Title: Diffusion Tensor Imaging of Somatosensory Tract in Cervical Spondylotic Myelopathy and Its Link with Electrophysiological Evaluation

Running Title: DTI in CSM

Chun-Yi Wen #, Jiao-Long Cui #, Kin-Cheung Mak, Keith Dip-Kei. Luk, and Yong Hu *
Department of Orthopaedics and Traumatology, Li Ka Shing Faculty of Medicine, The University of Hong Kong

“#” Equally contribution to this manuscript

“*” Corresponding Author

Dr Yong Hu
Dept. of Orthopaedics and Traumatology,
The University of Hong Kong
Address: 12 Sandy Bay Road, Pokfulam, Hong Kong

Email address: yhud@hku.hk
Tel: (852) 29740359; Fax: (852) 29740335

Acknowledgements

The General Research Fund of the University Grant Council of Hong Kong (771608M/774211M) provided financial support to this study. The authors also would like to thank Prof EX Wu, Drs. Henry Mak, Queenie Chan and Mr TH Li for their assistance during DTI protocol and scanning technical development. The authors thank American Journal Experts (AJE) for language editing.
Abstract

Background and Context: Abnormal somatosensory evoked potential (SEP), i.e., prolonged latency, has been associated with poor surgical prognosis of cervical spondylotic myelopathy (CSM).

Purpose: To further characterize the extent of microstructural damage to the somatosensory tract in CSM patients using diffusion tensor imaging (DTI).

Study Design/Setting: Retrospective study.

Patient Sample: A total of 40 volunteers (25 healthy subjects and 15 CSM patients).

Outcome Measures: Clinical, electrophysiological and radiological evaluations were performed using the Japanese Orthopaedic Association (JOA) scoring system, SEP and cord compression ratio in anatomic MR images respectively. Axial diffusion MR images were taken using a pulsed gradient, spin-echo-echo-planar imaging (SE-EPI) sequence with a 3T MR system. The diffusion indices in different regions of the spinal cord were measured.

Methods: Comparison of diffusion indices among healthy and myelopathic spinal cord with intact and impaired SEP responses were performed using one-way ANOVA.

Results: In healthy subjects, FA values were higher in the dorsal (0.73±0.11) and lateral columns (0.72±0.13) than in the ventral column of WM (0.58±0.10), e.g., at C4/5 (p<0.05). FA was dramatically dropped in the dorsal (0.54±0.16) and lateral columns (0.51±0.13) with little changes in the ventral column (0.48±0.15) at the compressive lesions in CSM patients. There were no significant differences in the JOA scores or cord compression ratios between CSM patients with or without abnormal SEP. However, patients with abnormal SEP showed a FA decrease in the dorsal column cephalic to the lesion (0.56±0.06), i.e., at C1/2, compared with healthy subjects (0.66±0.02), but the same decrease was not observed for those without a SEP abnormality (0.67±0.02).

Conclusion: Spinal tracts were not uniformly affected in the myelopathic cervical cord. Changes in diffusion indices could delineate focal or extensive myelopathic lesions in CSM,
which could account for abnormal SEP. DTI analysis of spinal tracts might provide additional information not available from conventional diagnostic tools for prognosis of CSM.

**Key words:** Cervical Spondylotic Myelopathy, Diffusion Tensor Imaging, Spinal Cord, Fractional Anisotropy, Microstructure
DTI in CSM

1 Introduction

Cervical spondylotic myelopathy (CSM) is the most common type of spinal cord dysfunction in patients older than 55 years [1-3]. The severity of somatosensory dysfunction, i.e. the prolonged latency of somatosensory evoked potential (SEP), has been identified as an indicator for the poor prognosis in CSM patients after surgical management. However, the information regarding the extent of somatosensory tract damage in CSM patients remains to be explored.

