



Title	The relationship between glucose metabolism, metabolic syndrome, and bone-specific alkaline phosphatase: A structural equation modeling approach.
Author(s)	Cheung, CL; Tan, KCB; Lam, KSL; Cheung, BMY
Citation	Journal of Clinical Endocrinology and Metabolism, 2013, v. 98 n. 9, p. 3856-3863
Issued Date	2013-08-20
URL	http://hdl.handle.net/10722/185985
Rights	This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License.

The relationship between glucose metabolism, metabolic syndrome, and bone-specific alkaline phosphatase: A structural equation modeling approach

Running title: Insulin resistance and alkaline phosphatase

¹Ching-Lung Cheung, PhD,

^{2,3}Kathryn CB Tan, MD, FRCP,

^{2,3}Karen SL Lam, MD, FRCP, FRACP,

^{1,3}Bernard MY Cheung, PhD, FRCP

¹Division of Clinical Pharmacology and Therapeutics, ²Division of Endocrinology

and Metabolism, ³Research Centre of Heart, Brain, Hormone and Healthy Aging,

Department of Medicine, Li Ka Shing Faculty of Medicine,

The University of Hong Kong, Pokfulam, Hong Kong, China

Correspondence:

Bernard MY Cheung,

University Department of Medicine,

Queen Mary Hospital,

102 Pokfulam Road, Hong Kong

Email: mycheung@hku.hk

Tel: +85222554347

Fax: +85228186474

Disclosure summary: The authors have nothing to disclose.

Words count:

Abstract: 239; Main text: 3195

Number of tables: 3

Number of figure (in black and white): 1

Abstract

Context: Serum alkaline phosphatase plays a role in vascular calcification. It is found in various tissues, whereas bone-specific alkaline phosphatase (BAP) more specifically reflects mineral metabolism. The relationship of serum alkaline phosphatase (total and bone-specific) with diabetes and metabolic syndrome, two major risk factors of vascular calcification, is largely unknown.

Objective: We aimed to investigate the relationships between glucose metabolism, components of MetS, and alkaline phosphatase.

Design: This was a cross-sectional study.

Setting: Nationally representative sample of the US population in 1999-2004.

Participants: 3,773 non-diabetic participants of the National Health and Nutrition Examination Survey 1999-2004.

Main Outcome Measures: Serum BAP and total alkaline phosphatase.

Results: In multivariable linear regression, HOMA2-IR ($\beta=0.068$), HOMA2-B ($\beta=0.081$), insulin ($\beta=0.065$), mean arterial pressure ($\beta=0.15$), and HDL-cholesterol ($\beta=0.209$) were positively associated with BAP, whereas HOMA2-IS ($\beta=-0.065$) was negatively associated with BAP. On the other hand, only mean arterial pressure and HDL-cholesterol were significantly associated with total alkaline phosphatase. Moreover, structural equation model revealed that hypertension, low HDL, and

insulin resistance had significant direct effects on serum BAP levels, whereas obesity and inflammation might have indirect effects on serum BAP levels. The overall model showed very good fit to the data (comparative fit index = 0.995, root mean square error of approximation = 0.037, and standardized root mean square residual = 0.006).

Conclusion: Glucose metabolism and MetS are significantly related to serum BAP levels. How BAP mediates vascular calcification in diabetes and MetS warrants further studies.

Keywords: bone alkaline phosphatase; insulin resistance; vascular calcification; metabolic syndrome

Introduction

Diabetes mellitus and metabolic syndrome (MetS) are major risk factors of vascular calcification (1). Patients with diabetes have more extensive coronary artery disease, and atherosclerotic plaque morphology in diabetic patients may be predisposed to rupture and thrombosis (2). Among patients with acute coronary syndromes, patients with co-morbidity diabetes or MetS have a greater lesion length, plaque burden, necrotic core, and calcium content among nonculprit lesions (3). Moreover, higher incidence and progression of coronary artery calcium have also been demonstrated in subjects with MetS or diabetes (4). However, the underlying mechanism is poorly understood.

Extracellular pyrophosphate and alkaline phosphatase have been identified as key molecules in vascular calcification (1). Pyrophosphate is a potent inhibitor of tissue mineralization and vascular calcification (5). Pyrophosphate is produced by an enzyme, ectonucleotide pyrophosphatasephosphodiesterase 1 (NPP1), which is located in the cell membrane. It is then transported across the cell membrane to the extracellular matrix by the progressive ankylosis protein (ANK). Bone-specific alkaline phosphatase (BAP) hydrolyzes pyrophosphate and so promotes tissue mineralization (6-7). Knockout mice deficient in NPP1 and ANK have increased

aortic calcification (5) and medial calcification (8), respectively. On the other hand, blocking the hydrolyzing action of BAP ameliorated the calcification in vascular smooth muscle cells from NPP1- and ANK-deficient mice (7). Thus, alkaline phosphatase has a critical role in calcification, and it has been extensively used as a marker of calcification (6-7) and exploited as therapeutic target of vascular calcification (7, 9). Previous studies also showed that serum alkaline phosphatase is a predictor of mortality (10-11) and cardiovascular events (11).

A number of studies have investigated the association between serum alkaline phosphatase and diabetes/MetS owing to its role as a hepatobiliary marker. However, serum alkaline phosphatase consists of several alkaline phosphatase isoforms from various tissues, such as liver, bone, and kidney, with a large proportion are derived from liver and bone. Previous studies showed that BAP reflects mineral metabolism with a higher sensitivity and specificity than total alkaline phosphatase (12). Therefore in the current study, we aimed to evaluate the relationship between glucose metabolism, metabolic syndrome, alkaline phosphatase, and BAP in a large nationally-representative population.

Materials and Methods

The current study utilized data from National Health and Nutrition Examination Survey (NHANES) 1999–2004 (13). The NHANES included a stratified multistage probability sample which represented the civilian non-institutionalized U.S. population. Selection was based on counties, blocks, households, and individuals within households. It also included non-Hispanic blacks and Mexican-Americans to provide an adequate estimate of these ethnic groups. Participants were required to sign a consent form before their participation, and ethical approval was obtained from the Human Subjects Committee of the U.S. Department of Health and Human Services.

