant to FDA approved antiviral drugs, including the adamantanes and neuraminidase inhibitors.

[0007] In previous research the current inventors have identified a series of potent inhibitors of the H5N1, H1N1 and H9N2 neuraminidases at a level comparable to the known neuraminidase inhibitor oseltamivir. They have also used bioactivity-guided purification technologies to analyze and identify the bioactive molecules responsible for anti-inflammatory activities in medicinal herbs including Panax ginseng and Cimicifuga species. While these advances are impressive, efforts to isolate and identify useful therapeutic compounds from natural sources remain difficult and largely empirical. Also, as virus strains change over time, the emergence of resistant mutations further diminishes the efficacy of anti-viral agents. Therefore, the development of additional novel anti-viral therapeutics is urgently needed.

BRIEF SUMMARY OF THE INVENTION

[0008] The present invention provides novel and advantageous materials and methods for preventing and/or treating viral infection. Specifically exemplified herein are therapeutic uses of forsythoside A and jacaranone, compounds isolated from traditional Chinese medicinal material such as Fructus Forsythiae (Lian Qiao). Also provided are herbal formulations, and compositions comprising the herbal formulation, for preventing and/or treating viral infection. Preferably, the compounds and compositions of the present invention are formulated for oral administration.

[0009] In one embodiment, the therapeutic methods of the subject invention can be used to prevent and/or treat viral infection. The method comprises administering, to a subject in need of such treatment, an effective amount of a composition comprising isolated compound A or a prodrug, metabolite or salt thereof, having the following formula:

![Chemical Structure 1]

wherein

[0010] R₁, R₂, and R₃ are, independently, —H, acyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkyl, alkoxy, hydroxyalkyl, carboxyl, carbalkoxy, or carboxamide.

[0011] In another embodiment, the method comprises administering, an effective amount of isolated prodrug, and/or metabolite of Compound A (or salt thereof), to a subject. An exemplified metabolite of Compound A is Compound B. The chemical structure of Compound B is shown as follows:

![Chemical Structure 2]

wherein

[0012] R₁, R₂, and R₃ are, independently, —H, alkyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkoxy, hydroxyalkyl, carboxyl, carbalkoxy, or carboxamide.

[0013] Another exemplified metabolite of Compound A is Compound C. The chemical structure of Compound C is shown as follows:

![Chemical Structure 3]
wherein

0014 \[ R''^n, R''^m \] are, independently, —H, acyl, halo, haloalkyl, amino, alkyaminooxy, hydroxyl, alkyl, alkoxy, hydroxyalkyl, carboxyl, carboxalkoxy, or carboxamide.

0015 In another embodiment, the method comprises administering, to a subject in need of such treatment, an effective amount of a composition comprising isolated compound D or a prodreg, metabolite or salt thereof, having the following formula:

![Chemical Structure](image)

wherein

0016 \[ R''''^n, R''''^m \] are, independently, —H, acyl, haloalkyl, alkyaminooxy, hydroxyl, alkyl, alkoxy, hydroxyalkyl, carboxyl, carboxalkoxy, or carboxamide.

0017 \[ R''''''^n, R''''''^m \] are, independently, —H, alkyl, halo, haloalkyl, amino, alkyaminooxy, hydroxyl, alkyl, alkoxy, hydroxyalkyl, carboxyl, carboxalkoxy, or carboxamide.

0018 The present invention also provides use of compounds of Formula A-D as medicaments for the prevention and/or treatment of viral infection. In one embodiment, the present invention provides uses of forsythoside A and jacarandone, and prodrugs, metabolites or salts thereof, as medicaments for the prevention and/or treatment of viral infections.

0019 In a further embodiment, the method comprises administering a composition comprising isolated compounds in a form, including but not limited to, a salt, stereoisomer, tautomer, crystalline, polymorph, amorphous, solvate, hydrate, ester, prodrug, metabolite, and any combination thereof.

0020 In a still further embodiment, the subject invention provides therapeutic methods for treating, preventing, or ameliorating a viral infection by administering an effective amount of the Yin Qiao Sun (YQS) composition, or a fraction thereof, to a subject. Also provided is the use of the Yin Qiao Sun composition, or a fraction thereof, as medicament for treating, preventing, or ameliorating viral infection.

0021 The present invention further embodies fractions of the Yin Qiao Sun composition, fractionated by reversed-phase HPLC method described herein. In a specific embodiment, the present invention provides one or more fractions F2-F5 of a methanol extract of the Yin Qiao Sun composition, whose HPLC chromatogram is shown in FIG. 12. In a specific embodiment, the present invention provides fraction F2 and/or F3 of the methanol extract of the Yin Qiao Sun composition, whose HPLC chromatogram is shown in FIG. 12. In a specific embodiment, the present invention provides fraction S11 of the Yin Qiao Sun composition, whose HPLC chromatogram is shown in FIG. 14.

0022 In one embodiment, the therapeutic methods of the subject invention can be used to prevent and/or treat viral infection caused by influenza virus A, B or C, including but not limited to, H1N1, H9N2, H3N2, H5N1, H2N2, H7N7, H7N1, and any combination thereof.

0023 In a further embodiment, the methods for preventing and/or treating viral infection further comprise: administering to a subject a second anti-viral agent selected from, for example, zanamivir, oseltamivir, peramivir, seltamivir, and any of the combination thereof; and wherein the second anti-viral agent can be administered prior to, simultaneously with, or subsequent to the administration of a compound of the subject invention.

0024 In a still further embodiment, the methods for preventing and/or treating viral infection further comprise administering to a subject, in addition to a compound of the present invention, a traditional Chinese medicinal material including, but not limited to, Fructus Forsythiae, Herba seu Flav Schizonepetae Tenuifoliae, Fructus Arctii Lappae, Semen Sojae Preparatum, Herba Lophatheri Gracilis, Radix Glycyrrhizae Uralensis, and Radix Platycodi Grandiflori.

0025 In a still further embodiment, the methods for treating viral infection further comprise a step of determining the level of viral infection in a subject, wherein the determination is optionally made at multiple times to monitor the change over time.

0026 In another further embodiment, the methods for treating viral infection further comprise a step of determining the level of one or more bio-markers in a subject, wherein the determination is optionally made at multiple times to monitor the change over time.

BRIEF DESCRIPTION OF THE FIGURES

0027 FIG. 1 shows an extraction scheme of bioactive compounds including forsythoside A (Compound A1) and jacarandone (Compound D1) from Fructus Forsythiae. Fructus Forsythiae is milled and extracted with 10x of milli-Q water under reflux and boiled for 2 hours. The extraction process is repeated once. Each time, the resulting extract is collected, combined, evaporated to dryness, and redissolved in MeOH. Using a bioassay guided isolation scheme, the fractions showing significant anti-viral and/or COX-2 suppressive effects are further purified using the reversed-phase high-performance liquid chromatography (HPLC) until pure compounds are obtained.

0028 FIGS. 2A-B show reverse-phase high performance liquid chromatography (HPLC) chromatograms (A) and UV absorbance (B) of forsythoside A (Compound A1). Compound A1 is purified by reversed-phase HPLC using gradient elution from 10% to 90% of acetonitrile at a flow rate of 1 mL/min. FIG. 2A shows that a single peak is detected using Photo-diode Array detector at 254, 210 and 280 nm. Compound A1 elutes at approximately 6.8 min. FIG. 2B shows that the UV absorbance of Compound A1 is maximized at 210, 290 and 330 nm.

0029 FIGS. 3A-B show reverse-phase high performance liquid chromatography (HPLC) chromatograms (A) and UV absorbance (B) of jacarandone (Compound D1). Compound D1 is purified by reversed-phase HPLC using gradient elution from 10% to 90% of acetonitrile at a flow rate of 1 mL/min. FIG. 3A shows that a single peak is detected using Photo-diode Array detector at 254, 210 and 280 nm. Compound D1 is eluted at approximately 10.75 min. FIG. 3B shows that UV absorbance of Compound D1 is maximized at 230 nm.

0030 FIG. 4 shows the 1H (upper panel) and 13C (lower panel) chemical shifts of forsythoside A (Compound A1). The structure of Compound A1 is elucidated by a Bruker 500 MHz
DRX NMR spectrometer, operating at 500 MHz for H and at 125 MHz for 13C NMR, using methanol-d with TMS as the solvent.

[0031] FIG. 5 shows the H (upper panel) and 13C (lower panel) chemical shifts of jacraronine (Compound D1). The structure of Compound D1 is elucidated by a Bruker 500 MHz DRX NMR spectrometer, operating at 500 MHz for H and at 125 MHz for 13C NMR, using methanol-d with TMS as the solvent.

[0032] FIG. 6 shows the chemical structures of forsythoside A and jacarane.

[0033] FIGS. 7A-E show inhibitory effects of forsythoside A (Compound A1) or oseltamivir on influenza virus replication. MDCK cells are infected with influenza viruses (A) H1N1 (A/HK/154/98), (B) H1N1-R (A/Victoria/07159200/ 07), (C) H9N2 (A/Quail/HK/G1/97) or (D) H3N2 (A/H3N2/1174/99). After 48-hour incubation with forsythoside A (Compound A1) (100 μg/ml), viral titters (TCID50) in cell culture supernatant are measured by titration in MDCK cells.

[0034] FIG. 8 shows inhibitory effects of forsythoside A (Compound A1) on influenza virus replication in primary human blood macrophages. Primary human blood macrophages are infected with human influenza viruses including H1N1 (A/HK/54/98); H9N2 (A/Quail/HK/G1/97); and H3N2 (A/H3N2/1174/99) at an m.o.i. of 2. Cells are then incubated with forsythoside A (Compound A1) (100 μg/ml) or DMSO for 48 hours. After that, cell culture supernatants are collected and viral titters (TCID50) are measured by titration in MDCK cells. (All figures are representative results from three experiments.)

[0035] FIGS. 9A-C show inhibitory effects of forsythoside A (Compound A1) on influenza virus as determined by immunocytochemistry (A-B) and Western blot analysis (C). (A) MDCK cells are infected with human influenza virus H9N2/G1 (A/Quail/HK/G1/97) at an m.o.i. of 0.2. Cells are then incubated with Compound A1 (100 μg/ml) or DMSO for 6 or 10 hours. After that, cells are fixed. Influenza nucleoprotein and M1 protein are stained with the Influenza Detection Kit. (B) The numbers of cells expressing viral proteins at 6 and 10 hours post-infection are counted and percentages of cells expressing viral proteins are calculated. (C) MDCK cells are infected with human influenza virus H9N2/G1 (A/Quail/HK/G1/97) at an m.o.i. of 2 or mock infected (M). After that, cells are then incubated with forsythoside A (Compound A1) (100 μg/ml) or DMSO. Proteins are harvested at 2, 4, 9 or 24 hour post-infection. Expressions of the influenza M1 protein are analyzed by Western blot analysis. Actin is used as a loading control. A & C, representative figures from three experiments; B: n = 3; *P<0.05.

