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Peiris et al.

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C12N 7/04 (2006.01)
C12Q 1/70 (2006.01)

U.S. Cl. 424/204.1; 435/235.1; 435/236; 435/306; 435/5; 536/23.72

ABSTRACT
The present invention relates to an isolated novel virus causing Severe Acute Respiratory Syndrome (SARS) in humans ("hSARS virus"). The hSARS virus is identified to be morphologically and phylogenetically similar to known members of Coronaviridae. The present invention provides the complete genomic sequence of the hSARS virus. Furthermore, the invention provides the nucleic acids and peptides encoded by and/or derived from the hSARS virus and their use in diagnostic methods and therapeutic methods, including vaccines. In addition, the invention provides chimeric or recombinant viruses encoded by said nucleotide sequences and antibodies immunospecific to the polypeptides encoded by the nucleotide sequences.
FIG. 1
Murine hepatitis virus strain ML-11
Murine hepatitis virus
Murine hepatitis virus strain 2
Murine hepatitis virus
Murine hepatitis virus (strain JHM)
Bovine coronavirus
Bovine coronavirus
SARS virus, Hong Kong isolate
Avian infectious bronchitis virus
Avian infectious bronchitis virus (strain Beaudette CK)
Transmissible gastroenteritis virus
Human coronavirus 29E
Porcine epidemic diarrhea virus

Group | Accession number
--- | ---
2 | AAF68920
2 | AAF69332
2 | AAF19384
2 | NP_068668
2 | VFIHJH
2 | AAK83365
2 | AAL40397
3 | NP_066134
3 | CAC39112
1 | NP_058422
1 | NP_073549
1 | NP_598309

FIG. 6
FIG. 8
FIG. 8 Con't
c aga acc atg cct aac atg ctt agg ata atg gcc tct ctt gtt ctt gct 49
Arg Thr Met Pro Asn Met Leu Arg Ile Met Ala Ser Leu Val Leu Ala
1 5 10 15

cgc aaa cat aac act tgc tgt aac tta cca cac cgt ttc tac agg tta 97
Arg Lys His Thr Cys Cys Asn Leu Ser His Arg Phe Tyr Arg Leu
20 25 30

gct aac gag tgt ggc caa gta tta agt gag atg tgt gtc tgt ggg ggc 145
Ala Asn Glu Cys Ala Gin Val Leu Ser Glu Met Val Met Cys Gly Gly
35 40 45

tca cta tat gtt aaa cca ggt gga aca tca tcc ggt gat gct gca aca act 193
Ser Leu Tyr Val Lys Pro Gly Thr Ser Ser Gly Asp Ala Thr Thr
50 55 60

gct tat gct aat agt gtc ttt aac att tgt cca ggt gtt aca gcc aat 241
Ala Tyr Ala Asn Ser Val Phe Asn Ile Cys Gin Val Thr Ala Asn
65 70 75 80

gta aat gca ctt ctt tca act gat gtt aat aag ata gct gac aag tat 289
Val Asn Ala Leu Leu Ser Thr Asp Gly Asn Lys Ile Ala Asp Lys Tyr
85 90 95

gtc ggc aat cta cca cac agg ctc tat ggt tgt ctc tat aga aat ggg 337
Val Arg Asn Leu Gin His Arg Leu Tyr Gly Cys Leu Tyr Arg Arg
100 105 110

gat gtt gat cat gaa ttc gtg gat gga gtt ttc tac gct tac ctc gct aca 385
Asp Val Asp His Glu Phe Val Asp Glu Phe Tyr Ala Tyr Leu Arg Lys
115 120 125

cat ttc tcc atg atg att ctt tct gat gat gcc gtt gtt tgt gtt tat aac 433
His Phe Ser Met Met Ile Leu Ser Asp Ala Val Val Cys Tyr Asn
130 135 140