Our analysis included non-pregnant participants aged 20 or above whose BAP and fasting glucose levels were available (n=5,057). We excluded participants with missing data in the multivariable models or without weight in the complex sampling model (n=757) and participants with diabetes (n=527, defined as a plasma glucose level ≥ 126 mg/dl [7.0 mmol/L] after fasting for a minimum of 8 hours, a glycohemoglobin (HbA1C) value $\geq 6.5\%$, self-reported diabetes or self-reported current use of oral hypoglycemic medication or insulin). After exclusion, 3,773 participants were included in the analysis.

BAP and total alkaline phosphatase

Serum BAP was measured using the Hybritech Tandem-MP ostease immunoenzymetric assay (1999-2001, 2003-2004) and the Beckman access ostease assay (2002). Since there were two methods in measuring BAP, therefore NHANES recommends the following regression equation to convert the Hybritech assay values to the Beckman assay values: Beckman assay serum BAP = $\exp(-0.5326 * 1.1139 * x - 0.7963 * (\max(0, x - 4.5151))) + 0.9660 * (\max(0, x - 4.9030))$, where $x = \log(\text{Hybritech assay serum BAP})$. This conversion was used in the present analysis.

Serum total AP was measured as a routine laboratory blood parameter using Hitachi Model 704 multichannel analyzer in NHANES 1999–2000 and Beckman Synchron LX20 in NHANES 2003–2004. In NHANES 2001–2002, they were measured by either Hitachi Model 704 multichannel analyzer or Beckman Synchron LX20, and the reported values had been adjusted by regression equations to allow comparison across the two methods.

Indices of glucose metabolism

Serum insulin, plasma glucose, and HbA1C were measured at the Diabetes Diagnostic Laboratory, University of Missouri–Columbia. Plasma glucose and serum insulin were measured from fasting blood samples using hexokinase enzymatic method and

radioimmunoassay, respectively. HbA1C was measured using Primus CLC330 and Primus CLC 385 analyzers (Primus Corporation, Kansas City, MO). The updated homeostasis model assessment (HOMA2) is more accurate than the original HOMA1, therefore Homeostasis model assessment calculator (14) was used to calculate the HOMA2 beta cell function (%B), insulin sensitivity (%S), and hepatic insulin resistance (IR).

Components of MetS

Lipids (total cholesterol, HDL-cholesterol, triglycerides) were measured enzymatically using Hitachi 704 Analyzer at the lipoprotein analytical laboratory, Johns Hopkins University School of Medicine. Seated systolic and diastolic blood pressures (mm/Hg) were measured using a mercury sphygmomanometer according to the American Heart Association and Seventh Joint National Committee recommendations. Mean arterial pressure (MAP, mm/Hg) was calculated as $\frac{2}{3}$ diastolic blood pressure + $\frac{1}{3}$ systolic blood pressure.

Independent variables

Age (years), gender (male/female), race/ethnicity (Hispanics, non-Hispanic white, non-Hispanic Black, others), smoking status (current/ former/ non-smokers), alcohol

use (drinkers/ non-drinkers), level of education (< high-school, high-school, > high-school), and physical activity (active/ sedentary) were assessed using a questionnaire. We used same definition for never, current and former smokers that is used in the National Health Interview Surveys (NHIS) conducted by Center for Disease Control. Individuals who had not smoked more than 100 cigarettes in their lifetime were considered non-smokers; those who had smoked more than 100 cigarettes in their lifetime were considered former smokers if they answered negatively to the question “Do you smoke now?” and current smokers if they answered affirmatively. Individuals who had 12 alcohol drinks or more per year were considered as drinkers. Participants who reported doing moderate or vigorous activity for at least 10 minutes over the past 30 days on the Physical Activity Questionnaire were considered to be physically active. Body mass index (BMI, kg/m²) was calculated as weight in kilograms divided by height in meters squared. However, as BMI is highly correlated with waist circumference (r>0.8), we only included waist circumference (a component of MetS) as the independent variable to avoid collinearity. C-reactive protein (CRP, mg/dL), gamma-glutamyl transpeptidase (GGT, U/L), aspartate aminotransferase (AST, U/L), alanine transaminase (ALT, U/L), N-terminal telopeptides, and fatty liver index (15) were also included as independent variables. Fatty liver index was derived using the equation: $FLI = (e^{0.953 \cdot \log_e(\text{triglycerides})})$

$+ 0.139 \cdot \text{BMI} + 0.718 \cdot \log_e(\text{ggt}) + 0.053 \cdot \text{waist circumference} - 15.745) / (1 + e^{0.953 \cdot \log_e(\text{triglycerides}) + 0.139 \cdot \text{BMI} +$

$0.718 \cdot \log_e(\text{ggt}) + 0.053 \cdot \text{waist circumference} - 15.745) * 100$. Details regarding the measurement of

the independent variables are available at the NHANES website (13).

Statistical analyses

Glycemic traits and components of MetS were analyzed as continuous and quartile variables. Multivariable linear regression was used to evaluate the association when glycemic traits and components of MetS were modeled as continuous variables.

Estimated mean and 95% CI of BAP or tissue non-specific alkaline phosphatase for each quartile were calculated using ANCOVA in a general linear model, and linear trend P was obtained using contrast coefficients for linear trend analysis. Structural

equation modeling (SEM) was used to estimate the model goodness-of-fit and directional estimates between variables. The predefined model used in the current

study was modified from an existing model of MetS from literature.(16) The global

Goodness of Fit Index for the SEM included the comparative fit index (CFI), root mean square error of approximation (RMSEA), and standardized root mean square

residual (SRMR). These indices are less sensitive to sample size and complexity than

χ^2 test for evaluating model fit (17). Values for the CFI of ≥ 0.90 and ≥ 0.95 indicate

acceptable and good fit, respectively (18). Values for the RMSEA values of ≤ 0.06

represent good fit while values >0.10 represent unacceptable fit (18). A SRMR ≤ 0.08 and <0.10 represent good and acceptable fit, respectively (18). SEM was carried out using R package “lavaan.survey” that performs SEM while taking account of the complex sampling of NHANES data. Variables with skewed distributions were log-transformed before analysis. Sample weights that account for the unequal probabilities of selection, oversampling, and non-response were applied for all analyses using complex sampling module in SPSS version 18.0 software (SPSS Inc, Chicago, IL). All values presented are weighted to represent the U.S. civilian population.