[0036] FIGS. 10A-D show the ultrastructural examination of virions by transmission electron microscopy. MDCK cells are infected with human influenza virus H9N2/G1 (A/Quail/ HK/G1/97) at an m.o.i. of 2. Cells are then incubated with forsythoside A (Compound A1) (100 μg/ml) (C-D) or DMSO (A-B) for 18 hours. Cells are then fixed and ultrastructures are examined by transmission electron microscopy. Arrows in (B) and (D) indicate influenza virions. Magnifications: A, 32000x; B, 63000x; C, 48000x; D, 89000x.

[0037] FIGS. 11A-B show inhibitory effects of jacraronine (Compound D1) on cyclooxygenase 2 (COX-2). (A) Primary human blood macrophages are infected with H9N2/G1 (A/Quail/HK/G1/97) at an m.o.i. of 2. Cells are then incubated with Compound D1 (10 μg/ml or 20 μg/ml) or DMSO for 3 hours. After that, total RNA are harvested and COX-2 mRNA levels are examined by TaqMan Gene Expression assays (n=5). (B) Cells are either infected with H9N2/G1 (A/Quail/HK/G1/97) at an m.o.i. of 2 or mock-infected. After that, cells are treated with Compound D1 (10 μg/ml or 20 μg/ml) for 48 hours. Cells culture supernatants are then harvested and prostaglandin E2 levels are measured by the Prostaglandin E2 ELISA Kit (n=4). *P<0.05; **P<0.01; ***P<0.005.

[0038] FIG. 12 shows reverse-phase high performance liquid chromatography (HPLC) chromatogram of Yin Qiao San (YQS) formula. The fractions are separated by reversed-phase HPLC using gradient elution from 10% to 90% of acetonitrile at a flow rate of 1 mL/min. Total five fractions are obtained.

[0039] FIGS. 13A-C show inhibitory effect of YQS and YQS fractions on virus replication. MDCK cells are infected with H1N1 (A/HK/54/98) (A) or H9N2/G1 (A/Quail/HK/ G1/97) (B) at an m.o.i. of 2. Cells are then incubated with YQS (100 μg/ml) or DMSO for 48 hours. After that, cell culture supernatants are collected and viral titters (TCID50) are measured by titration in MDCK cells. (A-B) two (C) independent experiments.

[0040] FIG. 14 shows a typical chromatogram of YQS obtained by reverse-phase high performance liquid chromatography (HPLC). The fractions are separated by reversed-phase HPLC using gradient elution from 10% to 90% of acetonitrile at a flow rate of 1 mL/min. Total thirteen fractions are obtained.

[0041] FIGS. 15A-B show YQS and YQS fractions induce interleukin (IL)-1β. (A) Primary human blood macrophages are infected with H1N1 (A/HK/54/98) at an m.o.i. of 2 or mock infected. H1N1-infected cells are then incubated with YQS (100 μg/ml) or DMSO for 3 hours. After that, total RNA are harvested and IL-1β mRNA levels are examined by TaqMan Gene Expression assays. (B) Primary human blood macrophages are infected with H1N1 (A/HK/54/98) at an m.o.i. of 2 or mock infected. H1N1-infected cells are then incubated with YQS fractions (S1-S13) (100 μg/ml) or DMSO for 3 hours. After that, total RNA are harvested and IL-1β mRNA levels are examined by TaqMan Gene Expression assays. Representative figures from three experiments.

DETAILED DESCRIPTION OF THE INVENTION

[0042] The subject invention provides novel and advantageous materials and methods for preventing, treating and/or ameliorating viral infection in a subject. Specifically exemplified herein are therapeutic uses of forsythoside A and jacarane, compounds isolated from traditional Chinese medicinal material such as Fructus Forsythiae (Lar Qiao). Also provided are herbal formulations, and compositions thereof, for preventing and/or treating viral infection. Preferably, the compounds and compositions of the present invention are formulated for oral administration.
In one embodiment, the method comprises administering, an effective amount of isolated Compound A or a prodrug, metabolite or salt thereof, to a subject. The chemical structure of Compound A is shown as follows:

wherein

R<sub>1</sub>-R<sub>7</sub> are, independently, —H, acyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkyl, alkoxy, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide.

In another embodiment, the method comprises administering, an effective amount of isolated prodrug and/or metabolite of Compound A (or salt thereof), to a subject. An exemplified metabolite of Compound A is Compound B. The chemical structure of Compound B is shown as follows:

wherein

R<sub>1</sub>-R<sub>6</sub> are, independently, —H, acyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkoxy, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide.

Another exemplified metabolite of Compound A is Compound C. The chemical structure of Compound C is shown as follows:

wherein

R<sub>1</sub>′-R<sub>6</sub>′ are, independently, —H, acyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkoxy, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide.

In another embodiment, the method comprises administering, to a subject in need of such treatment, an effective amount of a composition comprising isolated compound D or a prodrug, metabolite or salt thereof, having the following formula:

wherein

R<sub>n</sub>′-R<sub>m</sub>′ are, independently, —H, acyl, haloalkyl, alkylamino, alkyl, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide, and

R<sub>n</sub>-R<sub>m</sub> are, independently, —H, alkyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkoxy, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide.

In an embodiment, C—R<sub>n</sub> forms an ether or ester bond. In another embodiment, R<sub>n</sub>′-R<sub>m</sub>′ are, independently, —H, alkyl, halo, haloalkyl, amino, or hydroxylalkyl.

In a further embodiment, the subject invention provides methods for treating, preventing, or ameliorating viral infection by administering, an effective amount of any of a combination of Compound A, Compound B, Compound C, and Compound D (and prodrugs, metabolites and salts thereof), to a subject.

The present invention also includes the use of compounds of Formula A-D as medicaments for the prevention and/or treatment of viral infections. In one embodiment, the present invention provides use of forsythoside A and jacarandone, and prodrugs, metabolites or salts thereof, as medicaments for the prevention and/or treatment of viral infections.

“Alkyl” means linear saturated monovalent radicals of one to eight carbon atoms or a branched saturated monovalent of three to eight carbon atoms. It may include hydrocarbon radicals of one to four or one to three carbon atoms, which may be linear. Examples include methyl, ethyl, propyl, 2-propyl, n-butyl, iso-butyl, tert-butyl, pentyl, and the like.

“Acyl” means a radical —C(O)R where R is, for example, hydrogen, alkyl or cycloalkyl, heterocycloalkyl, halo, or alkyl halo. Examples further include formyl, acetyl, ethylcarbonyl, and the like.

“Carbonyl” means the radical —C(O)OH.

“Carboxalkoxy” means a radical —C(O)R where R is, for example, hydrogen, alkyl or cycloalkyl, heterocycloalkyl, halo, or alkyl halo.

“Carboxamide” means a radical —C(O)R, where R is, for example, —NH<sub>2</sub> or Alkylamino.

“Halo” means fluoro, chloro, bromo, or iodo, such as bromo and chloro.
[0061] “Halooalkyl” means alkyl substituted with one or more, same or different, halogen atoms, e.g., —CH₂Cl, —CH₂Br, —CF₃, —CH₂CH₂Cl, —CH₂CCl₂, and the like.

[0062] “Amino” means the radical —NH₂.

[0063] “Alkylaminoo” means a radical —NHR or —NR₂, where each R is, independently, an alkyl group. Examples include methylaminoo, (1-methylethyl)aminoo, dimethylaminoo, methylethyleminoo, di(1-methylethyl)aminoo, and the like.

[0064] “Hydroxy” means the radical —OH.

[0065] “Hydroxyalkyl” means an alkyl radical as defined herein, substituted with one or more, preferably one, two or three, hydroxy groups. Representative examples include, but are not limited to, hydroxymethyl, 2-hydroxyethyl, 2-hydroxypropyl, 2-hydroxypropyl, 1-(hydroxymethyl)-2-methylpropyl, 2-hydroxybutyl, 2-hydroxyethyl, 2-hydroxypropyl, 2-hydroxypropyl, 1-(hydroxymethyl)-2-methylpropyl, preferably 2-hydroxyethyl, 2,3-di-hydroxypropyl and 1-(hydroxymethyl)2-hydroxyethyl.

[0066] “Alkoxy” means the radical —OR, where R is an alkyl group. Exemplary alkoxy groups include methoxy, ethoxy, propoxy, and die, like.

[0067] The subject invention further provides methods for preventing, treating, or ameliorating viral infections by administering a composition comprising isolated enantiomeric compounds. The isolated enantiomeric forms of the compounds of the invention are substantially free from one another (i.e., in enantiomeric excess). In other words, the “R” forms of the compounds are substantially free from the “S” forms of the compounds and are, thus, in enantiomeric excess of the “S” forms. Conversely, “S” forms of the compounds are substantially free of “R” forms of the compounds and are, thus, in enantiomeric excess of the “R” forms. In one embodiment of the invention, the isolated enantiomeric compounds are at least about 80% enantiomeric excess. In a preferred embodiment, the compounds are in at least about 90% enantiomeric excess. In a more preferred embodiment, the compounds are in at least about 95% enantiomeric excess. In an even more preferred embodiment, the compounds are in at least about 97.5% enantiomeric excess. In a most preferred embodiment, the compounds are in at least about 99% enantiomeric excess.

[0068] The subject invention further provides methods for preventing, treating, or ameliorating viral infections by administering a composition comprising isolated compounds in a form, including but not limited to, a salt, stereoisomer, tautomer, crystalline, polymorph, amorphous, solvate, hydrate, ester, prodrug, metabolite, and any combination thereof.

[0069] The term “prodrug,” as used herein, refers to a metabolic precursor of a compound of the present invention or pharmaceutically acceptable form thereof. In general, a prodrug comprises a functional derivative of a compound, which may be inactive when administered to a subject, but is readily convertible in vivo into an active metabolite compound.