FIG. 9
FIG. 10 Con't
<table>
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<tr>
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</tr>
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<tr>
<td>10081 gatgcacacag tatactgcgcc aagacatgtg aatggagccag cagaagacact gcttaacctt</td>
<td>10141 aactatgaag atctgctcatc tcgaacaaact aacatagtct tctctgttgtc gagctgcaaat</td>
<td>10201 gcttaaatctt gtattatggct cacatcact tagt ttaaatagtgctt</td>
<td>10261 actctcaacc tcaagacacc caagttaaaa tgtctgaatc ttcacacctg tcaacacttt</td>
</tr>
</tbody>
</table>

**FIG. 10 Con’t**
|--------------------------------|--------------|----------------|-------------------|

13441 cagggcactag tactgatgctc gtcctacagg ctttttgatat ttacaagcga aaaaaggtcctg
13501 gttttgcgaa gttcctaaaa acataaggct gtcctgcctca ggaggaaggt gaggagaagga
13561 attatattag cctctacttt cttgataaag gcagactaat gctcaactac caacatgaag
13621 agactattta ccactatgctt tcagctcttc gcgtctcag ttccttcgcgtcgattcagtt
13681 ttggctgtatt tctctcacta tgggtcagtg atgtattatt caataaagaag gattgttagt
13741 tggctgtcatt atctccttcag ccctgttgat atcccttcag ttcctgtctgctgtagcagc
13801 aatcactcgt cgcatacttc tgggtataa ggtacatcta aaacatccag aacccagcagtt
13861 acctgatcag gattcattctt ttccttcttc ctgatccttg gctgtaataaa
13921 cccctctatt tttcctcttc cctgtagtct gctctctctt ggtttctttccagc
14081 tcccttcttttt cacccaaattt cctcctctcc gtttctgatg cggctgagctt cctcagtgttctc
14141 cccctctcttcc gttttcttttt tttttttttt ttttttttttt ttttttttttt ttttttttttt
14201 ccctcccttttt ttttttttttt ttttttttttt ttttttttttt ttttttttttt ttttttttttt
14261 aatcgttattg aatcgttattg aatcgttattg aatcgttattg aatcgttattg aatcgttattg
14321 ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt ccctctctct
14381 cgcctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14441 ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt ccctctctct
14501 atctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14561 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14621 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14681 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14741 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14801 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14861 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14921 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
14981 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15041 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15101 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15161 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15221 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15281 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15341 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15401 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15461 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15521 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15581 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15641 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15701 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15761 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15821 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15881 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
15941 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16001 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16061 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16121 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16181 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16241 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16301 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16361 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16421 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16481 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16541 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16601 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16661 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt
16721 gctctctctct ctgctcattt gttctgctctt ccctctctct ctgctcattt gttctgctctt