The emerging diffusion tensor imaging (DTI) technique provides in vivo detection of the microstructure of the spinal cord parenchyma [5]. Fractional anisotropy (FA) and diffusivities, e.g., mean diffusivity (MD), axial and radial diffusivities (AD and RD), were derived from the diffusion tensor matrix, which are commonly used in DTI analysis to describe the voxels’ diffusion properties [6]. Above diffusion indices are attributed to the densely packed axonal membranes in the spinal cord, and they may reflect microarchitectural changes associated with the demyelination process and axon damage in neurological injury and disease [7,8]. Feasibility of DTI has been used for CSM patients in previous studies [9-17]. However, little is known about the specific spinal tract damages in CSM due to the poor quality of diffusion MR images in previous studies under relatively lower magnetic field strengths, i.e., 0.2 and 1.5 Tesla, or sagittal slicing of the cervical spinal cord [9-17]. Several approaches were employed to improve the quality of diffusion MR images, including use of a 3.0-Tesla MRI scanner, optimizing the axial slice thickness to achieve a good signal/noise ratio (SNR) and reducing motion artifacts through cardiac/respiratory gating [18,19]. The improved image quality, with a clear separation of gray and white matter structures, makes it possible to analyze the microarchitecture of the spinal tracts anatomically.

This study aimed to (1) characterize the diffusion properties of the ventral, lateral and dorsal columns in the healthy and myelopathic cervical cord using diffusion MR images and (2) correlate SEP status, i.e., normal or prolonged latency, with DTI findings in CSM patients.
Materials and Methods

Subjects

The institutional review board of research ethics approved all experimental procedures in this study. A total of forty volunteers were recruited with informed consent (25 healthy subjects at the age of 52±7 years old and 15 CSM patients at the age of 60±9 years old). All volunteers were screened to confirm their eligibility before the study. The inclusion criteria for healthy subjects were with intact sensory and motor function and a negative Hoffman’s sign under physical examination. Those having any neurological signs or symptoms or any past history of neurological injury, disease or surgeries were excluded. Experienced spinal surgeons made clinical diagnoses based on the insidious and chronic course, neurological deficit and radiological findings of degenerative intervertebral discs and spondylosis. The CSM patients’ neurological deficits were evaluated via physical examination and the modified Japanese Orthopaedic Association (mJOA) score, with the highest score being 17 [20,21].

Electrophysiological assessments

The functional integrity of the spinal cord was evaluated using somatosensory evoked potential (SEP) [4]. In brief, stimulation was applied to the median nerve on the wrists, while SEP signals were recorded from the C3 in response to right limb stimulation and from the C4 in response to left limb stimulation, with the reference electrode at Fz according to the international 10–20 system [6]. The data were inspected for the presence of the main peaks N19/P22 by an experienced electrophysiologist. The latency and amplitude of SEP signals from CSM patients were compared with previously published healthy criteria (latency: 18.40±0.71 ms; amplitude: 1.23±0.50 µV) [4]. The impaired SEP in CSM patients were defined as delayed N19 latency (exceeding 2.5 SD), regardless of the peak-to-peak amplitude (< 0.5 µV), or waveform disappearance [4].

MRI Scanning
All images were taken via a 3.0-Tesla MRI scanner (Philips Achieva). During the acquisition process, the subject was placed in a supine position with the sense neuro-vascular (SNV) head and neck coil enclosing the cervical region and instructed not to swallow to minimize motion artifacts. The subject was then scanned to produce anatomical T1-weighted (T1W) images, T2-weighted (T2W) images and diffusion tensor images (DTI).