[0070] Conventional procedures for the selection and preparation of suitable prodrug derivatives are described, for example, in “Design of Prodrugs”, ed. H. Bundgaard, Elsevier, 1985. Preferably, a prodrug of the present invention enhances desirable qualities of the compound of the present invention, including not limited to, solubility, bioavailability, and stability. Hence, the compounds employed in the present methods may, if desired, be delivered in a prodrug form. Prodrugs of the compounds employed in the present invention may be prepared by modifying functional groups present in the compound such that the modifications are cleaved, either in route manipulation or in vivo, to the parent compound.

[0071] The term “metabolite,” refers to a pharmacologically active product, including for example, an active intermediate or an ultimate product, produced through in vivo metabolism of a compound of the present invention in a subject. A metabolite may result, for example, from the anabolic and/or catabolic processes of the administered compound in a subject, including but not limited to, the oxidation, reduction, hydrolysis, amidation, deamidation, esterification, deesterification, enzymatic cleavage, and the like.

[0072] Metabolites are typically identified by preparing a radiolabelled (e.g., 14C or 3H) isotope of a compound of the present invention, administering it parenterally in a detectable dose (e.g., greater than about 0.5 mg/kg) to an animal such as rat, mouse, guinea pig, monkey, or to a human, allowing sufficient time for metabolism to occur (typically about 30 seconds to about 30 hours), and isolating its conversion products from the urine, blood or other biological samples. These products are easily isolated since they are labeled (others are isolated by the use of antibodies capable of binding epitopes surviving in the metabolite). The structure of metabolites can be determined in conventional fashion, e.g., by MS, LC/MS or NMR analysis. In general, analysis of metabolites is performed according to techniques well known to those skilled in the art of drug metabolism studies.

[0073] The term “subject,” as used herein, describes an organism, including mammals such as primates, to which treatment with the compositions according to the present invention can be provided. Mammalian species that can benefit from the disclosed methods of treatment include, but are not limited to, apes, chimpanzees, orangutans, humans, monkeys; and domesticated animals such as dogs, cats, horses, cattle, pigs, sheep, goats, chickens, mice, rats, guinea pigs, and hamsters.

[0074] The term “antiviral,” as used herein, includes but is not limited to, preventing, inhibiting, suppressing, reducing, adversely impacting, and/or interfering with the growth, survival, replication, function, and/or dissemination of a virus.

[0075] In one embodiment, the subject invention provides therapeutic methods for treating, preventing, or ameliorating viral infection by administering an effective amount of the compounds of the subject invention.

[0076] In a specific embodiment, the subject invention provides therapeutic methods for treating, preventing, ameliorating viral infection by administering an effective amount of forsythoside A (Compound A1) or a prodrug, metabolite or salt thereof, to a subject. The chemical structure of forsythoside A (Compound A1) is: 

[0077] In another specific embodiment, the subject invention provides therapeutic methods for treating, preventing, and/or ameliorating viral infection by administering, an
effective amount of jacarandine (Compound D1) or a prodrug, metabolite or salt thereof, to a subject. The chemical structure of jacarandine (Compound D1) is:

\[ \text{Chemical structure of jacarandine (Compound D1)} \]

In a further embodiment, the subject invention provides therapeutic methods for treating, preventing, or ameliorating viral infection by administering an effective amount of any of a combination of forsythoside A (Compound A1), Compound B1, Compound C1, and Compound D1 (and prodrugs, metabolites or salts thereof) to a subject.

For another specific embodiment, the subject invention provides therapeutic methods for treating, preventing, or ameliorating viral infection by administering an effective amount of prodrug and/or metabolite of Compound A1 (or salt thereof), to a subject. An exemplified metabolite of Compound A1 is Compound B1. The chemical structure of Compound B1 is:

\[ \text{Chemical structure of Compound B1} \]

Another exemplified metabolite of Compound A1 is Compound C1. The chemical structure of Compound C1 is:

\[ \text{Chemical structure of Compound C1} \]

In one embodiment, the composition comprises an herbal formula consisting of herbs having relative weight amounts as indicated in the parenthesis: 3% Liangqiao (FFS) (Fructus Forsythiae suspensae), 2% Jin yin hua (FL2) (Lonicerae Japonicae), 2% Niu bang zi (FAL) (Fructus Arctii Lappaceae), 2% Jie geng (RPG) (Platycodi Grandiflori), 2% Ba He (Herba Menthae haplocalycis), 1% Dan Dou Chi (Semen Sojae Preparatum), 1% Niu Bang Zi (FAL) (Fructus Arctii Lappaceae), 1% Jie geng (RPG) (Platycodi Grandiflori), and 1% Dan Dou Chi (Semen Sojae Preparatum)

In one embodiment, the composition comprises an herbal formula consisting of herbs having relative weight amounts as indicated in the parenthesis: 2% Liangqiao (FFS) (Fructus Forsythiae suspensae), 2% Jin yin hua (FL2) (Lonicerae Japonicae), 2% Niu bang zi (FAL) (Fructus Arctii Lappaceae), 2% Jie geng (RPG) (Platycodi Grandiflori), 2% Ba He (Herba Menthae haplocalycis), 1% Dan Dou Chi (Semen Sojae Preparatum), 1% Niu Bang Zi (FAL) (Fructus Arctii Lappaceae), 1% Jie geng (RPG) (Platycodi Grandiflori)

Table 1 illustrates embodiments of the herbal formulations of the subject invention.

<table>
<thead>
<tr>
<th>Herbs</th>
<th>YQS-F1</th>
<th>YQS-F2</th>
<th>YQS-F3</th>
<th>YQS-F4</th>
<th>YQS-F5</th>
<th>YQS-F6A</th>
<th>YQS-F9 Preferred</th>
<th>YQS-Traditional</th>
</tr>
</thead>
<tbody>
<tr>
<td>Liangqiao</td>
<td>32%</td>
<td>33%</td>
<td>23.5%</td>
<td>22.2%</td>
<td>30.8%</td>
<td>30%</td>
<td>30%</td>
<td>22%</td>
</tr>
<tr>
<td>Jin yin hua</td>
<td>32%</td>
<td>17%</td>
<td>23.5%</td>
<td>22.2%</td>
<td>30.8%</td>
<td>20%</td>
<td>30%</td>
<td>22%</td>
</tr>
<tr>
<td>Niu bang zi</td>
<td>19%</td>
<td>33%</td>
<td>23.5%</td>
<td>22.2%</td>
<td>15.4%</td>
<td>20%</td>
<td>5%</td>
<td>9%</td>
</tr>
<tr>
<td>Jie geng</td>
<td>—</td>
<td>—</td>
<td>1.4%</td>
<td>2.7%</td>
<td>1.8%</td>
<td>4%</td>
<td>5%</td>
<td>9%</td>
</tr>
</tbody>
</table>

Table 1: Percentage Distribution of herbal constituents in Ying Qiao San (YQS)
<table>
<thead>
<tr>
<th>Herbs</th>
<th>YQ8-F1</th>
<th>YQ6-F2</th>
<th>YQ8-F3</th>
<th>YQ8-F4</th>
<th>YQ8-F5</th>
<th>YQ8-F6A</th>
<th>YQ8-F9</th>
<th>YQ8-Traditional</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bo he (HG)</td>
<td>— —</td>
<td>1.4%</td>
<td>2.7%</td>
<td>1.8%</td>
<td>4%</td>
<td>5%</td>
<td>9%</td>
<td></td>
</tr>
<tr>
<td>Dan dou chi (SSP)</td>
<td>— —</td>
<td>1.2%</td>
<td>2.2%</td>
<td>1.5%</td>
<td>4%</td>
<td>5%</td>
<td>7.5%</td>
<td></td>
</tr>
<tr>
<td>Guancao (BGU)</td>
<td>16%</td>
<td>17%</td>
<td>23.5%</td>
<td>22.2%</td>
<td>15.4%</td>
<td>30%</td>
<td>10%</td>
<td>7.5%</td>
</tr>
<tr>
<td>Dan zha ye (HIG)</td>
<td>— —</td>
<td>0.9%</td>
<td>1.8%</td>
<td>1.2%</td>
<td>4%</td>
<td>5%</td>
<td>6%</td>
<td></td>
</tr>
<tr>
<td>Jing jie (HST)</td>
<td>— —</td>
<td>0.9%</td>
<td>1.8%</td>
<td>1.2%</td>
<td>4%</td>
<td>5%</td>
<td>6%</td>
<td></td>
</tr>
<tr>
<td>Anti-viral effects (% of suppression)</td>
<td>85%</td>
<td>&lt;50%</td>
<td>&lt;50%</td>
<td>~50%</td>
<td>80%</td>
<td>85%</td>
<td>~80%</td>
<td></td>
</tr>
</tbody>
</table>

[0087] Total Dry Weight in Traditional YQ8 Decoction is 66 g.

[0088] Advantageously, the compositions of the subject invention provide surprisingly superior anti-viral effects. Specifically, the compositions of the subject invention enhance IL-1β level in a subject who has viral infection.

[0089] In an embodiment, the subject invention provides therapeutic methods for treating, preventing, or ameliorating viral infection by administering an effective amount of the composition of the subject invention, or a fraction thereof, to a subject. Also provided is use of the composition of the subject invention, or a fraction thereof, as a medicament for treating, preventing, or ameliorating viral infection.

[0090] In a still further embodiment, the subject invention provides therapeutic methods for treating, preventing, or ameliorating viral infection by administering an effective amount of the Yin Qiao San (YQS) composition (YQS-traditional formulation as shown in Table 1), or a fraction thereof, to a subject. Also provided is use of the Yin Qiao San composition, or a fraction thereof, as a medicament for treating, preventing, or ameliorating viral infection.


[0092] In a specific embodiment, the Yin Qiao San composition consists of herbs having relative weight amounts as indicated in the parenthesis: *Herba menthae haplocalycis* (about 60), *Herba sen Flos Schizonetiae Tenuifoliae* (about 40), *Fructus forsythiae suspenesae* (about 100), *Fructus Arctii Lappae* (about 60), *Semen Sojae Preparatum* (about 50), *Herba Lophatheri Gracilis* (about 40), *Radix Glycyrrhizae Uralensis* (about 50), *Flos Lonicerae Japonicae* (about 100) and *Radix Platycodi Grandiflori* (about 60).

[0093] In one embodiment, the present invention provides fractions of the Yin Qiao San composition, fractionated by reversed-phase HPLC (LiChrospher 100 RP C18 EC 5 μm, 250 x 4.6 mm ID) using a gradient elution from 10% acetonitrile (CH₂CN) to 90% CH₂CN at a flow rate of 1 mL/min. Peak detection is achieved using an Agilent 1200 series of fast scanning Photo-diode Array detector set at 210, 254 and 280 nm.