**FIG. 10 Con't**
FIG. 1O Con't
FIG. 10 Con't
FIG. 10 Con't
FIG. 11 Con't
FIG. 11 Con't
ATG GTG ATT GAG TTA AAT GCT GTA AAC TAC ACA CTT GAA AAT GGT GTG TGG TGT GAG CAG GTGCA
M V I V N A A N I H L K H G G V A G A
W * L * M L L T T Y T * N M V V V * Q V H
G D C K C * H T P E T T E W W C S B C T
CTCA ACA AGG GCA ACCA ATG TGG TCC ATG AAA AAG GAG NTG ATG ATT C ATTA CAG CT AAT
L N K A T N G A M Q K E S D D Y I K L N
ST R Q P M V P C K R R V M I T L S * M
Q G G N Q W C H A K G E * L H * A K W
GGCC CCT CTAC A GTG AGG AGG GC TGT T G TCT GGAC AT AA AT CT TGG CT AAG AGA GTT
G P L T V G G S C L S G S N H N L A K K C
AL L O * E G L V C F L D I I L L R S V
PS Y S R R V L P A F W T * S C * E V S
CTG CA ATG TT TGG ACC AT CTA AAT GCAG GT TGG AGC AT CAC GC TCT CT TGA GGCA GCA
L H V G P N L N A G E D I Q L L K A A
C M L L D L T * M Q V R T S S F L R Q H
A C C W T * P K C R * G H P A S * G S I
TAT GA AA TT TCC AAT TC GAG GAC AT CTT ACT TT GC AAT TGT G TT GAC GC A A TTT P
Y E N F N S Q D I L A P L L S A G I F
MK I S I H R T S Y L H H C C Q Q A Y L
* K F Q F T G H L T C T I V S R H I L
GGCT C T T A A C C A C T C T A C G T C T T A A C A G T G T G T G C A G C A C G T G T G
G A K P L Q S L Q V C Q V T Q V T Q V Y
V L N H P S L Y K C A C R R F V H R F I
C * T T S V F T S V R A D G S Y T G L Y
ATG CAG CTG CAC ACA AAC GCT CT T T T T A T G C C G A C AG T T A T C G T C T C T G G
I A V N D K A L Y E Q V V M D Y L D N L
L Q S M T K L F M S R L S W I L I T *
C S Q * Q S S L * A G C H G L S * P E
K P R V E A P K Q E E P P N T E D S K T
S L E W K H L N K R S H Q T O K I P K L
A * S G T * T R G A T K H R R F Q N *
E E K S V V K P V D V K E K I K A C T
R R N L S Y R S L S M * S Q K L R P A L
G E I C R T E A C R C E A R N * G L H *
D E V T T T L E E T K F L T N K L L L F
M R L P Q H W K K L S F L P I S Y S C L
* G Y H N T G R N * V S Y Q * V T L V C
A D I N G H Y H D S Q N M L R G E D M
L I N G V S F T M I L R T C L E V K I C
* Y Q W * A L P * F S E H A R R * R Y V
T C T T C C T T C T C C T G A A A G A G T G A C C T T A C T G T G A T G A C T C T C T G T G A T A C T
S F L E K D A P Y M V G D V I T S G D I
L S L R R M H L T W * V M L S L V V I S
F P * E G C T L H G R * C Y H * W * Y H
T C V V I P S K K A G G T E T E M L S R A
L V L * Y P P K R L V A L L R C S Q E L
L C C N T L Q K G W W H Y * D A L K S F
L K K V P V D E Y I T T Y P Q G Q C A G
* R K C Q L M S I * P R T L D K D V L V
E E S A S * * V Y N H V F W T R M C W L

FIG. 11 Con’t
FIG. 11 Con’t
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FIG. 11 Con't
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FIG. 11 Con't
10921 - GTTAAGGGCCACTCATCATTGGATGCTTTAATCTTTCTTTGACAACTGATTGATTCCCTTGTT - 10980
-VKGTTHWMLLFTFLTSLLILV
-LRALIGFSLHSHYFLF
-*GSSSLDAFNFLDIDSTCS

10981 - CAAAGTGACAGGTTGAGCTGACCTCTTCTTTTTGTACAGATGTGCTTTGACCTTACATCCT - 11040
-QSTQWSSFYVYENAFLPFT
-KHSHGHCFSLFTSRMGLSCHL
-KYTVTVTFLCRLRECFLAIYS

11041 - CTGTTGTATATATGGCATATGGCTATGTTGCTATGTTGCTATGTTAGCATAACGGCATTC - 11100
-LGIMATIAACAMLGLVHKKHAP
-LVWLQLLHVLCLLSSLISTSHS
-WYGLQCCCMCYAAC*AARIL

11101 - TTGTGCTTGGTTCATCTTCATCTGCAATCAGGTGCTATCTTTATATATGTCCTTCATG - 11160
-LCLFLPLPSLATVAYFPNMVYM
-CACFCYLLQLLLTTLIWSCT
-VLVSVTFCSCNSSLYGLHLA