Sagittal and axial T1W and T2W images were acquired for each subject. A fast spin echo (FSE) sequence was employed. A total of 11 sagittal images covering the whole cervical spinal cord were acquired. Cardiac vectorcardiogram (VCG) triggering was applied to minimize the pulsation artifact from CSF. A total of 12 transverse images covering the cervical spinal cord from C1 to C7, each of which was placed at the center of either a vertebrae or intervertebral disc, were acquired. Diffusion MRI images were acquired using the pulsed sequence of single-shot spin-echo echo-planar imaging (SE-EPI). Diffusion encoding was in 15 non-collinear and non-coplanar diffusion directions with a b-value = 600 s/mm². The image slice planning was the same as that for the anatomical axial T1W and T2W images, with 12 slices covering the cervical spinal cord from C1 to C7. The duration of diffusion tensor imaging (DTI) averaged 24 minutes per subject with an average heart rate of 60 beats per minute. Spatial saturation with Spectral Presaturation with Inversion Recovery (SPIR) was applied to suppress the fold-over effect.

Image analysis

The morphometry of the spinal cord were analyzed using the previously reported methods [22], including measurement of cervical cord compression using the anterior-posterior diameter/transverse diameter ratio in axial T2W images. Intramedullary signal changes were recorded based on both T2W and T1W images.

Diffusion measurement was performed using DTIStudio software (Version 2.4.01 2003, Johns Hopkins Medical Institute, Johns Hopkins University). Image volume realignment and 3D rigid body registration with different diffusion gradients were conducted using the
Automated Image Registration (AIR) program (Laboratory of Neuroimaging, UCLA) to reduce the effect of motion artifacts. The realigned and co-registered diffusion-weighted data were double checked for image quality and then used to estimate diffusion tensors, including three eigenvalues ($\lambda_1$, $\lambda_2$ and $\lambda_3$) and the corresponding eigenvectors. Maps of the fractional anisotropy (FA) and axial and radial diffusivity (AD and RD) were derived from the diffusion matrix accordingly.

The regions of interest (ROIs) were defined in different areas of the cervical spinal cord: the ventral, lateral and dorsal columns of white matter (WM) (Figure 1) [23]. The diffusion indices were calculated by averaging all selected voxels in the ROIs using ImageJ (National Institute of Health, USA).

Statistical analysis

The FA, AD and RD values in different regions of the spinal cord were calculated at each vertebrae and disc level along the whole cervical spine (Figure 2). The degenerated disc level(s) and adjacent vertebrae level(s) were defined as the spondylotic myelopathic lesion segment for statistical analysis. Comparisons among healthy and myelopathic spinal cord with intact and impaired SEP responses were performed using one-way ANOVA. The level of significance was set at $p<0.05$. All data analyses were performed using SPSS 15.0 analysis software (SPSS Inc., Chicago, IL, USA).
Results

Clinical, radiological and electrophysiological data

A total of fifteen CSM patients presented with severe neurological deficits as indicated by their significant mJOA scores (CSM: 9.8±1.0, full score 17) and the compression of the cervical cord (0.35±0.07) compared to healthy subjects (0.52±0.05, p<0.001) (Table 1, Figure 2).

Among fifteen CSM patients, five of them presented prolonged latency in SEP (latency: 21.90±1.22 ms; amplitude: 0.87±0.42 µV) and were classified as the CSM_lat+ group. The remaining CSM patients, who presented normal SEP or only decreased amplitude (latency: 17.81±1.06 ms; amplitude: 1.14±0.64 µV), were classified as the CSM_lat- group. There were no significant differences in the age of patients (CSM_lat+: 62±8 years, CSM_lat-: 59±10 years), duration of disease (CSM_lat+: 6.1±8 years, CSM_lat-: 6.1±2.3 years) or mJOA score (CSM_lat+: 10.0±0.9, CSM_lat-: 9.4±1.1) between these two groups. Intramedullary signal changes in T2 or T1 images appeared more frequently in the CSM_lat+ group (Table 1).