[0094] In a specific embodiment, the present invention provides one or more fractions F2-F5 of the Yin Qiao San composition, whose HPLC chromatograph is shown in FIG. 12. In a specific embodiment, the present invention provides fraction F2 and/or F3 of the Yin Qiao San composition, whose HPLC chromatograph is shown in FIG. 12. In a specific embodiment, the present invention provides fraction S11 of the Yin Qiao San composition, whose HPLC chromatograph is shown in FIG. 14.

[0095] The term “treating,”’ as used herein, includes but is not limited to, reducing, suppressing, inhibiting, lessening, or affecting the progression, severity, and/or scope of a condition, chance of re-occurrence or returning of a disease after a remission. In one embodiment, treating may include directly affecting or curing, suppressing, inhibiting, reducing the severity of, delaying the onset of, reducing symptoms associated with an infection, or a combination thereof. In another embodiment, treating includes delaying progression, expediting remission, inducing remission, augmenting remission, speeding recovery, increasing efficacy of or decreasing resistance to alternative therapeutics, or a combination thereof.

[0096] The term “suppressing” or “inhibiting,” as used herein, includes but is not limited to, reducing the severity of symptoms, reducing the severity of an acute episode, reducing the number of symptoms, reducing the incidence of disease-related symptoms, reducing the latency of symptoms, ameliorating symptoms, reducing secondary symptoms, reducing secondary infections, prolonging patient survival, or a combination thereof.

[0097] The term “preventing,” as used herein, includes but is not limited to, delaying the onset of symptoms, preventing relapse to a disease, decreasing the number or frequency of relapse episodes, increasing latency between symptomatic episodes, or a combination thereof. The term “effective amount,” as used herein, refers to an amount that is capable of preventing, ameliorating, or treating viral infection. The term “effective amount” also includes an amount that is capable of preventing, ameliorating, or treating viral infections and/or inhibiting or reducing the level of COX-2 and/or PGE2. In certain embodiments, the effective amount enables at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 60%, 70%, 80%, 90%, or 100% decrease in viral titers (e.g., TCID₅₀). The decrease in viral titers can be determined from, for example, biological samples obtained from a subject at different time points.
Advantageously, the compounds and compositions of the subject invention are capable of preventing, inhibiting, reducing and/or retarding viral replication. In specific embodiments, the replication of influenza viruses H1N1 (A/HK/54/98), H1N1 (A/Victoria/0715/2000), H9N2 (A/Quail/HK/G1/97) and H3N2 (A/H3N2/1174/99) in MDCK cells is suppressed by forsythoside A (Compound A1) for about 15-, 3-, 24- and 12-fold, respectively.

In addition, the compounds and compositions of the subject invention are useful for modulating immune responses during viral infection. For instance, the compounds and compositions of the subject invention enhance IL-1β levels. The compounds and compositions of the subject invention can also inhibit COX-2 and prostaglandin E2 (PGE2) activity, particularly during viral infection. In another embodiment, the novel and advantageous therapeutic method is capable of preventing, treating, or ameliorating viral infection of a subject resistant to existing anti-viral therapies, including but not limited to, compounds such as adamantanes and neuraminidase inhibitors, zanamivir, oseltamivir, peramivir and seltamivir, interferons, nucleosides, siRNAs, or any of the combination thereof.

The compounds, pharmaceutical compositions, and therapeutic methods of the subject invention are useful for preventing, treating, or ameliorating infections caused by influenza viruses, including but not limited to: any of the subtypes of A influenza, B influenza, or influenza C.


Influenza A virus, as used herein, encompasses any strain of influenza A virus that is capable of causing disease in an animal or human subject, or that is an interesting candidate for experimental analysis. A large number of influenza A isolates have been partially or completely sequenced, as is described in see Macken, C. L., Goodman, J., & Boykin “The value of a database in surveillance and vaccine selection.” in Options for the Control of Influenza IV, A.D.M.E. Osterhaus, N. Cox & A. W. Hampson (Eds.) Amsterdam: Elsevier Science, 2001, 103-106. This database also contains complete sequences for influenza B and C genome segments.

Influenza viruses are enveloped, negative-stranded RNA viruses of the Orthomyxoviridae family. They are classified as influenza types A, B, and C, of which influenza A is the most pathogenic and is believed to be the only type able to undergo re-assortment with animal strains. Influenza types A, B, and C can be distinguished by differences in their nucleoprotein and matrix proteins. As discussed further below, influenza A subtypes are defined by variation in their hemagglutinin (HA) and neuraminidase (NA) genes and usually distinguished by antibodies that bind to the corresponding proteins.

The influenza A viral genome consists of ten genes distributed in eight RNA segments. The genes encode 10 proteins: the envelope glycoproteins hemagglutinin (HA) and neuraminidase (NA); matrix protein (M1); nucleoprotein (NP); three polymerases (PB1, PB2, and PA) which are components of a RNA-dependent RNA transcriptase also referred to as a polymerase or polymerase complex herein; ion channel protein (M2), and non-structural proteins (NS1 and NS2). The organization of the influenza B viral genome is extremely similar to that of influenza A whereas the influenza C viral genome contains seven RNA segments and lacks the NA gene.

Influenza A virus classification is based on the hemagglutinin and neuraminidase genes. Potential hemagglutinin and neuraminidase genes can be determined using the UniProtKB/Swiss-Prot Release 57.5 (7 Jul 2009) or the UniProtKB/TrEMBL Release 40.5 (7 Jul 2009), as is known in the art. World Health Organization (WHO) nomenclature defines each virus strain by its animal host of origin (specified unless human), geographical origin, strain number, year of isolation, and antigenic description of HA and NA. For example, human epidemics have been caused by viruses with HA types H1, H2, and H3 and NA types N1 and N2. For example, the H5N1/97 “bird-fli” incident was the first documented direct transmission of an avian influenza virus to humans, causing devastating infections with severe viral pneumonia and a mortality rate of >30-50%. Other influenza A epidemics include the Asian flu pandemic in 1957 (H2N2), the Hong Kong flu pandemic in 1968 (H3N2), the re-emergence of H1N1 (Russian flu) in 1970, and the most recent swine flu H1N1 in April 2009.

In one embodiment, the subject compounds, pharmaceutical compositions, and therapeutic methods are useful for preventing, treating, or ameliorating infections caused by influenza A viruses, including but not limited to, any of the strains of H1N1, H1N2, H1N3, H1N4, H1N5, H1N6, H1N7, H1N8, H1N9, H2N1, H2N2, H2N3, H2N4, H2N5, H2N6, H2N7, H2N8, H2N9, H3N1, H3N2, H3N3, H3N4, H3N5, H3N6, H3N7, H3N8, H3N9, H4N1, H5N2, H5N3, H5N4, H5N5, H5N6, H5N7, H5N8, H5N9, H6N1, H6N2, H6N3, H6N4, H6N5, H6N6, H6N7, H6N8, H6N9, H7N1, H7N2, H7N3, H7N4, H7N5, H7N6, H7N7, H7N8, H7N9, H8N1, H8N2, H8N3, H8N4, H8N5, H8N6, H8N7, H8N8, H8N9, H9N1, H9N2, H9N3, H9N4, H9N5, H9N6, H9N7, H9N8, H9N9, H10N1, H10N2, H10N3, H10N4, H10N5, H10N6, H10N7, H10N8, H10N9, H11N1, H11N2, H11N3, H11N4, H11N5, H11N6, H11N7, H11N8, H11N9, H12N1, H12N2, H12N3, H12N4, H12N5, H12N6, H12N7, H12N8, H12N9, H13N1, H13N2, H13N3, H13N4, H13N5, H13N6, H13N7, H13N8, H13N9, H14N1, H14N2, H14N3, H14N4, H14N5, H14N6, H14N7, H14N8, H14N9, H15N1, H15N2, H15N3, H15N4, H15N5, H15N6, H15N7, H15N8, and H15N9.

In a specific embodiment, the subject compounds, pharmaceutical compositions, and therapeutic methods are useful for preventing, treating, or ameliorating infections caused by influenza A viruses, including but not limited to, any of the strains of H1N1, H9N2, H3N2, H5N1, H2N2, H7N7, and H7N1.

In another specific embodiment, the subject compounds, pharmaceutical compositions, and therapeutic methods are useful for preventing, treating, or ameliorating infections caused by influenza A viruses, including but not limited to, H1N1 (A/HK/54/98, oseltamivir-resistant strain), H1N1 (A/Victoria/0715/2000), oseltamivir-resistant strain), H9N2 (A/Quail/HK/G1/97), H3N2 (A/H3N2/1174/99), H5N1(A/H5N1/97), H1N1 (A/54/98), H9N2/G1, and H6N1 (A/Total/HE/312/97).

In another specific embodiment, the subject compounds, pharmaceutical compositions, and therapeutic methods are useful for preventing, treating, or ameliorating infections caused by viruses, including but not limited to, respiratory syncytial virus, rhinovirus, HIV virus; hepatitis
viruses including hepatitis A virus, hepatitis B virus, hepatitis C virus, hepatitis D virus, hepatitis E virus, hepatitis F virus, and hepatitis G virus; oncoviruses; human papillomavirus (HPV); human T-lymphotropic virus Type I (HTLV-1); bovine leukemia virus (BLV); Epstein-Barr virus; herpes simplex virus 1; herpes simplex virus 2; coronavirus; and poliovirus.

[0110] In one embodiment, subject compounds, pharmaceutical compositions, and therapeutic methods are useful for preventing, treating, or ameliorating infections caused by, including but not limited to, Varicella zoster (also known as chickenpox or herpes zoster), herpes simplex, cytomegalovirus and herpes simplex virus-6 (also known as AIDS-associated Kaposi sarcoma virus). In a specific embodiment, the subject invention is used to treat neutralgia or neurasthenia associated with herpes zoster reactivation, commonly known as shingles. In another specific embodiment, the subject invention is used to treat chronic fatigue syndrome caused by viral infections.

[0111] As is known in the art, genetic variation occurs by two primary mechanisms in viruses such as influenza A. Genetic drift occurs via point mutations, which often occur at antigenically significant positions due to selective pressure from host immune responses, and genetic shift, involving substitution of a whole viral genome segment of one subtype by another. In addition, many different types of animal species including humans, swine, birds, horses, aquatic mammals, and others, may become infected with viruses. Therapeutic methods useful for preventing, treating, or ameliorating infections caused by viral variants are embodiments of the subject invention.