11161 - CCTGCATCGCTGGTATGCTACGATACACGGCATTGAATGGCTGACCTGACTGCTTGA - 11220
-PASWVMXRMIWTWLELADTSL
-LLAGCVSHGLNNWLTTLACL
-CLCGDAYHAIHGINHLYW

11221 - GGTATTAGGTGTTAAGAGTGTGTATAGCTGCTATGTATATCATATACGCTTCTTCATG - 11280
-GYRLRDCMVKYASALVRLLILM
-VLRLRVISSLMLQLFCLFS
-LA*GLCYVCFSSFSAFAYSHD

11281 - ACACATCGTGCACTGCTTTATGCTGGTACGATGCTGACATGGTGAATGCTCTATTT - 11340
-TARTYVDDAARRVWTLMNV
-QPASLMMFLLLDLLVFHMSL
-SSSHCCACCTC[CLDTDECHY

11341 - ACACATTGTTAAAGAGGCTACATAGCTGCTATTACTATACAGCATATAGCTATTGAGGC - 11400
-TLVYKVYGGYGNALDQAISMAW
-HLFTKSTMVMLIKLFPCGP
-TCLQSLLLWCFRSSYFHVGL

11401 - TTATAGATTCTCTCTACATTCTACTATGGGCTGATGCTATCATATCTGCTTTAGCT - 11460
-LVISVTSSNYGSSVVTITMLA
-*FLPLPTLTVSLRSLSCFL
-SYFCNLFLFWRCYDYHVFS

11461 - AGAGCTATAGGGTTGTGTTGAGTATTTACCCATGATATTATTTATACGGGCAACACC - 11520
-RAIVFVFCVEYYPLYLLFITGNT
-ECLCLCVLSLITHCRYLLLLATP
-SYSVCVCVVLPIIVYYQH

11521 - TTACAGGCTATCATCTGCTTTTATATTGCTGCTTTGCTGCTTACTTTTGCC - 11580
-LQCMYLVYCVFLGMYCYYCGYFG
-YSVSCLFIYVSAILVAARAL
-TVYHACLLFLRLILLLLP

11581 - CTTTCTCTGTTACTCTACAGGTTACTTCAGCTCTCTCTTCTTCTATGACTACTTTGTC - 11640
-LFCNLNRYFRLTGLSVYDLYLV
-FSVYSTTSGGLLLVFMRTWS
-FLFTQPLLAQGAYSWCLLLLLG

11641 - TCTACCAAGAATTATGATATAGACTCTCAAGGAGCTTTTTGACTTTAAGAGTATATT - 11701
-STQEFYHMSQGLPFPKSSI
-LHKNLGI*TPRFGCLRLRVL
-YYTRIVYELPAGAS*E*IY

11701 - CATGGTTTCACGGTCATATGATTGATGGTTAGATGAATACCGTATGACTGATTT - 11760
-DAFKLNIKLGLIGGKPCIKV
-MLSSLTLTSCWVLLEVNHVSR
-CFQA*HVVGYWR*TMYQGC

FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
15121 - GRRGCATCTGTTGAAATATGGAAGACCCAGAACTGCTTATAGGCTGCTGGCATAATATTTTATAAGGCTTGRGTGCCTGTGGCACATAATATTTTA
- ELLWLMELQASFTVAGIC*K
- SYCGNWKNQVLRWL*A*YVKN
- ATTVVIGTSKFGYGGWHNLMLK

15181 - CTGGTTACGATGATGAAATTGAGTCCACTCCGATGTTGGAGTATCCGCAACATGCA
- LFTVM*KLHTLWVQNNVT
- CLQ*CRNSTPYGLGLGSKM*Q
- VYSVDETPTHLMGWDYPKCDR