Regional differences in diffusion anisotropy in the healthy cervical spinal cord

The tissue microarchitecture was not uniform in the cervical spinal tracts of healthy subjects. FA values were significantly higher in the dorsal (0.73±0.11) and lateral columns (0.72±0.13) than those in the ventral column of WM (0.58±0.10) (p<0.05), e.g., at C4/5. At the same level, there were no statistically significant differences in AD values between the different regions of white matter (dorsal column: 2.067±0.197×10^{-3}; lateral column: 2.081±0.191×10^{-3}; ventral column: 2.130±0.242×10^{-3}), whereas RD values were relatively lower in the somatosensory tracts (dorsal column: 0.596±0.243×10^{-3}; lateral column: 0.612±2.23×10^{-3}; ventral column: 0.770±0.177×10^{-3}) (Figure 3).

Changes in diffusion anisotropy were region-dependent

The diffusion indices of the myelopathic cord changed in all three columns in the white matter. For example, at the level of C4/5, the AD values were significantly higher in all
regions of the myelopathic cord (dorsal column: $3.139\pm0.447\times10^{-3}$; lateral column: $2.857\pm0.371\times10^{-3}$; ventral column: $3.356\pm0.266\times10^{-3}$) than those in the healthy cord (dorsal column: $2.067\pm0.197\times10^{-3}$; lateral column: $2.081\pm0.191\times10^{-3}$; ventral column: $2.130\pm0.242\times10^{-3}$; \(p<0.001\)). Increased RD values were also detected in the myelopathic cord (dorsal column: $1.500\pm0.487\times10^{-3}$; lateral column: $1.498\pm0.320\times10^{-3}$; ventral column: $1.610\pm0.080\times10^{-3}$) in comparison with the healthy cord (dorsal column: $0.596\pm0.210\times10^{-3}$; lateral column: $0.612\pm0.223\times10^{-3}$; ventral column: $0.770\pm0.177\times10^{-3}$; \(p<0.001\)).

By contrast, the FA changes in the myelopathic cord were region-dependent. As shown in Figures 1 and 2, a significant change in FA was observed in the dorsal (0.54±0.16) and lateral columns (0.51±0.13) of the myelopathic spinal cord under anterior compression; while the ventral column of myelopathic spinal cord were relatively spared (0.48±0.15). The regional differences in FA, observed in healthy spinal cord, were absent in the myelopathic cord.

*Diffusion anisotropy drop cephalic to the lesion*

As compared to healthy subjects, CSM patients with intact SEP (normal latency) showed the decrease of FA localized at the dorsal and lateral columns of white matter in myelopathic spinal cords. By contrast, the decrease of FA was much more extensive in patients with impaired SEP (prolonged latency), involving three columns of white matter (dorsal column: $0.57\pm0.05$; lateral column: 0.58±0.03; ventral column: 0.48±0.03) (Figure 4). In addition, it not only occurred at the compressive lesion, but also at the cephalic level to the lesion in all three columns of white matter (dorsal column: $0.57\pm0.06$; lateral column: $0.57\pm0.04$; ventral column: $0.53\pm0.02$) (Figures 4, 5).
**Discussion**

DTI was employed to evaluate regional deficits in the myelopathic spinal cord. It was found that the spinal tracts were not uniformly affected in CSM. The CSM-related changes of diffusion anisotropy were region-dependent, afflicting the dorsal and lateral columns and relatively sparing the ventral column. It was in good agreement with histopathological findings under clinical autopsy examination [1].

In consistence with previous DTI studies of CSM[11-19], the present study demonstrated FA decrease and apparent diffusion coefficient (ADC) or mean diffusivity increase in CSM. The diffusivity changes in CSM reflect the increase in the strength of water molecule movement in the enclosed spinal cord when passing through a narrow canal. This increase may be part of the spinal cord’s initial adaptation under chronic compression in a progressive stenotic canal. The unconstrained water molecules in the myelopathic cord present the decrease of diffusion anisotropy under DTI examination.

AD and RD of the myelopathic cord were elevated in all three columns, and they did not show the same regional differences as those observed for FA. By contrast, FA pattern of the myelopathic cord was more compatible with histopathological features of previously published clinical autopsy studies [4,5]. FA appeared to reflect demyelination and axon damage more appropriately in CSM cases in comparison with AD and RD, although they were once used to detect microstructural changes in other spinal cord disorders, e.g., multiple sclerosis [9,10].