Results

Table 1 shows the sex-stratified characteristics of the 3,773 NHANES 1999-2004 participants who were free from diabetes. Women were older, more likely to be non-Hispanic white, non-smokers., non-active drinkers and had lower physical activity. They had higher HDL-cholesterol levels, CRP, HOMA2-IS, and HOMA2-B. Conversely they had lower waist circumference, BAP, total alkaline phosphatase, triglyceride levels, MAP, ALT, AST, GGT, HbA1C, fasting glucose levels, insulin concentrations, and HOMA2-IR.

Table 2 shows the association between glycemic traits and alkaline phosphatase. After adjustment of age, race/ethnicity, education, smoking, drinking, physical activity, and waist circumference in model 1, HOMA2-IR, HOMA2-B, fasting glucose, and insulin were positively associated with BAP, while HOMA2-IS was negatively associated with BAP ($P < 0.05$). After further adjustment of metabolic risk factors (CRP, total cholesterol, AST, ALT, GGT, triglycerides, HDL-cholesterol, and MAP) in model 2, the association between BAP and fasting glucose became insignificant. **Similar results were obtained when urinary N-terminal telopeptides and fatty liver index were adjusted in model 3.** On the other hand, three indices (HOMA2-IR, HOMA2-B, and insulin) and HOMA2-IS were positively and negatively associated with tissue non-specific alkaline phosphatase in model 1, respectively. All associations became insignificant after further adjustment in models **2 and 3**. Similar results were obtained when glycemic traits were categorized as quartiles (Supplementary Table 1).

Table 3 shows the association between components of MetS and alkaline phosphatase. MAP and HDL-cholesterol showed significant association with BAP, while waist circumference and triglycerides showed a trend toward significant association with BAP ($0.05 < P < 0.1$). On the other hand, MAP and HDL-cholesterol showed significant association with tissue non-specific alkaline phosphatase, while no

association was observed with waist circumference and triglycerides. Similar results were obtained when components of MetS were categorized as quartiles (Supplementary Table 2).

Figure 1 illustrates our hypothesized relationships between BAP, inflammation, and components of MetS. All latent variables (denoted by oval circle) were adjusted for age, gender, race/ethnicity, drinking, smoking, and physical activity. As expected, obesity has direct association with hypertension, inflammation, and insulin resistance; inflammation has direct association with insulin resistance; insulin resistance has direct association with vascular calcification and dyslipidemia; and dyslipidemia and hypertension have direct association with vascular calcification. All of the standardized regression coefficients shown in the figure were statistically significant ($P \leq 0.001$). The CFI was 0.995 (value >0.95 indicates good fit), RMSEA was 0.037 (value <0.06 indicates good fit), and SRMR was 0.006 (value <0.08 indicates good fit), these fit indices consistently indicate that our hypothesized model is a good fitting model.

Discussions

The associations and inter-relationships between insulin resistance, components of

MetS, and BAP were unraveled in the current study of the US-population using structural equation modeling in addition to multivariable linear regression. Although tissue non-specific alkaline phosphatase is known to correlate with insulin resistance and metabolic syndrome owing to its role as a hepatobiliary marker, our study clearly shows that both hyperinsulinemia, insulin resistance, and MetS are in fact more strongly associated with BAP.

Our study excluded participants with diabetes allowing us to study the relationship of hyperinsulinemia or insulin resistance with BAP, without the confounding effect of significant hyperglycaemia. Note that the direct effect of insulin on osteoblasts is a stimulation of osteoblast proliferation & differentiation, and an increase in BAP activity *in vitro* (19). On the other hand, streptozotocin-induced insulin deficiency (type 1 diabetes) in rats is associated with reduced BAP which is reversed by insulin therapy (20). In humans, osteopenia is observed in type 1 diabetic patients even with good control. On the other hand, high glucose *per se* has deleterious effect on osteoblast function *in vitro* (rat primary osteoblast cell culture) and decreases BAP secreted (21). Only subjects with poorly controlled type 2 diabetes are at risk of osteoporosis.

Our study sheds light on the mechanism of elevated BAP. While insulin resistance is known to be a risk factor for vascular calcification, the mechanism is not completely understood. In the current study, we showed that multiple HOMA2 indices and insulin levels are robustly associated with BAP rather than total alkaline phosphatase. Although total alkaline phosphatase could be elevated due to obesity, insulin resistance, fatty liver, and hepatosteatosis, no significant association was observed between HOMA2-IR and total alkaline phosphatase in the fully adjusted model. This may be explained by the fact that other liver markers included in the model were more specific to liver dysfunction; ALT, AST, and GGT also showed significant associations ($P < 0.05$) with HOMA2-IR in the fully adjusted model. When ALT, AST, and GGT were removed from the fully adjusted model, the association between alkaline phosphatase and HOMA2-IR became significant ($P = 0.015$). On the other hand, elevated BAP could be due to the presence of fatty liver. However, further adjustment for fatty liver index (15), a validated marker of fatty liver, did not change the association of HOMA2-IR, HOMA2-IS, HOMA2-B, and insulin with BAP (Table 2), suggesting that the association was independent of fatty liver. In addition, our study also showed that BAP is associated with multiple components of MetS, which suggested that hyperinsulinemia, insulin resistance, and MetS could lead to serum BAP elevation. In line with our previous studies (22-23), serum CRP was significantly

associated with alkaline phosphatase in the current study, while no association was observed with BAP ($P > 0.5$; data not shown). These observations further suggested that although total alkaline phosphatase and BAP are strongly correlated, the causes of elevation of total alkaline phosphatase and BAP could be different.

There have only been a few studies of BAP with insulin resistance and MetS. In agreement with a cross-sectional study carried out in 328 type 2 diabetes patients, we also observed no significant association between BAP and fasting plasma glucose and HbA1c (24). In a study of 54 healthy postmenopausal women, there was no correlation between BAP and any components of MetS (25), which could be due to the small sample size.