[0112] In addition, the subject compounds, pharmaceutical compositions, and therapeutic methods are useful for preventing, treating, or ameliorating infections caused by pathogens, including but not limited to, bacteria, fungi, parasitic microorganisms, RNA, DNA, retroviral vectors, tumor/oncogenic viruses, prions, protozoan, environmental toxins, and a combination of infectious pathonogens.

[0113] In one embodiment, the subject compounds, pharmaceutical compositions, and therapeutic methods are useful for treating or ameliorating fever in a subject.

[0114] In one embodiment, the subject method for treating or ameliorating viral infection, comprises:

[0115] (i) administering, to a subject in need of such treatment, an effective amount of an isolated compound selected from the group consisting of

[0116] a) Compound A

[0117] b) Compound B

[0118] c) Compound C

[0119] d) Compound D

[0120] e) prodrugs, metabolites and/or salts of these compounds; and

[0121] f) any combination thereof;

wherein

[0122] R_{1}^{1}-R_{1}^{1} 

[0123] R_{1}^{n}-R_{1}^{n}

[0124] R_{1}^{n}-R_{1}^{n}

[0125] (ii) determining the presence and/or level of one or more viruses in the subject; wherein the determination is optionally made at multiple times to monitor the change over time; and/or optionally, and

[0126] (iii) determining the presence and/or level of one or more bio-markers in the subject; wherein the determination is optionally made at multiple times to monitor the change over time.
The presence and/or level of a type of virus or multiple types of viruses can be determined from a sample of biological fluid obtained for the purpose of evaluation of a subject of interest, such as a patient. The presence and/or level of the viruses can be measured in a biological sample such as, blood, tissue, serum, plasma, urine, saliva, and tears. In one embodiment, the sample is a blood sample. In another embodiment, the sample is a urine sample. In another embodiment, the sample is a saliva sample. In another embodiment, the sample is a bodily fluid sample.

The presence and/or level of a type of virus or multiple types of viruses can be determined in a manner as is known in the art, such as for example a titration, a viral titer, enzyme-linked immunosorbant assay (ELISA), western blot, and immunological assays.

Upon viral infection, drastic changes are rapidly induced in the host cells. Among various consequences caused by viral infections, a shared feature is the shifting of the cellular response to pathogens from apoptotic death to autophagy. The switch to the autophagy state is associated with the induction of pro-inflammatory cytokines in the virus-infected human macrophages. For instance, coronavirus and poliovirus are capable of inducing the autophagy processes in the host cells in favor of the survival and replication of viruses.

Influenza virus-infected human macrophages exhibit delayed onset of apoptosis and hyper-induction of a variety of pro-inflammatory cytokines and related molecules. For instance, the H5N1 virus is found to induce high levels of pro-inflammatory cytokines in differentiated primary human blood macrophages. This cytokine dysregulation contributes to the pathogenesis and severity of the disease. In addition, pattern recognition receptors, MAPK, also plays a significant role in the hyper-induction of TNF-α in H5N1-infected macrophages. Influenza such as H5N1 and H9N2/G1 infection also triggers the hyper-induction of IFN-β and IFN-α (including IFN-α subtypes such as IFNA1, 2 and 8) in human blood macrophages. Additionally, interferon regulatory factor 3 (IRF3), and TNF-α plays a significant role in the hyper-induction of the cytokines including IFN-β, IFN-α, and TNF-α in human macrophages infected with H5N1 viruses.

A switch of cellular response from apoptotic death to the autophagy state, associated with the induction of pro-inflammatory cytokines in the host cells, is also found in other pathogenic infections such as bacterial, fungal, and microbial infections, including infections caused by intracellular microbes such as Listeria and Mycobacteria.

The compounds of the subject application have useful immunomodulatory properties by regulating or assisting the regulation of cytokines and cytokine-related molecules during infection. Also, these cytokines and related molecules can serve as bio-markers, such as for the determination of the severity of the condition and progression of the infection, appropriateness of therapeutic methods, dosage, route of administration, and/or the need for administration of other pharmaceutical agents.

The biomarkers include, but are not limited to, TNF-α, Interleukin-1beta (IL-1β), COX-2, prostaglandin (e.g., PEG2), interferons such as interferon-beta (IFN-β), interferon-alpha (IFN-α) which includes IFN-α subtypes such as IFNA1, 2 and 8, and Interferon-gamma (IFN-γ); p38K; IRF3; the interleukin family such as Interleukin-1 (IL-1), Interleukin-2 (IL-2), Interleukin-3 (IL-3), Interleukin-4 (IL-4), Interleukin-5 (IL-5), Interleukin-6 (IL-6), Interleukin-7 (IL-7), Interleukin-8 (IL-8), Interleukin-9 (IL-9), Interleukin-10 (IL-10), Interleukin-11 (IL-11), Interleukin-12 (IL-12), Interleukin-13 (IL-13), Interleukin-14 (IL-14), Interleukin-15 (IL-15), Interleukin-16 (IL-16), interleukin-17 (IL-17), Interleukin-18 (IL-18), Interleukin-19 (IL-19), Interleukin-20 (IL-20), Interleukin-21 (IL-21), Interleukin-22 (IL-22), Interleukin-23 (IL-23), Interleukin-24 (IL-24), Interleukin-25 (IL-25), Interleukin-26 (IL-26), Interleukin-27 (IL-27), Interleukin-28 (IL-28), Interleukin-29 (IL-29), Interleukin-30 (IL-30), Interleukin-31 (IL-31), Interleukin-32 (IL-32), Interleukin-33 (IL-33), Interleukin-34 (IL-34), Interleukin-35 (IL-35); the interleukin receptor family; the macrophage inflammatory protein family such as macrophage inflammatory protein 2 (MIP-2) and macrophage inflammatory protein 1α (MIP-1α); macrophage colony-stimulating factor (M-CSF); monocyte chemotactic protein-1 (MCP-1); and immunoglobulins such as IgA, IgG, IgM, IgD, and IgE.

Immunglobulins include IgG, IgM, IgD, IgE, IgA and subtypes such as for example IgG1, IgG2, IgG3, IgG4, IgA1, and IgA2. They further include molecules in monomeric or multimeric form, whether digested from whole antibody or produced by other means.

In a further embodiment, cytokines and related molecules mediated by the compounds of the subject invention are selected from the group consisting of TNF-α, IFN-β, IFN-α including subtypes 1, 2 and 8, IFN-γ, p38K, IRF3, IL-1, IL-2, IL-3, IL-5, IL-4, IL-6, IL-8, IL-10, IL-12, IL-13, IL-14, IL-17, IL-18, IL-23, IL-24, IL-25, IL-27, IL-32, G-CSF, M-CSF, MCP-1, MIP-2, MIP-1α, IgA, IgG, IgM, IgD and IgE.

The presence and/or level of the bio-markers can be determined from a sample of biological fluid obtained from a subject of interest, such as a patient. The presence and/or level of the viruses can be measured in a sample such as, blood, tissue, serum, plasma, urine, saliva, and tears. In one embodiment, the sample is a blood sample. In another embodiment, the sample is a saliva sample. In another embodiment, the sample is a bodily fluid sample.

The compounds of the subject invention, the level of the bio-marker can be determined by quantitative immunological detection methods, such as for example enzyme-linked immunosorbant assay (ELISA), western blot, immunological assays, microarray and radioimmunoassay.

Further, a plurality of markers can be measured. In addition, analysis of a plurality of markers may be carried out separately or simultaneously. Several markers may be combined into one test for efficient processing of multiple samples from a subject.

In one embodiment, the subject invention provides a therapeutic method by administering isolated compounds. As used herein, “isolated” refers to compounds that have been removed from any environment in which they may exist in nature. For example, isolated forsythoside A would not refer to the forsythoside A compound as it exists in Pratos forsythiae. In preferred embodiments, the compounds of the subject invention are at least 75% pure, preferably at least 90% pure, more preferably are more than 95% pure, and most preferably are more than 99% pure (substantially pure).

In one specific embodiment, a therapeutic composition of the subject invention includes compounds A, B, C, and/or combination thereof, in the absence of other forsythoside compounds.
In another specific embodiment, a therapeutic composition of the subject invention includes compounds A1 (forsythoside A), B1, C1, D1, produgs, metabolites or salts thereof, and any combination thereof, in the absence of other forsythoside compounds.

The present invention also provides for a therapeutic method by administering therapeutic or pharmaceutical compositions in a form that can be combined with a pharmaceutically acceptable carrier. In this context, the compound may be, for example, isolated or substantially pure. The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the compound is administered. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum oil such as mineral oil, vegetable oil such as peanut oil, soybean oil, and sesame oil, animal oil, or oil of synthetic origin. Saline solutions and aqueous dextrose and glyceral solutions can also be employed as liquid carriers, particularly for injectable solutions.

Suitable methods of administration include, but are not limited to, oral, nasal, pulmonary administration including intravenous, subcutaneous, topical, transdermal, intradermal, transmucosal, intraperitoneal, intramuscular, intracapular, intrarbitral, intracardiac, transtracheal, subcutaneous, subcuticular, intraarticular, subcapsular, subarachnoid, intraspinal, epidural and intrathecal injection, infusion, and electroproporation, as well as co-administration as a component of any medical device or object to be inserted (temporarily or permanently) into a subject.

In one embodiment, the subject method further comprises administering to a subject a second anti-viral agent, including but not limited to, compounds such as adavante and neuraminidase inhibitors, zanamivir, oseltamivir, peramivir, and seltenamivir; interferons; nucleotides; siRNAs; or any of the combination thereof; and wherein the second anti-viral agent can be administered prior to, subsequently to, or simultaneously with the compounds or pharmaceutical compositions of the subject invention.

In a specific embodiment, the subject method further comprises administering to a subject a second anti-viral agent, selected from the group consisting of zanamivir, oseltamivir, peramivir, and seltenamivir, and any of the combination thereof; wherein the second anti-viral agent can be administered prior to, subsequently to, or simultaneously with the compounds or pharmaceutical compositions of the subject invention.

In another embodiment, the subject compounds or pharmaceutical compositions can be used in combination with at least one second anti-viral agent, optionally administered together with at least one second anti-viral agent as a vaccine; or administered prior to, simultaneously with, or subsequently to at least one second anti-viral agent.

Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, gelatin, propylene, glycol, water, ethanol and the like. The therapeutic composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions of emulsion, tablets, capsules, powders, sustained-release formulations and the like. The composition can be formulated with traditional binders and carriers such as triglycerides. Examples of suitable pharmaceutical carriers are described in "Remington's Pharmaceutical Sciences" by E. W. Martin. Such compositions contain a therapeutically effective amount of the therapeutic composition, together with a suitable amount of carrier so as to provide the form for proper administration to the patient. The formulation should suit the mode of administration.

In one embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for local injection administration to human beings. Typically, compositions for local injection administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lidocaine to ease pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water free concentrate in a hermetically sealed container such as an ampoule or sachet indicating the quantity of active agent. Where the composition is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

The therapeutic or pharmaceutical composition of the invention can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include, but are not limited to, hydrochloric, phosphoric, acetate, oxalic, tartaric acids, sodium, potassium, ammonium, calcium, ferric hydroxides, isopropyl amine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

The present invention also provides for the modification of the composition such that it is more stable once administered to a subject, i.e., once administered it has a longer time period of effectiveness as compared to the unmodified compound. Such modifications are well known to those of skill in the art, e.g., microencapsulation, etc.

The amount of the therapeutic or pharmaceutical composition of the invention which is effective in the treatment of a particular disease, condition or disorder will depend on the nature of the disease, condition or disorder and can be determined by standard clinical techniques. In general, the dosage ranges from about 0.001 mg/kg to about 2 mg/kg. In addition, in vitro assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed in the formulation will also depend on the route of administration, and the seriousness of the disease, condition or disorder, and should be decided according to the judgment of the practitioner and each patient's circumstances. Effective doses may be extrapolated from dose-response curves derived from in vitro or animal model test systems. For example, in order to obtain an effective mg/kg dose for humans based on data generated from rat studies, the effective mg/kg dosage in rats is divided by six.

The invention also provides a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients, e.g., compound, carrier suitable for administration.

The method of administration can also be practiced consistent with traditional Chinese medicine practices. The composition and dosage of the formulation that are effective in the treatment of a particular disease, condition or disorder will depend on the nature of the disease, condition or disorder by standard clinical techniques.

The traditional Chinese medicine in prescription amounts can be readily made into any form of drug, suitable for administering to humans or animals. Suitable forms include, for example, tinctures, decoctions, and dry extracts. These can be taken orally, applied through venous injection.
mucous membranes or inhalation. The active ingredient can also be formulated into capsules, powder, pellets, granules, tablets, pastille, suppositories, oral solutions, pasteurized gastroenteric suspension injections, small or large amounts of injection, frozen powder injections, pasteurized powder injections and the like. All of the above-mentioned methods are known to people skilled in the art, described in books and commonly used by practitioners of herbal medicine.

A tincture is prepared by suspending herbs in a solution of alcohol, such as, for example, wine or liquor. After a period of suspension, the liquid (the alcohol solution) may be administered for example, two or three times a day, one teaspoon each time.

A decoction is a common form of herbal preparation. It is traditionally prepared in a clay pot, but can also be prepared in glass, enamel or stainless steel containers. The formulation can be soaked for a period of time in water and then brought to a boil and simmered until the amount of water is reduced by, for example, half.

An extract is a concentrated preparation of the essential constituents of a medicinal herb. Typically, the essential constituents are extracted from the herbs by suspending the herbs in an appropriate choice of solvent, typically, water, ethanol/water mixture, methanol, butanol, isobutanol, acetone, hexane, petroleum ether or other organic solvents.

The extracting process may be further facilitated by means of maceration, percolation, repercolation, counter-current extraction, turbo-extraction, or by carbon-dioxide hypercritical (temperature/pressure) extraction. After filtration to rid of herb debris, the extracting solution may be further evaporated and thus concentrated to yield a soft extract (extractum spissum) and/or eventually a dried extract (extractum siccum), by means of spray drying, vacuum oven drying, fluid-bed drying or freeze-drying. The soft extract or dried extract may be further dissolved in a suitable liquid to a desired concentration for administering or processed into a form such as pills, capsules, injections, etc.

In one embodiment, the subject method further comprises administering to a subject, in addition to a compound of the present invention, a traditional Chinese medicinal material, including but not limited to, Fructus Forsythiae, Herba seu Flos Schizonepetae Tenuifoliae, Fructus Arctii Lappace, Semen Sojae Preparatum, Herba Lophatheri Gracillis, Radix Glycyrrhizae Uralensis, and Radix Platycodi Grandiflori.

Materials and Methods

Plant Material

The medicinal herbs, Herba menthae haplocalycis (60 g), Herba seu Flos Schizonepetae Tenuifoliae (40 g), Fructus Forsythiae suspensae (100 g), Fructus Arctii Lappace (60 g), Semen Sojae Preparatum (50 g), Herba Lophatheri Gracillis (40 g), Radix Glycyrrhizae Uralensis (50 g), Flos Lonicerae Japonicae (100 g) and Radix Platycodi Grandiflori (60 g) as well as herbal extracts are provided by PuraPharm International (H.K.) Ltd.

Fructus Forsythiae obtained from PuraPharm International (H.K.) Ltd. is dried and ground into powders. Methods for extracting bioactive compounds are shown in FIG. 1 and illustrated as follows.

Briefly, Fructus Forsythiae (100 g) is dried, ground into powders, and then extracted twice for bioactive compounds. During each extraction, powders are suspended in 10× milli-Q water under reflux for 2 hours. The supernatant from each extraction is collected, combined and evaporated to dryness under vacuum to yield 22.23 g light yellowish powders. The powders are re-dissolved in MeOH and fractionated.

The resulting MeOH extract is purified by reversed-phase high-performance liquid chromatography (HPLC) (Lichrospher 100 RP C18 EC 5 μm, 250×4.6 mm ID) using a gradient elution from 10% acetonitrile (CH3CN) to 90% CH3CN at a flow rate of 1 mL/min.

Peak detection is achieved using an Agilent 1200 series of fast scanning Photo-diode Array detector set at 210, 254 and 280 nm. Eluting peaks are scanned between 200 nm and 300 nm with 1 nm intervals to determine absorbance maxima and minima. A total of 13 fractions are obtained.

Each fraction is examined for its biological activities. Fraction 4 shows COX-2 inhibitory effects, and fraction 5 exhibits significant anti-viral effects; thus, they are subject to further purification. By repeating the purification process using HPLC, a pure compound (Compound A1) (6.1 mg) with anti-viral effects and a pure compound (Compound D1) (2.8 mg) with COX-2 inhibitory effects are obtained.

Elucidation of the Molecular Structure

The structure of the pure compounds is elucidated through H and 13C NMR spectroscopy as well as through TOF-ESI-MS and FT-MS spectrometry. The NMR spectra are recorded on a Bruker 500 MHz DRX NMR spectrometer, operating at 500 MHz for 1H and at 125 MHz for 13C NMR, using methanol-d4 with TMS as the solvent. The accurate mass determination for Compound A1 is recorded on a microTOF II ESI-TOF mass spectrometer (Bruker Daltonics), and the sample is dissolved in MeOH. The accurate mass determination for Compound D1 is recorded on a 5975C ESI-MS (Agilent Technologies).

Preparation of Yiqingaoxsan (YQS) Extract

The YQS extract is prepared according to the Chinese Pharmacopeia (2005). Briefly, 3 g of Herba menthae haplocalycis and 2 g of Herba seu Flos Schizonepetae Tenuifoliae are extracted twice with 20-fold milli-Q water under reflux for 1 hour. The essential oil and the water extract are collected. The residue is combined with other five herbs including Fructus Forsythiae suspensae (5 g), Fructus Arctii Lappace (3 g), Semen Sojae Preparatum (2.5 g), Herba Lophatheri Gracillis (2 g), and Radix Glycyrrhizae Uralensis (2.5 g) and then extracted twice with 10-fold of milli-Q water under reflux for 2 hours. The supernatant is collected and combined with the previous water extract. The resulting extract is then lyophilized under reduced pressure. The dried paste is combined with Flos Lonicerae Japonicae (5 g) and Radix Platycodi Grandiflori (3 g) and then extracted three times with 10-fold of MeOH. Around 5 g of MeOH extract is obtained.

Fractionation of YQS Extract for Anti-Viral Bioassay

The MeOH extract of YQS is fractionated by reversed-phase HPLC (Lichrospher 100 RP C18 EC 5 μm, 250×4.6 mm ID), using a gradient elution from 10% acetonitrile (CH3CN) to 90% CH3CN at a flow rate of 1 mL/min.
0169] Peak detection is achieved using an Agilent 1200 series of fast scanning Photo-diode Array detector set at 210, 254 and 280 nm. A total of 5 fractions are obtained.

Fractionation of YQS Extract for IL-1β Bioassay

0170] The YQS extract is fractionated by reversed-phase HPLC (Lichrospher 100 RP C18 EC 5 μm, 250x4.6 mm ID) using a gradient elution from 10% acetonitrile (CH₃CN) to 90% CH₃CN at a flow rate of 1 ml/min.

0171] Peak detection is achieved using an Agilent 1200 series of fast scanning Photo-diode Array detector set at 210, 254 and 280 nm. A total of 13 fractions are obtained.

Preparation of Human Influenza Viruses

0172] Human influenza H1N1 virus (A/HK/54/98 (oseltamivir-sensitive strain) and A/Victoria/0715/2000/07 (oseltamivir-resistant strain)), H9N2 (A/Quail/HK/G1/97), and H3N2 (A/H3N2/1174/99) are prepared as described in Lee et al. 2005 and Mok et al. 2007 (14, 15), which are incorporated by reference in their entirety.

0173] To briefly illustrate, first, the viruses are isolated from human beings. The isolated human influenza H1N1 virus (A/HK/54/98 (oseltamivir-sensitive strain) and A/Victoria/0715/2000/07 (oseltamivir-resistant strain)), H9N2 (A/Quail/HK/G1/97), and H3N2 (A/H3N2/1174/99) are cloned by limiting dilution. Seed virus stocks are prepared in MDCK (Madin-Darby canine kidney) cells.

Viral Infection

0174] MDCK cells and macrophages are infected with the indicated viruses at a multiplicity of infection (m.o.i.) of 0.2 or 2 for 30 min at 37°C. The supernatant containing the virus inoculum is then removed, and the cells are incubated in MEM or macrophage serum-free medium (Invitrogen).