Clinically, the prolonged latency of SEP has been reported as an indicator for the poor prognosis of CSM after surgeries [4]. It was also found in a rat model that the normal or prolonged latency of SEP were in a good association with the severity of microstructural damages in the chronic compressive spinal cord [37]. In this study, diffusion MR imaging of spinal tracts unveiled that a decrease of FA at the cephalic level of myelopathic cord to the
compressive lesion indicated anterograde degeneration of somatosensory spinal tract, so-called Wallerian degeneration. This finding specifically provided the structural basis of prolonged latency of SEP in myelopathic human spinal cord. The JOA assessment is a global assessment for myelopathic severity [28]. However, spinal tracts are not uniformly affected in CSM, which cannot be reflected by a global assessment such as the JOA score. We did not find a difference in the sums of the JOA scores between CSM patients with or without prolonged latency. The value of the JOA score system in predicting surgical outcomes for CSM patients remains controversial [29]. As such, the regional analysis of diffusion MR images of the myelopathic cord might provide additional information to the current assessments, including the JOA score system, anatomic MR images and SEP, to formulate a comprehensive evaluation approach for clinical diagnosis and prognosis of CSM.

The severity of cord compression did not necessarily correlate with the signs and symptoms of CSM patients [30-32]. For example, there were cases with significant cord compression but without any neurological signs, or with mild cord compression but with development of neurological signs [33,34]. We found that there was no significant difference in the compression ratio of the myelopathic cord with or without prolonged latency in SEP, although there was a difference in the extent of cord damage between the two groups.

The clinical significance of T2 hyperintensity [29,35-38], and T1 hypointensity [39,40] was also documented in CSM patients. It was found that signal changes of the myelopathic cord were commonly present in CSM patients with prolonged latency of SEP. However, such signal changes in the cervical cord are non-specific, which covers a wide spectrum of pathological changes such as edema and hemorrhage (T2 hyperintensity) or cyst (T1 hypointensity). In contrast, DTI might provide more specific information on demyelination and axon damage in spinal tracts of the myelopathic cord.

In summary, DTI could provide a more sensitive and specific measurement for spinal tract damage in CSM than the conventional clinical, electrophysiological and radiological
DTI in CSM

1 assessments. Limited to a cross-sectional observation on a small number of CSM patients, the
2 exact diagnostic and prognostic values of DTI in CSM needs to be verified in a large-scale,
3 prospective study in the near future.
4
13 DTI in CSM

References


Figure Legends

**Figure 1.** The representative anatomic (A, F), diffusion MR images (B, C, G, H) and fiber tractography (D, E, I, J) of healthy (A–E) and myelopathic spinal cord (F–G). The regions of interest are defined based on the anatomy of the spinal cord in axial slices of the fractional anisotropy (FA) mapping (C, F). The gray matter (“*”) is defined as the central portion of the spinal cord with low gray scale on the FA map; then the ventral, lateral and dorsal columns of white matter are defined accordingly (C, H). Compared with the healthy cord, the tracking of water molecules movement significantly changes in the dorsal and lateral aspects of the myelopathic cord (I, J: white arrow).

**Figure 2.** The characterization of diffusion properties of healthy (left column) and the myelopathic spinal cord (right column) in the ventral, lateral and dorsal columns of white matter by anatomic levels along the length of the cervical spine. FA values significantly drop in the dorsal and lateral columns with relative sparing in the ventral column (shown in the upper row). Yet the AD and RD values are increased in all three columns of white matter (shown in the middle and lower rows). Abbreviations: FA: fractional anisotropy; AD: axial diffusivity; RD: radial diffusivity.