Serum alkaline phosphatase is routinely measured clinically, while BAP is not. Therefore, most epidemiological studies examined the association of serum alkaline phosphatase with mortality and cardiovascular diseases. Among 4,115 participants of the Cholesterol And Recurrent Events study, alkaline phosphatase was associated with all-cause mortality. This finding was subsequently replicated using NHANES III data, with significant association being observed between alkaline phosphatase and all-cause and cardiovascular mortality (10). Similar results are observed in patients

with maintenance hemodialysis (26) and acute stroke (27). In a retrospective study including 10,743 outpatients, serum alkaline phosphatase was robustly associated with mortality (including cardiovascular mortality) and cardiovascular related hospitalization (28). In addition to mortality, serum alkaline phosphatase has been reported to be associated with multiple cardiovascular diseases, such as stroke incidence (29), coronary artery calcification score (30), myocardial infarction, stent thrombosis, major adverse cardiac event, and coronary calcification (11). Our earlier study also showed that elevated alkaline phosphatase is associated with peripheral arterial disease (31). A few studies examined the role of BAP in mortality and cardiovascular diseases. In 130 osteoporotic patients, serum BAP was significantly higher in those with vascular calcification than those without (32). In a prospective study carried out in 800 dialysis patients, serum BAP was robustly associated with cardiovascular and non-cardiovascular mortality, and the effect size was much stronger than that of serum total alkaline phosphatase (33). Altogether, these studies suggested that alkaline phosphatase is an important predictor of mortality and cardiovascular events, while our current study provided evidence that BAP may be the link between insulin resistance and vascular calcification, cardiovascular diseases, or even mortality. Future studies on clinical outcomes should measure BAP in addition to total alkaline phosphatase (11).

BAP is known as a bone formation marker. Our previous study showed that genetic variation in *ALPL* gene was associated with hip geometry and bone mineral density in 1,513 Framingham Offspring Cohort participants (34). Therefore the associations observed in the current study could reflect difference in bone turnover. We then further adjusted for another bone turnover marker, urinary N-terminal telopeptides, in the logistic regression model (Table 2). Further adjustment for urinary N-terminal telopeptides attenuated the estimates but the associations remained statistically significant. These findings suggest that at least part of the association between glycemic traits and BAP is independent of bone turnover. Similarly, BAP may also be associated with growth. We then performed sensitivity analysis by excluding participants aged <25, and the results were essentially similar to the results before the exclusion (Supplementary Table 3). Future study is required to confirm our findings.

The association of MetS with osteoporosis is controversial, with MetS was reported to be associated with higher BMD (35) and lower BMD (36). In previous study, blood pressure was shown to be negatively associated with bone mineral density (36).

However, blood pressure was found to be positively associated with BAP in the current study. It should also be noted that, although BAP is mainly expressed in osteoblast, it is also expressed in other cells, such as B-cell (37) and vascular smooth muscle cells (38). Therefore, serum BAP could also be contributed by other cells,

although it is expected to be contributed predominantly by osteoblasts.

Population studies have shown that diabetes is preventable or even reversible. Therefore, identification of biomarkers may have the potential for early diagnosis of insulin resistance and hence prevention of diabetes (39-40). In this study, we showed that serum BAP concentration is associated with several metabolic traits, therefore BAP could potential be used as a biomarker. Future studies are required to examine whether addition of BAP as a predictive biomarker could enhance the performance of existing prevention strategies such as increased physical activity, healthy diet, and early treatment of hyperglycemia (39-40).

Our study has several strengths. The study population is large, multiethnic, nationally-representative, and well-characterized with data on multiple risk factors and confounders. The findings were shown to be robust in multiple statistical tests. Nevertheless, there are limitations. First, cross-sectional data cannot prove causality. Second, although the model in the current study showed a good fit in SEM, this may not be the only plausible model since the mechanisms underlying metabolic risk factors is complex. Future studies incorporating evidence from laboratory and human studies are required to investigate the mechanism in detail. Third, HOMA2 indices are

only surrogate markers, for example, HOMA2-IR is a surrogate marker of hepatic insulin resistance; and these indices are strongly correlated with insulin levels. It is unclear whether insulin resistance, insulin levels, or both leads to increased BAP. Future studies using euglycemic hyperinsulinemic and hyperglycemic clamp techniques are required to confirm our findings. Fourth, as aforementioned, the regulation of BAP is complex, therefore the results should be interpreted with caution.

In conclusion, our study provides a possible mechanism for the increased vascular calcification in subjects with increased insulin resistance such as MetS or type 2 diabetes. Future studies are required to examine whether improving insulin resistance and MetS could reduce the expression of BAP and the risk of vascular calcification and cardiovascular disease.

Conflict of interest

The authors reported no conflict of interest.

Acknowledgements

BMYC receives support from the Sun Chieh Yeh Heart Foundation. C.L. Cheung is supported by the Faculty Development Fund, Li Ka Shing Faculty of Medicine, The

