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Review Article

Hypoxia-Induced Oxidative Stress in Ischemic Retinopathy

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Oxidative stress plays a crucial role in the pathogenesis of retinal ischemia/hypoxia, a complication of ocular diseases such as diabetic retinopathy (DR) and retinopathy of prematurity (ROP). Oxidative stress refers to the imbalance between the production of reactive oxygen species (ROS) and the ability to scavenge these ROS by endogenous antioxidative systems. Free radicals and ROS are implicated in the irreversible damage to cell membrane, DNA, and other cellular structures by oxidizing lipids, proteins, and nucleic acids. Anti-oxidants that can inhibit the oxidative processes can protect retinal cells from ischemic/hypoxic insults. In particular, treatment using anti-oxidants such as vitamin E and lutein, inhibition of nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase) or related signaling pathways, and administration of catalase and superoxide dismutase (SOD) are possible therapeutic regimens for DR, ROP, and other retinal ischemic diseases. The role of oxidative stress in the pathogenesis of DR and ROP as well as the underlying mechanisms involved in the hypoxia/ischemia-induced oxidative damage is discussed. The information provided will be beneficial in understanding the underlying mechanisms involved in the pathogenesis of the diseases as well as in developing effective therapeutic interventions to treat oxidative stress-induced damages.

1. Introduction

The retina is highly susceptible to oxidative damage by reactive oxygen species (ROS). Besides having the highest oxygen consumption in the body [1], the retina is also prone to photo-oxidation due to its constant exposure to incoming light. The high oxygen consumption and endless light exposure in the retina may in turn generate ROS. Moreover, the high lipid content in the retina (due to abundant polyunsaturated fatty acids in the photoreceptor outer segment) makes it prone to lipid peroxidation. During pathological conditions such as retinal ischemia, the imbalance between the production of ROS and the ability to scavenge these ROS by endogenous antioxidant systems is exaggerated. ROS triggers several signaling pathways, affects DNA and lipids inside the cell, and subsequently leads to cell death. Antioxidants that can inhibit or prevent the oxidative processes can protect retinal cells from ischemic damage.

2. Sources of Oxidants

A free radical is an atom, a molecule, or an ion having unpaired electrons. Due to the presence of the “free” electron in the outer shell, free radical is chemically very unstable and reactive. In order to achieve stability, free radical will participate in further reduction and oxidation reactions. Therefore, production of one free radical leads to further radical formation via sequential chain reactions [2]. Free radicals that are derived from oxygen are ROS, being one of the major contributors of oxidative stress. They include superoxide anion (O$_2^-$), perhydroxyl radical (also known as hydroperoxy radical, HO$_2$), and hydroxyl radical (OH). Superoxide is formed by either the enzymatic reduction of oxygen by nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase) or nonenzymatic reaction of mitochondrial respiration [3]. It is converted into hydrogen peroxide (H$_2$O$_2$) enzymatically by superoxide dismutase (SOD) or
nonenzymatically to H$_2$O$_2$ and singlet oxygen [3, 4]. Singlet oxygen is a powerful oxidant and is able to oxidize lipids, proteins and nucleic acids [5]. Perhydroxyl radical is the protonated form of superoxide, acting as an oxidant and reacts with polyunsaturated fatty acids in the membrane lipid bilayer, thereby initiating lipid peroxidation. Hydroxyl radical is another reactive radical which can be formed either from the reaction of singlet oxygen and H$_2$O$_2$ by the Haber-Weiss reaction [6] or from the interaction between H$_2$O$_2$ and reduced transition metals such as ferrous ions by the Fenton reaction [3, 6, 7]. Generation of superoxide, singlet oxygen and hydroxyl radical causes detrimental effects on the physiological states of cells by cleaving covalent bonding in protein and carbohydrates, leading to lipid peroxidation and compromising the integrity of cell membrane [3, 6, 7].

Another free radical, nitric oxide (NO) is generated from the oxidation of the guanidine group of L-arginine catalyzed by the enzyme nitric oxide synthase (NOS). NO is in fact a signaling molecule. Moreover, NO can react with superoxide to form a strong oxidant, peroxynitrite (ONOO$^-$), one of the reactive nitrogen species (RNS). Similar to ROS, excessive production of RNS (nitrosative stress) results in nitrosylation, causing deleterious effects such as lipid peroxidation, DNA damage and SOD inactivation [8]. Therefore, the toxic effects of ONOO and NO should also be taken into account in developing the antioxidative strategy [9].