**Figure 3.** Gross morphometry of the spinal cord was evaluated via measurement of the compression ratio (anterior-posterior distance divided by transverse distance of the spinal cord). Generally, the compression ratio decreases in the myelopathic spinal cord. There is no statistically significant difference in the compression ratio between the myelopathic cord with (CSM_lat+) or without prolonged latency (CSM_lat-). (“**” Indicates statistical significance at p<0.05 with one-way ANOVA and post-hoc test).

**Figure 4.** The representative anatomic (A, E, I), diffusion MR images (B, F, J) and fiber tractography (C, D, G, H, K and M) of healthy (A–D) and the myelopathic spinal cord with (I–M) or without prolonged latency (E–H) at C1/2 level cephalic to myelopathic lesions. The myelopathic cord with prolonged latency demonstrate significantly lower FA mapping (J) and disturbance of fiber tracking (K, M) at the upper cervical region cephalic to the chronic compressive lesions compared with those without prolonged latency, as well as the healthy cord.

**Figure 5.** A comparison of the diffusion anisotropy of the dorsal (A), lateral (B) and ventral columns (C) of white matter among healthy and the myelopathic spinal cord with (CSM_lat+) or without prolonged latency (CSM_lat-). In the CSM_lat- group, FA drops mainly in the dorsal and lateral columns. Yet in the CSM_lat+ group, the changes in FA are much more extensive not only at the lesion level but cephalic to the lesion, and involved in all three columns (“**” Indicates statistical significance at p<0.05 with one-way ANOVA and post-hoc test).
Table 1 Summary of clinical and radiological data of the patients of cervical spondylotic myelopathy

<table>
<thead>
<tr>
<th>Case</th>
<th>Gender/age</th>
<th>From symptom onset to imaging</th>
<th>JOA score</th>
<th>Hoffman Sign</th>
<th>Finger Escape Sign</th>
<th>Babinski Sign</th>
<th>Ankle Clonus</th>
<th>Romberg Test</th>
<th>Spinal canal</th>
<th>Spinal cord</th>
<th>Stenotic level(s)</th>
<th>SEP</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>F/44</td>
<td>3 years</td>
<td>10.0</td>
<td>-</td>
<td>1</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C5–6</td>
</tr>
<tr>
<td>2</td>
<td>F/46</td>
<td>5 years</td>
<td>11.5</td>
<td>+</td>
<td>2</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C4–5, C5–6</td>
</tr>
<tr>
<td>3</td>
<td>M/54</td>
<td>5 years</td>
<td>9.5</td>
<td>-</td>
<td>0</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID, spondylosis</td>
<td>-</td>
<td>Focal hyperintense signals</td>
<td>C5–6</td>
</tr>
<tr>
<td>4</td>
<td>F/61</td>
<td>4 years</td>
<td>11.0</td>
<td>+</td>
<td>1</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C4–5, C5–6</td>
</tr>
<tr>
<td>5</td>
<td>M/57</td>
<td>8 years</td>
<td>8.5</td>
<td>+</td>
<td>1</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C3–4</td>
</tr>
<tr>
<td>6</td>
<td>F/58</td>
<td>4.5 years</td>
<td>10.0</td>
<td>-</td>
<td>1</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID, spondylosis</td>
<td>-</td>
<td>Focal hyperintense signals</td>
<td>C3–4, C4–5, C5–6</td>
</tr>
<tr>
<td>7</td>
<td>M/61</td>
<td>8 years</td>
<td>9.5</td>
<td>+</td>
<td>1</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C3–4</td>
</tr>
<tr>
<td>8</td>
<td>F/68</td>
<td>7 years</td>
<td>10.0</td>
<td>-</td>
<td>4</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C4–5</td>
</tr>
<tr>
<td>9</td>
<td>M/71</td>
<td>&gt;10 years</td>
<td>11.0</td>
<td>+</td>
<td>2</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C3–4, C4–5, C5–6</td>
</tr>
<tr>
<td>10</td>
<td>M/72</td>
<td>10 years</td>
<td>9.0</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C4–5, C5–6</td>
</tr>
<tr>
<td>11</td>
<td>F/54</td>
<td>6 years</td>
<td>10.0</td>
<td>+</td>
<td>3</td>
<td>+</td>
<td>+</td>
<td>N.T.</td>
<td>PID, spondylosis</td>
<td>Multi-segmental hypointense signals</td>
<td>Multi-segmental hyperintense signals</td>
<td>C3–4, C4–5, C5–6</td>
</tr>
<tr>
<td>12</td>
<td>F/58</td>
<td>3 years</td>
<td>8.5</td>
<td>+</td>
<td>1</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C4–5, C5–6</td>
</tr>
<tr>
<td>13</td>
<td>M/65</td>
<td>5 years</td>
<td>11.0</td>
<td>+</td>
<td>3</td>
<td>+</td>
<td>+</td>
<td>N.T.</td>
<td>PID, spondylosis</td>
<td>-</td>
<td>Focal hyperintense signals</td>
<td>C4–5, C5–6</td>
</tr>
<tr>
<td>14</td>
<td>M/66</td>
<td>7 years</td>
<td>9.0</td>
<td>+</td>
<td>2</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C5–6</td>
</tr>
<tr>
<td>15</td>
<td>F/74</td>
<td>10 years</td>
<td>8.5</td>
<td>+</td>
<td>4</td>
<td>+</td>
<td>+</td>
<td>N.T.</td>
<td>PID</td>
<td>-</td>
<td>-</td>
<td>C3–4</td>
</tr>
</tbody>
</table>