References

1. **Shao JS, Cheng SL, Sadhu J, Towler DA** 2010 Inflammation and the osteogenic regulation of vascular calcification: a review and perspective. *Hypertension* 55:579-592
2. **Silva JA, Escobar A, Collins TJ, Ramee SR, White CJ** 1995 Unstable angina. A comparison of angioscopic findings between diabetic and nondiabetic patients. *Circulation* 92:1731-1736
3. **Marso SP, Mercado N, Maehara A, Weisz G, Mintz GS, McPherson J, Schiele F, Dudek D, Fahy M, Xu K, Lansky A, Templin B, Zhang Z, de Bruyne B, Serruys PW, Stone GW** 2012 Plaque composition and clinical outcomes in acute coronary syndrome patients with metabolic syndrome or diabetes. *JACC Cardiovasc Imaging* 5:S42-52
4. **Wong ND, Nelson JC, Granston T, Bertoni AG, Blumenthal RS, Carr JJ, Guerci A, Jacobs DR, Jr., Kronmal R, Liu K, Saad M, Selvin E, Tracy R, Detrano R** 2012 Metabolic syndrome, diabetes, and incidence and progression of coronary calcium: the Multiethnic Study of Atherosclerosis study. *JACC Cardiovasc Imaging* 5:358-366
5. **Johnson K, Polewski M, van Etten D, Terkeltaub R** 2005 Chondrogenesis mediated by PPI depletion promotes spontaneous aortic calcification in NPP1^{-/-} mice. *Arterioscler Thromb Vasc Biol* 25:686-691
6. **Hui M, Li SQ, Holmyard D, Cheng P** 1997 Stable transfection of nonosteogenic cell lines with tissue nonspecific alkaline phosphatase enhances mineral deposition both in the presence and absence of beta-glycerophosphate: possible role for alkaline phosphatase in pathological mineralization. *Calcif Tissue Int* 60:467-472
7. **Narisawa S, Harmey D, Yadav MC, O'Neill WC, Hoylaerts MF, Millan JL** 2007 Novel inhibitors of alkaline phosphatase suppress vascular smooth muscle cell calcification. *J Bone Miner Res* 22:1700-1710
8. **Ho AM, Johnson MD, Kingsley DM** 2000 Role of the mouse ank gene in control of tissue calcification and arthritis. *Science* 289:265-270
9. **Sage AP, Tintut Y, Demer LL** 2010 Regulatory mechanisms in vascular calcification. *Nat Rev Cardiol* 7:528-536
10. **Tonelli M, Curhan G, Pfeffer M, Sacks F, Thadhani R, Melamed ML,**

- Wiebe N, Muntner P** 2009 Relation between alkaline phosphatase, serum phosphate, and all-cause or cardiovascular mortality. *Circulation* 120:1784-1792
11. **Park JB, Kang DY, Yang HM, Cho HJ, Park KW, Lee HY, Kang HJ, Koo BK, Kim HS** 2013 Serum alkaline phosphatase is a predictor of mortality, myocardial infarction, or stent thrombosis after implantation of coronary drug-eluting stent. *Eur Heart J*
 12. **Bervoets AR, Spasovski GB, Behets GJ, Dams G, Polenakovic MH, Zafirovska K, Van Hoof VO, De Broe ME, D'Haese PC** 2003 Useful biochemical markers for diagnosing renal osteodystrophy in predialysis end-stage renal failure patients. *Am J Kidney Dis* 41:997-1007
 13. Centers for Disease Control and Prevention (CDC). National Center for Health Statistics (NCHS). National Health and Nutrition Examination Survey Data. Hyattsville, MD: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention, 1999-2004, http://www.cdc.gov/nchs/nhanes/nhanes_questionnaires.htm]. Accessed March 20, 2013.
 14. **Levy JC, Matthews DR, Hermans MP** 1998 Correct homeostasis model assessment (HOMA) evaluation uses the computer program. *Diabetes Care* 21:2191-2192
 15. **Bedogni G, Bellentani S, Miglioli L, Masutti F, Passalacqua M, Castiglione A, Tiribelli C** 2006 The Fatty Liver Index: a simple and accurate predictor of hepatic steatosis in the general population. *BMC Gastroenterol* 6:33
 16. **Lin LY, Kuo HK, Li HY, Hwang JJ, Lin JW** 2008 Confirming a biological pathway in the metabolic syndrome--insight from the NHANES 1999-2002. *Obesity (Silver Spring)* 16:2676-2681
 17. **Kline RB** Measurement models and confirmatory factor analysis. In: *Principles and Practice of Structural Equation Modeling*. 22005 ed. New York: Guilford Press; 133-145
 18. **Hu LT, Bentler PM** 1999 Cutoff Criteria for Fit Indexes in Covariance Structure Analysis: Conventional Criteria Versus New Alternatives. *Struct Equ Modeling* 6:1-55
 19. **Yang J, Zhang X, Wang W, Liu J** 2010 Insulin stimulates osteoblast proliferation and differentiation through ERK and PI3K in MG-63 cells. *Cell Biochem Funct* 28:334-341
 20. **Hie M, Iitsuka N, Otsuka T, Tsukamoto I** 2011 Insulin-dependent diabetes mellitus decreases osteoblastogenesis associated with the inhibition of Wnt signaling through increased expression of Sost and Dkk1 and inhibition of Akt

- activation. *Int J Mol Med* 28:455-462
21. **Zhen D, Chen Y, Tang X** 2010 Metformin reverses the deleterious effects of high glucose on osteoblast function. *J Diabetes Complications* 24:334-344
 22. **Webber M, Krishnan A, Thomas NG, Cheung BM** 2010 Association between serum alkaline phosphatase and C-reactive protein in the United States National Health and Nutrition Examination Survey 2005-2006. *Clin Chem Lab Med* 48:167-173
 23. **Cheung BM, Ong KL, Cheung RV, Wong LY, Wat NM, Tam S, Leung GM, Cheng CH, Woo J, Janus ED, Lau CP, Lam TH, Lam KS** 2008 Association between plasma alkaline phosphatase and C-reactive protein in Hong Kong Chinese. *Clin Chem Lab Med* 46:523-527
 24. **Kanazawa I, Yamaguchi T, Yamamoto M, Yamauchi M, Kurioka S, Yano S, Sugimoto T** 2009 Serum osteocalcin level is associated with glucose metabolism and atherosclerosis parameters in type 2 diabetes mellitus. *J Clin Endocrinol Metab* 94:45-49
 25. **Garcia-Martin A, Cortes-Berdonces M, Luque-Fernandez I, Rozas-Moreno P, Quesada-Charneco M, Munoz-Torres M** 2011 Osteocalcin as a marker of metabolic risk in healthy postmenopausal women. *Menopause* 18:537-541
 26. **Regidor DL, Kovesdy CP, Mehrotra R, Rambod M, Jing J, McAllister CJ, Van Wyck D, Kopple JD, Kalantar-Zadeh K** 2008 Serum alkaline phosphatase predicts mortality among maintenance hemodialysis patients. *J Am Soc Nephrol* 19:2193-2203
 27. **Ryu WS, Lee SH, Kim CK, Kim BJ, Yoon BW** 2010 Increased serum alkaline phosphatase as a predictor of long-term mortality after stroke. *Neurology* 75:1995-2002
 28. **Abramowitz M, Muntner P, Coco M, Southern W, Lotwin I, Hostetter TH, Melamed ML** 2010 Serum alkaline phosphatase and phosphate and risk of mortality and hospitalization. *Clin J Am Soc Nephrol* 5:1064-1071
 29. **Shimizu Y, Imano H, Ohira T, Kitamura A, Kiyama M, Okada T, Ishikawa Y, Shimamoto T, Yamagishi K, Tanigawa T, Iso H** 2012 Alkaline Phosphatase and Risk of Stroke Among Japanese: The Circulatory Risk in Communities Study (CIRCS). *J Stroke Cerebrovasc Dis*
 30. **Shantouf R, Kovesdy CP, Kim Y, Ahmadi N, Luna A, Luna C, Rambod M, Nissenson AR, Budoff MJ, Kalantar-Zadeh K** 2009 Association of serum alkaline phosphatase with coronary artery calcification in maintenance hemodialysis patients. *Clin J Am Soc Nephrol* 4:1106-1114
 31. **Cheung BM, Ong KL, Wong LY** 2009 Elevated serum alkaline phosphatase