3. Anti-Oxidative Mechanisms and Oxidative Stress

In physiological condition, ROS and RNS are products of normal cellular metabolism. ROS can be efficiently scavenged by the intrinsic antioxidant defense mechanisms. Manganese superoxide dismutase (MnSOD), copper/zinc superoxide dismutase (Cu/Zn SOD), catalase, and glutathione peroxidase (GPx) are members of the enzymatic antioxidants while ascorbic acid (vitamin C), α-tocopherol (vitamin E), glutathione (GSH), and β-carotene are nonenzymatic antioxidants. For example, MnSOD and Cu/Zn SOD convert superoxide to H$_2$O$_2$, which is then efficiently changed to water by GPx or catalase [10, 11]. On the other hand, vitamin E, vitamin C and β-carotene act as radical scavengers. Vitamin E also reduces and removes lipid peroxyl radical, a product of the reaction between lipid radical (a result of electron extraction from polyunsaturated fatty acid by hydroxyl radical) with molecular oxygen and an initiator of lipid peroxidation within the cell membrane. Subsequently, vitamin E is regenerated by vitamin C. Therefore, both vitamin E and vitamin C help to cease the lipid peroxidation chain reactions involving lipid peroxyl radicals [12]. GSH, a major cellular redox buffer, can directly scavenge hydroxyl radical and singlet oxygen. Through the action of GPx, GSH can also remove H$_2$O$_2$ and lipid peroxides. Moreover, GSH helps to regenerate both vitamin E and vitamin C back to their active forms. Normally, a balance is maintained between the levels of oxidants and antioxidants, which is essential for survival.

During ischemic injury, there is overproduction of free radicals that in turn leads to further production of radicals through chain reactions [10, 12]. The imbalance between the overproduction and scavenging of ROS/RNS leads to oxidative stress/nitrosative stress. The excess ROS can damage cellular lipids, proteins, or DNA and in turn inhibit their normal function. ROS also disturbs normal functioning of mitochondria by opening the mitochondrial membrane permeability transition pores (PTPs), leading to energy uncoupling and further ROS production [13]. This also induces the release of cytochrome c, leading to apoptosis [10, 14, 15]. In addition, superoxide reacts with NO to form ONOO$^-$ anion and in turn causes oxidation, lipid peroxidation, and subsequently cellular damage [8]. Accumulation of ONOO and free radicals in the nucleus also leads to DNA damage [9]. Indeed, an increased level of nitrotyrosine, the nitration product of ONOO, is observed after retinal ischemia/reperfusion (I/R) injury and is attributed to cell death [16, 17].

Retinal ischemia is a common feature in ischemic retinopathy such as diabetic retinopathy (DR) and retinopathy of prematurity (ROP). In both ocular diseases, there is initially an impairment in the normal retinal blood supply while the subsequent formation of abnormal new blood vessels (retinal neovascularization) further worsens the condition. The presence of ischemia and consequently oxidative stress makes the already vulnerable retina more prone to oxidative damage.
Oxidative stress causes the impairment of mitochondrial function by damaging the inner membrane of mitochondria, which leads to imbalance in the electron transport chain and consequently leads to further overproduction of superoxide, ONOO and hydroxyl radicals [35, 36]. Studies have shown that ROS also impairs mitochondrial function and transport machinery by damaging the mitochondrial DNA and increases apoptosis of retinal capillary cells during DR [37–39]. Increased ROS levels also trigger the release of cytochrome c, which in turn damages the mitochondrial membrane potential and initiate apoptosis by the activation of caspase-9 and caspase-3 [40–43]. Apoptosis in DR is also regulated by the family of Bcl-2 signaling proteins [43–45].