Abbreviations: SEP: Somatosensory evoked potentials; PID: protrusion of intervertebral disc; Note: “+” indicates the presence (or absence) of pathological signs. Finger escape signs were graded as: “0” all, none deficiency; “1” little finger unable to hold adduction; “2” little or little and ring finger unable to assume adduction; “3” little and ring finger unable to assume adduction or full extension; “4” little, right and middle unable to assume adduction or full extension.
**Supplementary Table** The scanning parameters of anatomic and diffusion MR image

<table>
<thead>
<tr>
<th>Scanning mode</th>
<th>Imaging parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sagittal T1W and T2W images</td>
<td>Field of view (FOV) = 250×250 mm, slice thickness = 3 mm, slice gap = 0.3 mm, fold-over direction = Feet/Head (FH), Number of excitation (NEX) = 2, resolution = 0.92×1.16×3.0 mm³ (T1W) and 0.78×1.01×3.0 mm³ (T2W), recon resolution = 0.49×0.49×3.0 mm³, Time of echo (TE) / Time of Repetition (TR) = 7.2 / 530 ms (T1W) and 120 / 3314 ms (T2W).</td>
</tr>
<tr>
<td>Axial T1W and T2W images</td>
<td>FOV = 80×80 mm, slice thickness = 7 mm, slice gap = 2.2 mm, fold-over direction = anterior/posterior (AP), NEX = 3, resolution = 0.63×0.68×7.0 mm³ (T1W) and 0.63×0.67×7.0 mm³ (T2W), recon resolution = 0.56×0.55×7.0 mm³ (T1W) and 0.63×0.63×7.0 mm³ (T2W), TE / TR = 8 / 1000 ms (T1W) and 120 / 4000 ms (T2W).</td>
</tr>
<tr>
<td>Axial diffusion tensor images</td>
<td>FOV = 80×80 mm, slice thickness = 7 mm, slice gap = 2.2 mm, fold-over direction = AP, NEX = 3, resolution = 1×1.26×7.0 mm³, recon resolution = 0.63×0.64×7.0 mm³, TE / TR = 60 ms / 5 heartbeats</td>
</tr>
</tbody>
</table>
Figure 5

A: Fractional anisotropy of dorsal column

B: Fractional anisotropy of lateral column

C: Fractional anisotropy of Ventral column