- and peripheral arterial disease in the United States National Health and Nutrition Examination Survey 1999-2004. *Int J Cardiol* 135:156-161
32. **Iba K, Takada J, Yamashita T** 2004 The serum level of bone-specific alkaline phosphatase activity is associated with aortic calcification in osteoporosis patients. *J Bone Miner Metab* 22:594-596
 33. **Drechsler C, Verduijn M, Pilz S, Krediet RT, Dekker FW, Wanner C, Ketteler M, Boeschoten EW, Brandenburg V** 2011 Bone alkaline phosphatase and mortality in dialysis patients. *Clin J Am Soc Nephrol* 6:1752-1759
 34. **Cheung CL, Livshits G, Zhou Y, Meigs JB, McAteer JB, Florez JC, Cupples LA, Demissie S, Kiel DP, Karasik D** 2010 Hip geometry variation is associated with bone mineralization pathway gene variants: The Framingham Study. *J Bone Miner Res* 25:1564-1571
 35. **Hernandez JL, Olmos JM, Pariente E, Martinez J, Valero C, Garcia-Velasco P, Nan D, Llorca J, Gonzalez-Macias J** 2010 Metabolic syndrome and bone metabolism: the Camargo Cohort study. *Menopause* 17:955-961
 36. **Jeon YK, Lee JG, Kim SS, Kim BH, Kim SJ, Kim YK, Kim IJ** 2011 Association between bone mineral density and metabolic syndrome in pre- and postmenopausal women. *Endocr J* 58:87-93
 37. **Hossain A, Jung LK** 2008 Expression of bone specific alkaline phosphatase on human B cells. *Cell Immunol* 253:66-70
 38. **Shioi A, Katagi M, Okuno Y, Mori K, Jono S, Koyama H, Nishizawa Y** 2002 Induction of bone-type alkaline phosphatase in human vascular smooth muscle cells: roles of tumor necrosis factor-alpha and oncostatin M derived from macrophages. *Circ Res* 91:9-16
 39. **Lindstrom J, Neumann A, Sheppard KE, Gilis-Januszewska A, Greaves CJ, Handke U, Pajunen P, Puhl S, Polonen A, Rissanen A, Roden M, Stemper T, Telle-Hjellset V, Tuomilehto J, Velickiene D, Schwarz PE, Acosta T, Adler M, AlKerwi A, Barengo N, Barengo R, Boavida JM, Charlesworth K, Christov V, Claussen B, Cos X, Cosson E, Deceukelier S, Dimitrijevic-Sreckovic V, Djordjevic P, Evans P, Felton AM, Fischer M, Gabriel-Sanchez R, Goldfracht M, Gomez JL, Hall M, Hauner H, Herbst J, Hermanns N, Herrebrugh L, Huber C, Huhmer U, Huttunen J, Jotic A, Kamenov Z, Karadeniz S, Katsilambros N, Khalangot M, Kissimova-Skarbek K, Kohler D, Kopp V, Kronsbein P, Kulzer B, Kyne-Grzebalski D, Lalic K, Lalic N, Landgraf R, Lee-Barkey YH, Liatis S, Makrilakis K, McIntosh C, McKee M, Mesquita AC, Misina D, Muylle**

F, Paiva AC, Paulweber B, Peltonen M, Perrenoud L, Pfeiffer A, Raposo F, Reinehr T, Robinson C, Rothe U, Saaristo T, Scholl J, Spiers S, Stratmann B, Szendroedi J, Szybinski Z, Tankova T, Terry G, Tolks D, Toti F, Undeutsch A, Valadas C, Valensi P, Vermunt P, Weiss R, Wens J, Yilmaz T 2010 Take action to prevent diabetes--the IMAGE toolkit for the prevention of type 2 diabetes in Europe. *Horm Metab Res* 42 Suppl 1:S37-55

40. **Schwarz PE, Greaves CJ, Lindstrom J, Yates T, Davies MJ** 2012 Nonpharmacological interventions for the prevention of type 2 diabetes mellitus. *Nat Rev Endocrinol* 8:363-373

Table 1. Characteristics of the study population (NHANES 1999-2004).