Several signaling pathways are involved in the pathogenesis of DR. Mitogen-activated protein kinase (MAPK) has been shown to play a role in hyperglycemia-induced cell death in DR [46]. In a cell culture model of hyperglycemia and hypoxia, phosphorylation of c-Jun N-terminal kinases (JNK) and p38 MAPK is induced, leading to overproduction of ROS and disruption of tight junctions in ARPE-19 cells [47]. In streptozotocin-induced diabetic mice, activation of cannabinoid-1 receptor contributes to DR by increasing MAPK activation, oxidative stress, and inflammatory signaling [48]. In addition, PARP and nuclear factor-kappa B (NFκB) are also downstream effectors of oxidative stress. PARP is a DNA nick-sensor enzyme that is activated by DNA single-strand breaks. It is involved in DNA repair, gene transcription, cell death, and apoptosis and acts as a coactivator in NFκB signaling pathway [34, 49, 50]. Increased levels of oxidative and nitrosative stress induce DNA single-strand break and activates PARP. This in turn inhibits the activity of GAPDH by poly(ADP-ribosylation) and consequently leads to endothelial dysfunction in diabetic conditions [34]. Therefore, inhibition of PARP could decrease the activation of PKC isoforms, hexosamine pathway flux and AGE formation induced by hyperglycemia [34]. As oxidative stress activates PARP, which in turn initiates the activation NFκB, application of PARP inhibitor could successfully inhibit hyperglycemia-activated NFκB signaling pathway in endothelia cell culture [51]. Also, inhibition of PARP or NFκB reduces the hyperglycemia-induced cell death in retinal endothelial cells [50]. NFκB also plays a role in apoptosis by triggering the Notch-1/Akt and PI3k/Akt signaling pathways. Increased activation of PARP, cleaved caspase-3 as well as reduced expression of Notch1 and pAkt have been observed, which consequently leads to apoptosis in hyperglycemic retina [49].

5. Oxidative Stress and the Pathogenesis of Retinopathy of Prematurity (ROP)

Similar to DR, retinal ischemia/hypoxia is a feature in ROP, which is a leading cause of blindness in children worldwide. Retinopathy of prematurity (ROP) was first described as a disease of prematurity [52, 53] and contributed to 50% of infant blindness in 1950. The percentage dropped to 4% in 1965 after identification of supplemental oxygen use as a

![Diagram of the relationship between oxidative stress and DR](image-url)
risk factor. Yet, with improved neonatal care and increased survival of very low birth weight preterm infants the incidence of ROP increased in recent years. ROP is now a major cause of childhood blindness in developed and developing countries. Latest estimates from National Eye Institute showed that 1,100–1,500 infants (<5% infants ≤1.25 kg at birth, <31 weeks of gestation) develop severe ROP that requires treatment. Despite treatment, 400–600 infants (40% with ROP, NIH figure) become legally blind. The resulting long-term disability and severely affected quality of life in ROP patients pose intense burden on the healthcare system worldwide as the population ages, making ROP a major public health issue.

ROP is an ocular disease characterized by vascular abnormalities induced by two phases of pathological changes. The first phase starts with the cessation of retinal vessel growth after premature birth. As premature infants are exposed to high oxygen inside the incubator after birth, a condition of relative hyperoxia happens, leading to downregulation of vascular endothelial growth factor (VEGF) and a subsequent regression of developed retinal vessels. After the cessation of oxygen therapy, infants are returned to normal oxygen tension and a condition of relative hypoxia occurs. In addition, the metabolic demand of the relatively vascular-depleted retina is now higher as the infant grows. Therefore, the increased demand for oxygen poses a hypoxic injury to the retina. The relatively hypoxic condition of the retina triggers the abnormal proliferation of vessels and leads to neovascularization; this is the second phase of ROP progression [54]. Hypoxia-induced oxidative stress therefore, plays a crucial role in the pathogenesis of ROP (Figure 2).

It has been shown that the production of enzymatic and nonenzymatic antioxidants such as SOD, catalase, GPx, vitamin C, and vitamin E increases dramatically in the late stage of gestation [55, 56]. Yet, premature infants are born before term; they are therefore, more susceptible to oxidative stress-induced damage due to the inadequate levels of antioxidants in premature birth and the inability of antioxidant synthesis during the high-oxygen intensive care period [54]. Indeed, the levels of 8-hydroxy 2-deoxyguanosine (8-OHdG), an indicator of oxidative stress, in leukocyte and urine in ROP infants are significant higher than the full-term infants, confirming the presence of oxidative stress in ROP patients [57].

During the two phases of ROP, the retina is subjected to fluctuating oxygen tensions, resulting in retinal hypoxia that triggers the overproduction of ROS, which activates NADPH oxidase and attributes to intravitreal neovascularization by the activation of signaling pathway such as JAK/STAT [58, 59]. On the other hand, inhibition of NADPH oxidase and JAK/STAT signaling pathway reduce the level of cleaved caspase-3 and hence, apoptosis and neovascularization in ROP retina [58, 59]. Increase intake of omega-3-polyunsaturated fatty acids has been shown to reduce the vaso-obliteration and neovascularization by suppressing the TNF-α in ROP retina [60]. Antioxidants such as vitamin E could also decrease the vaso-obliteration in ROP retina by its ROS scavenging property [61]. All this evidence points to the importance of oxidative stress in the pathogenesis of ROP.