Determination of Tissue Culture Infective Dose (TCID₅₀)

0175] Prior to TCID₅₀ assays, MDCK cells are seeded at 2x10⁶ cells per well on the 96-well plates. Culture supernatants are harvested from virus-infected cells at 48 hour post-infection. Serial 2-fold dilutions of the supernatant samples are prepared, and the diluted samples are incubated with MDCK cells for 1 hour for viral adsorption.

0176] The virus inoculum is then removed. Cells are washed once and replenished with MEM supplemented with 1 μg/ml TPCK-treated trypsin. After four days of incubation, the cells are fixed with 10% formaldehyde for 30 min and stained with 0.5% crystal violet to determine viral-induced cytopathic effects. TCID₅₀ titers are calculated using the Reed-Muench formula.

Immunocytochemical Staining

0177] At the indicated time points after infections, the cells are fixed and viral nucleoprotein and M1 protein are stained with the Influenza Detection Kit (Oxoid) according to the manufacturer’s protocol.

Protein Extraction and Western Blot Analysis

0178] Whole-cell lysates are prepared with total lysis buffer (50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 50 mM NaF, 10 mM glycerophosphate, 0.1 mM EDTA, 10% glycerol, 1% Triton X-100) containing a protease inhibitor cocktail. For Western blot analysis, 10 μg protein is heat denatured in a sample buffer (125 mM Tris, pH 6.8, 4% sodium dodecyl sulfate, 20% glycerol, 5% beta-mercaptoethanol, 0.01% bromophenol blue), separated by 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis, and transferred onto a nitrocellulose membrane for assaying protein levels with ECL™ Western Blotting Detection Reagents (GE Healthcare) solution.

Transmission Electron Microscopy

0179] At 18 hour post-infection, cells are fixed with 2.5% glutaraldehyde in the cacodylate buffer (0.1 M sodium cacodylate-HCl buffer, pH 7.4) for 15 min on ice. Cells are then scraped from the culture dish, and the pellet is harvested by centrifugation at 5000 rpm for 3 min. Cells are then resuspended in cacodylate buffer with 0.1 M sucrose. After washings with cacodylate buffer, cells are fixed in 1% osmium tetroxide in cacodylate buffer for 30 min at room temperature. Cells are washed three times and then dehydrated with graduated ethanol. After two washings with propylene oxide, cell pellets are embedded in epoxy resin. Ultrathin sections with a thickness of 100 nm are cut. Ultrastructural features of cells are examined under a transmission electron microscope (Philips EM208S).

Quantitative Reverse Transcription-PCR Analyses

0180] Total RNA are extracted with TRIzol reagent (Invitrogen) according to the manufacturer’s instructions. The cDNA is synthesized from total RNA with oligo(dT) primers and Superscript II reverse transcriptase (Invitrogen). The levels of mRNA encoding COX-2 or IL-1β are assayed with TaqMan gene expression assays.

Prostaglandin E₂ Assay

0181] At 48 hours post-infection, cell culture supernatants are harvested and prostaglandin E₂ (PGE₂) levels in the supernatant are measured by Prostaglandin E₂ ELA Kit (Cayman) according to manufacturer’s instructions.

EXAMPLES

0182] Following are examples that illustrate procedures for practicing the invention. These examples should not be construed as limiting.

Example 1

Extraction and Identification of Forsythoside A and Jacamine

0183] Light yellowish powders are obtained by repeated purification of the MoOH extracts prepared from Fructus Forsythiae using reversed-phase HPLC. The detailed procedures are summarized in FIG. 1.

0184] With bioactivity guided purification using sequential HPLC, an antiviral molecule designated as Compound A is eluted at approximately 6.8 min as a single compound (>95% purity) with UV absorbance maximized at 220, 290 and 330 (FIG. 2). The ¹³C NMR spectra of Compound A, as shown in FIG. 4, display signals at δ131.5 (C-1), 116.5 (C-2), 146.2 (C-3), 144.8 (C-4), 117.2 (C-5), 121.4 (C-6), 72.4 (C-7), 36.8 (C-8), 104.6 (C-9), 75.3 (C-10), 76.0 (C-11), 72.5 (C-12), 74.9 (C-13), 67.8 (C-14), 102.4 (C-15), 72.1 (C-16), 72.4 (C-17), 74.1 (C-18), 70.0 (C-19), 18.1 (C-20), 127.8
Inhibitory Effects of Forsythoside A on Viruses

To determine the inhibitory effects of forsythoside A (Compound A1) on viruses, MDCK cells are infected with human influenza viruses, and a viral titer (TCID₅₀) bioassay is performed according to the procedures illustrated as follows.

Briefly, MDCK cells are seeded at 1x10⁶ cells/well on a 24-well plate, and infected with human influenza viruses including H1N1 (A/HK/54/98), an oseltamivir-sensitive strain; H1N1 (A/Victoria/07159200/07) (H1N1-R), an oseltamivir-resistant strain; H9N2 (A/Quail/HK/G1/97); and H3N2 (A/H3N2/1174/99) at a multiplicity of infection (m.o.i.) of 2 for 30 minutes, respectively. After that, cells are washed with PBS once to remove non-absorbed viruses, and treated with forsythoside A (Compound A1) at a concentration of 100 µg/ml supplemented with minimum essential medium (MEM).

After incubation for 48 hours with forsythoside A (Compound A1) or DMSO, the cell culture supernatant is collected and subject to a viral titer (TCID₅₀) assay for measuring the inhibitory effects of forsythoside A (Compound A1) on influenza virus replication. Briefly, a two-fold serially diluted supernatant is added to MDCK cells. Cells are incubated for one hour at 37°C with 5% CO₂ for viral adsorption. The cells are then washed with MEM or phosphate-buffered saline to remove non-absorbed virus inoculum, and replenished with fresh MEM supplemented with 1 µg/ml N-tosyl-L-phenylalanyl chloromethyl ketone (TPCK)-treated trypsin. Cells are further incubated at 37°C with 5% CO₂ for four days, then fixed with 10% formaldehyde for 30 minutes and stained with 0.5% crystal violet for examining the virus-induced cytopathic effects. TCID₅₀ titers are calculated using the Reed-Muench formula as described in Methods and Techniques in Virology, 1993,137, which is hereby incorporated by reference in its entirety.

The results, as shown in Fig. 7, reveal that viral replication in the MDCK cells is suppressed by forsythoside A (Compound A1) at 100 µg/ml. Specifically, the replication of influenza viruses H1N1 (A/HK/54/98), H1N1 (A/Victoria/07159200/07), H9N2 (A/Quail/HK/G1/97) and H3N2 (A/H3N2/1174/99) is suppressed by forsythoside A (Compound A1) for about 15-, 3-, 24- and 12-fold, respectively when compared with DMSO-treated cells (Fig. 7A-D, Representative results from three experiments). After infecting the cells with H1N1 (A/HK/54/98), Tamiflu (100 µM) was also added to the cells as control (Fig. 7E). The virus was suppressed by more than 30-fold when compared with the diluent (PBS)-treated cells (FIGS. 7E).

Inhibitory Effects of Forsythoside A on Influenza Viruses in Primary Human Blood Macrophages

Primary human blood macrophages are infected with human influenza viruses including H1N1 (A/HK/54/98); H9N2 (A/Quail/HK/G1/97); and H3N2 (A/H3N2/1174/99) at a m.o.i. of 2. Cells are then incubated with forsythoside A (Compound A1, 100 µg/ml) or DMSO for 48 hours. After that, cell culture supernatants are collected and viral titers (TCID₅₀) are measured by titration in MDCK cells. (Representative results from three experiments.) The results,
as shown in FIG. 8, demonstrate that forsythoside A (Compound A1) possesses anti-influenza virus effects in primary human blood macrophages.

Example 5

Inhibitory Effects of Forsythoside A on Influenza Virus Replication Shown by Immunocytochemistry and Western Blot Analysis

[0195] This Example demonstrates that forsythoside A suppresses influenza virus replication. MDCK cells are infected with human influenza virus H9N2/G1 (A/Quail/11K/G1/97) at an m.o.i. of 0.2. Cells are then incubated with forsythoside A (Compound A1) at 100 μg/ml or DMSO for 6 or 10 hours. After that, cells are fixed and influenza nucleoproteins and M1 proteins are stained with the Influenza Detection Kit (Oxoid). The numbers of cells expressing viral proteins are counted and percentages of cells expressing viral proteins are calculated.

[0196] As shown in FIG. 9 (A & B), at 6 hour post-infection (h.p.i.), no significant difference is found when comparing the percentages of cells expressing viral proteins upon DMSO or forsythoside A (Compound A1) treatments. However, from 6 h.p.i. to 10 h.p.i., the number of cells expressing viral proteins increases by about 100% when treated with DMSO; while the percentage only increases by about 15% when treated with forsythoside A (Compound A1). This indicates that forsythoside A (Compound A1) decreases the number of viral-infected cells.

[0197] MDCK cells are infected with human influenza virus H9N2/G1 (A/Quail/11K/G1/97) at an m.o.i. of 2 or mock infected. After that, cells are then incubated with forsythoside A (Compound A1) (100 μg/ml) or DMSO. Proteins are harvested at 2, 4, 9 or 24 h.p.i.

[0198] Expressions of the influenza M1 protein are analyzed by Western blot analysis. Actin is used as a loading control. Expression level of viral M1 proteins is analyzed and band intensities are measured by Quantity One software (Bio-Rad). The relative intensities are determined by normalizing the M1 intensities to that of actin.

[0199] As shown in FIG. 9C, the M1 protein expression level is lower in the cells treated with forsythoside A (Compound A1), when compared to those treated with DMSO. At 2 h.p.i., the relative intensity of M1 in DMSO-treated cells is higher than that of forsythoside A-treated cells by more than four fold. Furthermore, over the period of 2 to 24 h.p.i., the relative intensity increases from 0.13 to 1.7 in DMSO-treated cells. On the contrary, in the cells with forsythoside A (Compound A1) treatment, the relative intensity only reaches 0.39 at 24 h.p.i. These results further show treatment with forsythoside A decreases M1 protein expression and reduces viral loads.