	Total (n=3,773)		Men (n=1,932)		Women (n=1,841)		P-value
	%	(95% CI)	%	(95% CI)	%	(95% CI)	
%			49.6	(48.4-50.8)	50.4	(49.2-51.6)	NA
Smoking (%)							
never	50.7	(47.9-53.6)	44.6	(41.3-48)	56.7	(53.2-60.1)	<0.001
former	23.3	(21.3-25.4)	26.3	(24.1-28.6)	20.3	(17.7-23.2)	<0.001
current	26	(23.8-28.4)	29	(26.2-32.1)	23	(20.7-25.6)	<0.001
Active drinker (%)	75	(71.2-78.4)	84.3	(80.5-87.6)	65.8	(61-70.3)	<0.001
Race/ethnicity (%)							
Mexican American	12.7	(9.8-16.4)	13.6	(10.5-17.4)	11.8	(8.8-15.7)	0.101
Non-Hispanic White	73.6	(69.9-77)	72.5	(68.5-76.2)	74.7	(71-78)	0.04
Non-Hispanic Black	10	(8.1-12.3)	9.7	(7.8-12.1)	10.2	(8.1-12.8)	0.516
Others	3.7	(2.8-5)	4.2	(3-5.9)	3.3	(2.1-5)	0.324
Education (%)							
< High school	18.5	(16.9-20.2)	18.8	(16.5-21.4)	18.2	(16.3-20.2)	0.667
High school	26.3	(23.9-28.9)	27.6	(24.8-30.5)	25.1	(22.2-28.3)	0.147
> High school	55.1	(52.2-58)	53.6	(50.2-56.9)	56.7	(53-60.3)	0.12
Had moderate or vigorous activity (%)	65.9	(63.5-68.2)	67.5	(64.6-70.3)	64.2	(61.4-66.9)	0.038
Age (yr)	39.94	(39.03-40.88)	38.84	(37.93-39.78)	41.05	(39.95-42.18)	<0.001
BMI (m/kg ²)	26.99	(26.75-27.23)	27.09	(26.82-27.36)	26.90	(26.53-27.26)	0.357
Waist circumference (cm)	93.35	(92.73-93.99)	96.92	(96.12-97.75)	89.97	(89.1-90.87)	<0.001
Bone-specific Alkaline phosphatase(ug/L)	13.76	(13.45-14.08)	14.96	(14.57-15.35)	12.68	(12.3-13.07)	<0.001
ALP (U/L)	67.05	(65.89-68.22)	70.36	(68.8-71.94)	63.94	(62.75-65.16)	<0.001
ALT (U/L)	22.50	(22.15-22.86)	27.25	(26.52-27.98)	18.65	(18.25-19.05)	<0.001
AST (U/L)	22.86	(22.51-23.2)	25.10	(24.58-25.63)	20.84	(20.46-21.23)	<0.001
GGT (U/L)	21.67	(21.09-22.26)	26.95	(25.98-27.96)	17.48	(16.93-18.06)	<0.001
C-reactive protein(mg/dL)	0.18	(0.17-0.19)	0.15	(0.14-0.16)	0.22	(0.21-0.24)	<0.001
Total cholesterol (mg/dL)	196.47	(194.4-198.52)	196.34	(193.55-199.11)	196.61	(194.4-198.79)	0.847
Triglycerides (mg/dL)	115.50	(112.05-119.07)	124.17	(118.77-129.81)	107.60	(104.35-110.92)	<0.001
HDL-cholesterol (mg/dL)	49.75	(49-50.5)	44.99	(44.1-45.89)	54.92	(53.91-55.92)	<0.001
Mean arterial pressure (mm/Hg)	87.76	(87.22-88.31)	89.23	(88.63-89.85)	86.34	(85.59-87.08)	<0.001
Glycohemoglobin (%)	5.23	(5.21-5.26)	5.25	(5.22-5.28)	5.22	(5.19-5.24)	0.006
Glucose, plasma (mg/dL)	94.54	(93.99-95.1)	96.38	(95.76-97.03)	92.77	(92.15-93.39)	<0.001
Insulin (uU/mL)	8.87	(8.59-9.17)	9.21	(8.84-9.59)	8.55	(8.24-8.88)	0.001

HOMA2-IR	1.00	(0.97-1.04)	1.05	(1.01-1.09)	0.96	(0.93-1)	<0.001
HOMA2-IS (%)	99.52	(96.32-102.83)	95.50	(91.69-99.47)	103.63	(99.79-107.65)	<0.001
HOMA2-B (%)	86.28	(84.63-87.98)	85.15	(83.2-87.16)	87.42	(85.39-89.47)	0.049

Data are geometric means (95% CI). All values presented are weighted to represent the U.S. civilian population, 1999-2004. P-value represents differences in means or proportions, using ANOVA or χ^2 test.

Table 2. Association between glyceemic traits and alkaline phosphatase (bone-specific or total) in NHANES 1999-2004

Glyceemic traits		Model 1 (N=3,773)			Model 2 (N=3,773)			Model 3 (N=3,009)*		
		Estimate	95% CI	P-value	Estimate	95% CI	P-value	Estimate	95% CI	P-value
Bone-specific alkaline phosphatase	HOMA-IR	0.109	(0.079-0.138)	<0.001	0.068	(0.038-0.097)	<0.001	0.044	(0.008-0.08)	0.018
	HOMA-IS	-0.107	(-0.137--0.077)	<0.001	-0.065	(-0.095--0.035)	<0.001	-0.041	(-0.079--0.004)	0.031
	HOMA-b	0.134	(0.088-0.18)	<0.001	0.081	(0.038-0.125)	<0.001	0.054	(0.009-0.1)	0.02
	Fasting glucose (mg/dL)	0.202	(0.038-0.366)	0.017	0.072	(-0.082-0.227)	0.351	0.021	(-0.167-0.208)	0.823
	Insulin (uU/mL)	0.107	(0.077-0.136)	<0.001	0.065	(0.035-0.095)	<0.001	0.042	(0.005-0.078)	0.026
	HbA1C (%)	0.22	(-0.033-0.473)	0.087	0.13	(0.085-0.346)	0.229	0.109	(-0.142-0.36)	0.382
Total alkaline phosphatase	HOMA-IR	0.072	(0.048-0.096)	<0.001	0.022	(-0.002-0.045)	0.071	-0.017	(-0.047-0.013)	0.259
	HOMA-IS	-0.07	(-0.095--0.046)	<0.001	-0.02	(-0.044-0.004)	0.098	0.02	(-0.011-0.051)	0.2
	HOMA-b	0.092	(0.055-0.13)	<0.001	0.032	(-0.002-0.065)	0.065	-0.014	(-0.05-0.023)	0.457
	Fasting glucose (mg/dL)	0.089	(-0.05-0.228)	0.202	-0.031	(-0.166-0.103)	0.643	-0.077	(-0.242-0.089)	0.35
	Insulin (uU/mL)	0.07	(0.046-0.094)	<0.001	0.02	(-0.004-0.043)	0.095	-0.019	(-0.049-0.011)	0.199
	HbA1C (%)	0.122	(-0.058-0.302)	0.179	0.004	(-0.157-0.164)	0.965	0.008	(-0.169-0.185)	0.93

Model 1: Adjusted for age, race/ethnicity, education, smoking, drinking, physical activity, and waist circumference; Model 2: Further adjusted for AST, ALT, GGT, total cholesterol, triglycerides, HDL-cholesterol, mean arterial pressure, and CRP; **Model 3: Further adjusted for fatty liver index and urinary N-terminal telopeptides, while waist circumference was removed from the model due to high correlation with fatty liver index.**

* Urinary N-telopeptides data was only available in NHANES 1999-2002.