Nitro-oxidation also contributes to the pathogenesis of ROP. Increased level of NOS, which contributes to the increase of NO production, is observed in neonatal retina exposed to hypoxia [62]. In an animal model of ROP, increased formation of ONOO− and apoptosis of endothelial cell were seen in association with increased tyrosine nitration of PI3K, cleaved caspase-3, activation of p38 MAPK signaling pathway, and decreased Akt phosphorylation. However, blocking tyrosine nitration of PI3K with epicatechin or N-acetylcysteine reverses the nitro-oxidation-induced pathogenesis [63]. In addition, application of NOS inhibitor or gene deletion of endothelial NOS effectively reduces the severity of ROP in mice, indicating the crucial role of nitro-oxidative stress in ROP [64]. Moreover, melatonin administration reduces the production of VEGF and NO as well as promotes cell proliferation, consequently reducing hypoxia-associated retinopathy [62].

VEGF signaling is a dominant pathway involved in the hypoxia-induced neovascularization and retinopathy in ROP. In the retina, VEGF is primarily secreted by Muller cells and astrocytes. In phase II of ROP, relative hypoxia induces VEGF production and hence promotes pathological vessel proliferation [65, 66]. IGF-1 is another determining factor in the pathogenesis of ROP and is critical in normal retinal vessel growth. IGF-I is a permissive factor of VEGF activation of Akt survival signaling in the growth of endothelial cells in phase I of ROP [67]. In preterm infants after birth, lower levels of IGF-I reduce Akt activation and endothelial cell survival, which makes the maturing avascular retina becomes hypoxic and leads to accumulation of VEGF in the vitreous [67]. IGF-1 level rises with high levels of VEGF, which triggers neovascularization through the activation of MAPK and Akt signaling pathways in phase II of ROP [67, 68]. Inhibition of IGF-1 and VEGF are capable to hinder the neovascularization.
6. Antioxidants in Ischemic Retinopathies

DR and ROP are blinding disorders that affect working-age adults and premature infants, causing enormous burden on the society. Yet, the treatment options are limited to laser photocoagulation and anti-VEGF therapy. Pathologically, they both follow a similar pattern as in ischemic retinopathy. As oxidative stress play a pivotal role in the pathogenesis of ischemic retinopathy, agents that can reduce the formation of ROS or increase the antioxidant mechanisms to counteract oxidative stress are beneficial to the situation. Examples of some of these agents are discussed below.

Catalase and SOD are potent ROS scavengers to block oxidative stress in the retina. SOD catalyzes the conversion of superoxide to oxygen and H₂O₂ while catalase prevents the formation of hydroxyl radical resulting from the reaction of H₂O₂ and ferrous ions [69]. Gene transfer of plasmids encoding MnSOD or catalase complexed with liposomes reduces apoptosis of retinal vascular cell and prevents retinal degeneration induced by retinal I/R [69]. In addition, pretreatment of intravitreal recombinant adenoassociated virus containing the catalase gene (AAV-CAT) efficiently increases the catalase activity and decreases the levels of H₂O₂, 8-OHdG and nitrotyrosine as well as attenuates the I/R-induced retinal function loss, suggesting the beneficial effects of catalase in ischemia/reperfusion-related eye diseases by reducing oxidative stress [70].

As discussed earlier, nitrosative stress also causes detrimental damages during retinal ischemia. NADPH oxidase catalyzes the formation of superoxide from oxygen and NADPH. In ischemic injury in mice, gene deletion of NADPH oxidase reduces the production of ROS, retinal cell loss and glial cell activation, together with an inhibition in the activation of extracellular-signal-regulated kinase (ERK) and NFκB signaling pathways [71]. Moreover, blockade of astrocytic NFκB signaling or administration of NADPH oxidase inhibitors reduces retinal ganglion cell death cocultured with astroglia under hypoxia [72]. Gene deletion of astroglial NF-κB also reduced oxidative stress in retinal I/R in mice [72, 73]. Most importantly, inhibition of NFκB activation by either gene deletion of NADPH oxidase or application of apocynin reduced the retinopathy in ROP and DR animal models [74].

Edaravone is a free radical scavenger which directly scaven ges hydroxyl radical and inhibits lipoxygenase activity [75]. It has been shown that application of edaravone can reduce oxidative stress-induced damage by lowering the levels of malondialdehyde (MDA), raising enzymatic activity of SOD, reducing apoptosis of retinal neurons, and preventing retinal functional deterioration in retinal I/R [76]. This suggests that edaravone protects the eye from I/R injury and may carry therapeutic potential in treatment of I/R-induced eye disorders.