Example 6

Inhibitory Effects of Forsythoside A on Influenza Virus Replication Shown by Electronic Microscopy

[0200] This Example demonstrates that forsythoside A suppresses influenza virus replication. MDCK cells are infected with human influenza virus H9N2/G1 (A/Quail/HK/11K/G1/97) at an m.o.i. of 2. Cells are then incubated with forsythoside A (Compound A1) (100 μg/ml) (C-D) or DMSO (A-B) for 18 hours. Cells are then fixed and ultrastructures are examined by transmission electron microscopy.

[0201] As shown in FIG. 10, in DMSO-treated cells (A-D), most virions have been completely separated from the cells. In contrast, in forsythoside A (Compound A1)-treated cells, a portion of the virions is still attached to the cells (C-D).

[0202] The infected cells are further observed under transmission electron microscopy with higher magnifications. A very small portion of virions, attached by thin neck, are still associated with cells incubated with DMSO (B, arrow heads). In contrast, in forsythoside A-treated cells, the budding virions remain connected to the cell by thick stalks (D, arrows). This indicates a slowed or abnormal budding process due to forsythoside A (Compound A1) treatment.

Example 7

Inhibitory Effects of Jacaralone on Cyclooxygenase 2 (COX-2) and Influenza Virus Infection

[0203] Primary human blood macrophages are infected with H9N2/G1 (A/Quail/HK/G1/97) at an m.o.i. of 2. Cells are then incubated with jacobarapon (Compound D1) or DMSO for 3 hours. After that, total RNA are harvested and COX-2 mRNA levels are examined by TaqMan Gene Expression assays.

[0204] As shown in FIG. 11A, jacobarapon (Compound D1) suppresses COX-2 mRNA levels upon H9N2/G1 infection and the suppression occurs in a dose-dependent manner. Jacobarapon suppresses COX-2 mRNA level by about 20% at 10 μg/ml and by 40% at 20 μg/ml.

[0205] After incubating the cells with jacobarapon (Compound D1) for 48 hours, prostaglandin E2 (PGE2) levels in the cell culture supernatant are measured by the Prostaglandin E2 EIA Kit (Cayman). FIG. 11B shows that treating non-infected macrophages with jacobarapon (Compound D1) alone does not cause any changes in PGE2 levels, when compared to the mock-treated cells. Infection with H9N2/G1 significantly induces PGE2 levels. Consistent with the RNA results, jacobarapon (Compound D1) suppresses PGE2 levels in a dose-dependent manner.

Example 8

Inhibitory Effects of YQS Formula on Influenza Virus Replication

[0206] MDCK cells are infected with H1N1 (A/HK/54/98) or H9N2/G1 (A/Quail/HK/G1/97) at an m.o.i. of 2. Cells are then incubated with the Yin Qiao San (YQS) (100 μg/ml) or DMSO for 48 hours. After that, cell culture supernatants are collected and viral titers (TCID₅₀) are measured by titration in MDCK cells.

[0207] FIG. 13A shows that YQS inhibits H1N1 and H9N2/G1 virus replication. In addition to YQS, cells are incubated with YQS fractions (F1-F5) (100 μg/ml) (as shown in FIG. 12) or DMSO after being infected with H9N2G1 (A/Quail/HK/G1/97).

[0208] After 48-hour incubation, cell culture supernatants are collected and viral titers (TCID₅₀) are measured by titration in MDCK cells. As shown in FIG. 13C, fractions F2-F5 show viral inhibitory effects. Among F2-F5, fractions F2 and F3 have the strongest anti-viral effects.
Example 9

Induction of Interleukin-1Beta by YQS Formula

[0209] Primary human blood macrophages are infected with H1N1 (A/HK/498/2009-A) at m.o.i. of 2 or mock infected. H1N1-infected cells are then incubated with YQS (100 μg/ml) or DMSO for 3 hours. After that, total RNA are harvested and interleukin(IL)-1β mRNA levels are examined by TaqMan Gene Expression assays. FIG. 15A shows that YQS enhances IL-1β mRNA production induced by H1N1.

[0210] Primary human blood macrophages cells are incubated with YQS fractions [S1-S-13, 100 μg/ml] (as shown in FIG. 14) or DMSO after being infected with H1N1. FIG. 15B shows that H1N1 infection increases IL-1β mRNA level by about 3 folds (DMSO), when compared with the mock-infected cells. Treatment with YQS fraction S11 increases IL-1β level by about 20 fold, when compared with the mock-infected cells.

[0211] All references, including publications, patent applications and patents, cited herein are hereby incorporated by reference to the same extent as if each reference was individually and specifically indicated to be incorporated by reference and was set forth in its entirety herein.

[0212] The terms “a” and “an” and “the” and similar references as used in the context of describing the invention are to be construed to cover both the singular and the plural, unless otherwise indicated herein or clearly contradicted by context.

[0213] Recitation of ranges of values herein are merely intended to serve as a shorthand method of referring individually to each separate value falling within the range, unless otherwise indicated herein, and each separate value is incorporated into the specification as if it were individually recited herein. Unless otherwise stated, all exact values provided herein are representative of corresponding approximate values (e.g., all exact exemplary values provided with respect to a particular factor or measurement can be considered to also provide a corresponding approximate measurement, modified by “about,” where appropriate).

[0214] The use of any and all examples, or exemplary language (e.g., “such as”) provided herein, is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention unless otherwise indicated. No language in the specification should be construed as indicating any element is essential to the practice of the invention unless as much is explicitly stated.

[0215] The description herein of any aspect or embodiment of the invention using terms such as “comprising,” “having,” “including” or “containing” with reference to an element or elements is intended to provide support for a similar aspect or embodiment of the invention that “consists of,” “consists essentially of,” or “substantially comprises” that particular element or elements, unless otherwise stated or clearly contradicted by context (e.g., a composition described herein as comprising a particular element should be understood as also describing a composition consisting of that element, unless otherwise stated or clearly contradicted by context).

[0216] It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application.

REFERENCES


We claim:

1. A method for preventing and/or treating viral infection in a subject, wherein said method comprises administering, to a subject in need of such prevention and/or treatment, an effective amount of an isolated compound selected from the group consisting of:

   a) Compound A

   b) Compound B

   c) Compound C

   d) Compound D

   c) a prodrug or metabolite of any of a)-d);

   f) a salt of any of a)-e);

   g) any combination thereof; and

   wherein

   \( R_{1}, R_{2}, R_{3}, R_{4}, R_{5}, R_{6}, \) and \( R_{7}-R_{13} \) are, independently, —H, acyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkyl, alkoxy, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide,

   \( R_{1}^{"}, R_{2}^{"}, \) and \( R_{3}^{"}, R_{4}^{"}, R_{5}^{"}, R_{6}^{"}, R_{7}^{"}, R_{8}^{"}, R_{9}^{"}, R_{10}^{"}, R_{11}^{"}, R_{12}^{"}, R_{13}^{"} \) are, independently, —H, acyl, haloalkyl, alkylamino, alkyl, hydroxylalkyl, carboxyl, carboxalkoxy, or carboxamide.

2. The method, according to claim 1, wherein the subject is a human.

3. The method, according to claim 1, wherein the compound is selected from the group consisting of:

   a) forsythiside A (Compound A1)

   b) Compound B1
c) Compound C1

\[
\text{HO-} \quad \text{OH} \\
\text{HO-} \quad \text{OH} \\
\text{HO-} \quad \text{OH} \\
\text{HO-} \quad \text{OH} \\
\text{HO-} \quad \text{OH}
\]

\[
\text{OH} \\
\text{OH} \\
\text{OH} \\
\text{OH}
\]

4. The method, according to claim 1, wherein the compound is foscarnet A or acyclovir.
5. The method, according to claim 1, used to prevent and/or treat an infection caused by influenza virus A or B.
6. The method, according to claim 5, used to prevent and/or treat an infection caused by a virus selected from the group consisting of H1N1, H9N2, H3N2, H5N1, H2N2, H7N7, and H7N1.
7. The method, according to claim 6, used to prevent and/or treat an infection caused by a virus selected from the group consisting of H1N1 (A/HK/54/98, oseltamivir-sensitive strain), H1N1 (A/Victoria/07/159200/07, oseltamivir-resistant strain), H3N2 (A/HK/105/97), H3N2 (A/H3N2/1174/99), H5N1 (A/H5N1/97), H1N1 (A/54/98), H9N2/G1, and H6N1 (A/Teil/HK/W512/97).

8. The method, according to claim 1, wherein said method further comprises administering to the subject a second antiviral agent selected from the group consisting of zanamivir, oseltamivir, peramivir, seltamivir, and any combination thereof.
9. The method, according to claim 1, used to enhance IL-1β level.
10. The method, according to claim 1, used to reduce COX-2 level.
11. The method, according to claim 1, used to reduce PGE2 level.
d) Compound D

e) a prodrug or metabolite of any of a)-d); 
f) a salt of any of a)-e); 
g) any combination thereof; and

wherein

R'1-R'2, R'3-R'4, and R''1-R''2, are, independently, —H, acyl, haloalkyl, amino, alkylamino, hydroxyl, alkyl, alkoxy, hydroxyalkyl, carboxyl, carboxalkoxy, or carboxamide,

R'2-R'4 are, independently, —H, acyl, haloalkyl, alky- lamino, alkyl, hydroxyalkyl, carboxyl, carboxalkoxy, or carboxamide, and

R'1-R''2 are, independently, —H, alkyl, halo, haloalkyl, amino, alkylamino, hydroxyl, alkoxy, hydroxyalkyl, carboxyl, carboxalkoxy, or carboxamide.

16. The composition, according to claim 15, wherein the compound is selected from the group consisting of:

a) forsythoside A (Compound A1)

b) Compound B1

c) Compound C1

d) Compound D1

e) a prodrug or metabolite of any of a)-d); 
f) a salt of any of a)-e); and 
g) any combination or salt thereof.

17. The composition, according to claim 15, wherein the compound is forsythoside A or jacaranone.

18. The composition, according to claim 15, wherein the composition is formulated for oral administration.

19. The composition, according to claim 15, which does not contain any other forsythoside compound.

20. A method for treating viral infection in a subject, comprising administering, to a subject in need of such treatment, a Yin Qiao San Composition, wherein the Yin Qiao San Composition consists of Herba men rhiae haploaclycis, Herba seu Flos Schizonepetae Temulifoliae, Fructus forsythiae suspensa, Fructus Arctii Lappae, Semen Sojae Preparatum, Herba Lophatheri Gracilis, Radix Glycyrrhizae Uralensis, Flos Lonicerae Japonicae, and Radix Platycodi Grandiflori, wherein the method enhances IL-1β level in a subject who has viral infection.