Table 3. Association between components of MetS and alkaline phosphatase (bone-specific or total) in NHANES 1999-2004

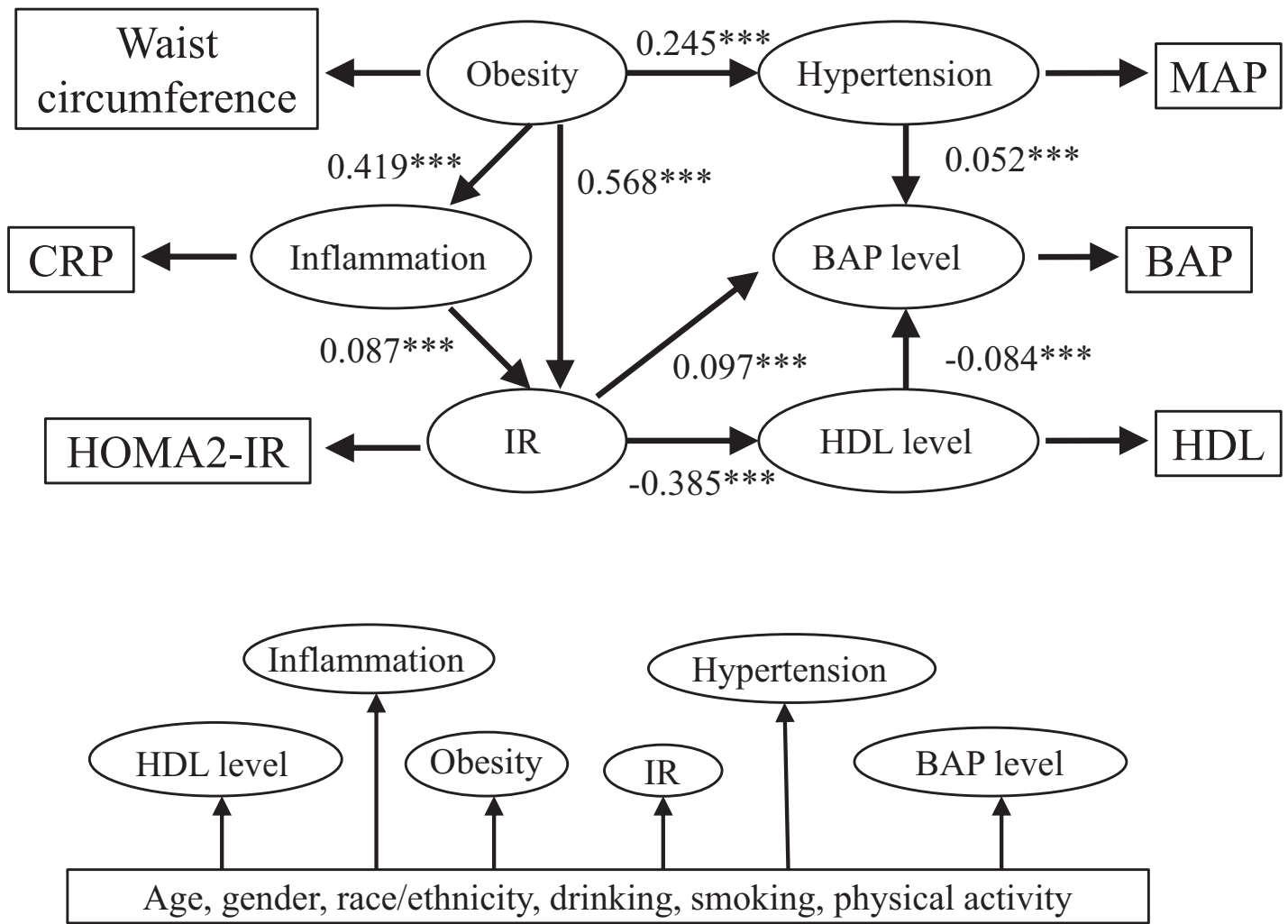
Components of MetS	Bone-specific alkaline phosphatase				Total alkaline phosphatase			
	Estimate	95% CI		P-value	Estimate	95% CI		P-value
		Lower limit	Upper limit			Lower limit	Upper limit	
Waist Circumferences (cm)	-0.122	-0.244	0.001	0.052	-0.049	-0.153	0.056	0.352
Mean Arterial Pressure (mm/Hg)	0.15	0.041	0.259	0.008	0.119	0.045	0.193	0.002
HDL-cholesterol (mg/dL)	-0.209	-0.272	-0.147	<0.001	-0.167	-0.217	-0.117	<0.001
Triglyceride (mg/dL)	-0.041	-0.086	0.003	0.069	-0.025	-0.062	0.013	0.191

Components of MetS simultaneously included in the model with further adjustment of age, race/ethnicity, education, smoking, drinking,

physical activity, AST, ALT, GGT, total cholesterol, HOMA-IR, and CRP.

Figure 1. Pre-defined structural equation model outlining relationships between insulin resistance, components of MetS, and BAP.

Oval circle indicates latent variable that is not measured directly. Standardized regression coefficients are at the base of each arrow. *** $P \leq 0.001$. HDL level, inflammation, obesity, IR, hypertension, and **BAP level** were adjusted for age, gender, race/ethnicity, drinking, smoking, and physical activity in the structural equation model.



Supplemental Material

[Click here to download Supplemental Material: 2013Manuscript_JCEM2_SI.docx](#)