Vitamin E is well known to have potent anti-oxidative properties in ocular disease related to retinal I/R, including ROP and diabetic retinopathy [77–81]. It is a lipid peroxyl radical scavenger via breaking the lipid peroxidation chain and acts against oxidative stress [77–81]. In an animal model of retinal I/R injury, reduced levels of oxidative stress marker such as MDA together with an inactivation of caspase-3 but increased levels of GSH were noted with vitamin E treatment [77–81]. In experimental DR models, oral intake of antioxidants, including vitamin E, significantly reduced lipid peroxidation and arrested retinal abnormalities in streptozotocin (STZ)-induced diabetic rats [82]. Intake of micronutrients based on the AREDS formula (vitamin E, ascorbic acid, beta-carotene, zinc, and copper) inhibited the increases in nitrotyrosine and inducible NOS and decreases in MnSOD in STZ-induced diabetic rats [83]. In ROP models, vitamin E can reduce the area of vaso-obliteration [61, 84], preserve retinal function [85] and protect the reduction of outer plexiform thickness [86]. Most importantly, vitamin E supplementation decreased the risk of severe retinopathy and blindness in very low birth weight infants [81]. All this evidence points to a beneficial role of vitamin E in ischemic retinopathy.

Numerous studies have investigated the role of erythropoietin (EPO) in ischemic retinopathy. EPO is a glycoprotein hormone that is involved in the pathogenesis of ROP. EPO level decreases and contributes to vessel obliteration in phase I of ROP; therefore, early administration of exogenous EPO minimizes vessel obliteration and subsequent hypoxia-induced neovascularization and neuron apoptosis by the activation of prosurvival NFκB signaling pathway via the EPO receptor [87]. However, the level of EPO mRNA increases during the neovascularization phase when late EPO application promotes pathologic neovascularization [87]. In addition, increased level of VEGF leads to vaso-obliteration by suppression EPO expression through the Janus kinase/STAT signaling pathway. Application of exogenous EPO inhibits the activation of STAT3 and hence, reduces the area of vaso-obliteration and consequently neovascularization in phase II of ROP [88]. Furthermore, administration of EPO siRNA suppresses neovascularization during phase II of ROP. Therefore, the time of exogenous treatment/inhibition is very critical to the prevention of ROP pathogenesis [89]. Similarly, EPO also exerts its anti-oxidative and protective effects in DR. Low-dose EPO inhibits oxidative stress and nitro-oxidative stress in experimental diabetic retina by reducing Ang-2 expression and pericyte loss as well as restoring pAkt and heat shock protein-27 levels [90]. In addition, EPO therapy decreases the formation of acellular capillaries and the loss of pericytes, which prevents the early vascular changes in DR [90]. EPO treatment also decreases VEGF levels and protects microvascular and neuronal damage in the experimental diabetic retina [91].

Lutein is a member of xanthophyll family of carotenoids and is contained in dark green leafy vegetables, such as spinach and kale [92–94]. It is a powerful anti-oxidant due to its unique chemical structure. It is characterized by having a hydroxyl group attached to each end of the molecule, making it more hydrophilic, and therefore, lutein reacts more strongly with singlet oxygen than other carotenoids [95, 96]. Increasing evidence has shown that lutein is neuroprotective in retinal I/R injury. We show that lutein prevents the increase of nitrotyrosine and PAR and hence, apoptosis and cell loss in inner retinal neuron in an animal model of retinal
We further show that lutein can directly protect RGC from H$_2$O$_2$-induced oxidative stress and cobalt chloride (CoCl$_2$)-induced hypoxia in vitro [97]. In retina of diabetic mice, lutein reduces the production of ROS, inactivates the NFKB signaling pathway, and decreases the levels of oxidative markers, thereby preserving the retinal function [98]. Lutein also prevents ERK activation, synaptophysin reduction, BDNF depletion, and consequently neuronal loss in the diabetic retina [99]. However, supplementation of anti-oxidants, including lutein, is ineffective in preventing ROP in preterm infants [100, 101]. Yet, the protective effects of lutein in ROP are still debatable due to the complicated pathogenesis of ROP.

## 7. Future of Antioxidant Treatment in Ischemic Retinopathy

DR and ROP are two examples of ischemic retinopathy. As mentioned earlier, DR and ROP are blinding disorders that affect working-age adults and premature infants, respectively. The subsequent long-term disability and poor quality of life pose enormous burden on the society. Much research has been conducted, but with no satisfactory outcome. As discussed by Kowluru and Zhong [18], there are serious challenges although promising results are observed in experimental DR models. Firstly, rodents only display the early retinopathy stages but not the later changes seen in patients with diabetic retinopathy. Secondly, DR is a complex diabetic complication; identification of the best molecular pathway for targeted therapeutic studies is not easy. Yet, the administration of the AREDS formula containing various antioxidants to diabetic rats has provided a significant step forward for antioxidant treatment for DR [83].

In ROP, lutein, vitamin E and EPO have all shown hopeful results in experimental models. Similar to the negative results in antioxidant therapy in DR, lutein is not able to yield beneficial effects in two randomized controlled clinical studies [100, 101]. However, vitamin E and EPO have shown some promising outcomes clinically. An early meta-analysis concluded that there is a 52% reduction in the incidence of Stage 3+ ROP with vitamin E prophylaxis in extremely low birth weight (<1000 gm) infants [80]. In a later clinical study, it is concluded that vitamin E is proven to be effective in prophylaxis of ROP development, and vitamin E supplementation relates to lower incidence of ROP [102]. Recently, interventions such as increasing retinal EPO and vitamin E supplements have been suggested [103]. Although well-designed randomized clinical trials are necessary before these interventions can be put into clinical practice, they have provided strong evidence that antioxidant therapy does have a potential role in treatment of ROP.

## 8. Conclusion

Hypoxia-induced oxidative stress leads to irreversible retinal damage in ocular diseases in which retinal ischemia is a feature. Treatments that can decrease the production of ROS or increase the ROS scavenging ability of the system may be beneficial to the situation. The positive outcomes in administration of AREDS antioxidants in DR experimental models as well as EPO and vitamin E supplements in ROP infants have strongly indicated the potential efficacy of antioxidant in treating ischemic retinopathy with hypoxia-induced oxidative stress.

### List of Abbreviations

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<thead>
<tr>
<th>Abbreviation</th>
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<tr>
<td>8-OHdG</td>
<td>8-hydroxy 2-deoxyguanosine</td>
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<td>AGE</td>
<td>Advanced glycation end-product</td>
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<td>AMD</td>
<td>Age-related macular degeneration</td>
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<tr>
<td>AR</td>
<td>Aldose reductase</td>
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<tr>
<td>Cu/Zn SOD</td>
<td>Copper/zinc superoxide dismutase</td>
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<td>DR</td>
<td>Diabetic retinopathy</td>
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<td>EPO</td>
<td>Erythropoietin</td>
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<tr>
<td>GPx</td>
<td>Glutathione peroxidase</td>
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<tr>
<td>GSH</td>
<td>Glutathione</td>
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<tr>
<td>H$_2$O$_2$</td>
<td>Hydrogen peroxide</td>
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<tr>
<td>I/R</td>
<td>Ischemia/reperfusion</td>
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<tr>
<td>JNK</td>
<td>c-Jun N-terminal kinases</td>
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<td>MAPK</td>
<td>Mitogen-activated protein kinase</td>
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<td>MDA</td>
<td>Malondialdehyde</td>
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<td>MnSOD</td>
<td>Manganese superoxide dismutase</td>
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<td>NADPH</td>
<td>Nicotinamide adenine dinucleotide phosphate oxidase</td>
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<tr>
<td>NFκB</td>
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<tr>
<td>NO</td>
<td>Nitric oxide</td>
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<tr>
<td>NOS</td>
<td>Nitric oxide synthase</td>
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<tr>
<td>ONOO</td>
<td>Peroxynitrite</td>
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<td>PARP</td>
<td>Poly(ADP-ribose) polymerase-1</td>
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<tr>
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<td>Protein kinase C</td>
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<tr>
<td>PTP</td>
<td>Permeability transition pores</td>
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<td>RAGE</td>
<td>Receptor for advanced glycation end-products</td>
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<td>RNS</td>
<td>Reactive nitrogen species</td>
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<td>Retinopathy of prematurity</td>
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<tr>
<td>ROS</td>
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<tr>
<td>SOD</td>
<td>Superoxide dismutase</td>
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<td>VEGF</td>
<td>Vascular endothelial growth factor</td>
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### References


[102] C. Dani, I. Lori, F. Favelli et al., "Lutein and zeaxanthin supplementation in preterm infants to prevent retinopathy of