15241 - GGACCAATCGTACTCAAGCTTATGGTACGGAAGACCCAGAACTGCTTATAGGCTGCTGGCATAATATTTTATAAGGCTTGRGTGCCTGTGGCACATAATATTTTA
- EPCLTCLG*WPLLFLLANIT
- SHA*H*ADNGLSCSCSQT*H
- ATTAYMLRIMASLVLRLKHTN

15301 - CTGGCTCTAACTTATCAGACCCAGCTTCTGATGATGAAATTGAGTCCACTCCGATGTTGGAGTATCCGCAACATGCA
- LAVTYHTVSTG*LTSVRKY*
- LLL*LITPFLQVS*RVCASIK
- CCNLSHRFYRLANECAQVLS

15361 - GTGAGATGGCCTACGAGGGCGCCTACATATATGTAAACAGAATGAGGCTGACTCAGCGGCAAGT
- VRSWCVAASHAHHLMNLQVEEHHPV
- D*GHVWRLTIC*TRWNIIIR*
- EMMVCSCGLSYWKPPTGSSGD

15421 - ATGGCTCAATCGCTTATGCGTAACATATATGAGGCTGACTCAGCGGCAAGT
- MLQLLMILMIVSTLFKLLQMPM
- CYNCILC*CL*HLSSCYSQC
- ATTAYMLRIMASLVLRLKHTN

15481 - TAAATGCACTCTCTCTCAGATGTTATGATGAAATTGAGTCCACTGCTCTATAGGCTGCTGGCATAATATTTTATAAGGCTTGRGTGCCTGTGGCACATAATATTTTA
- M*HFFQQLMVR*LTSMSAIY
- KCTSFS*NW**DS*QVCPQST
- NALLSTDGNKIAZDKYVRNLQ

15541 - AACACAGGCTACGACGGAAGAGCCTTATGATGAAATTGAGTCCACTGCTCTATAGGCTGCTGGCATAATATTTTATAAGGCTTGRGTGCCTGTGGCACATAATATTTTA
- NTGSMMSVSIEIGMGLIMNSW
- TQAL*VSLS*K*GC*S*IRG*
- HRLYESCLYRNRDVHDHFEFVDE

15601 - AGTGGTTACGATGATGAAATTGAGTCCACTCCGATGTTGGAGTATCCGCAACATGCA
- SFTLCVNIISP*FFLMHML
- VLRLPA*TFPHDDSDS*ACRC
- FLYAYLKHKFSMMLILDSDADV

15661 - GTGGCTTATACGAGACCTACACGCTGCAAGGACCTTATTGACTGACTGACTCAGACATATATTGAGGCTGACTCAGCGGCAAGT
- CNTVTMLRLKV**LARLTRLR
- VLQ*BLCGSFSS*H*ELG
- CYNSSNAAAGQLGVASIKNFKA

15721 - CAGTGTCTTATCTGTCAATATAGTGGCTTATGCTGCTGAGGCAAAATGTGAGATGACTCAGCGGCAAGT
- QFFIIKIMCSCSLRQNVNGRL
- SSLLS*K*CVDHV*KGMLOD*D*
- VLYYQNNVFMSAEAKCWETFETD

15781 - ACCTTACCTAAAGGACCTACACGCTTATTGACTGACTCAGACATATATTGAGGCTGACTCAGCGGCAAGT
- TLKDLTNTFAHSIC*C*LNKE
- PY*RTPSRIILTTAYNAS*TRR
- LTKGPHEFCSOQHTMLVQKGD

15841 - ATGGATTAACGATCCTGCTCTACCCGATCTCATCGACTAGATATATTAGGCGGCAAGCCTGTGTTGAGGCTGACTCAGCGGCAAGT
- MICTCLTQTIHQY*EAQAVL
- LRVAPALPSKINNIRRRRF
- DTVLYLPYDPERSIRILGAGCFV

15901 - TCGATGATAGTGGCTTAAACAGAGATGACCATACTTATGATGAAATTGAGTCCACTGCTCTATAGGCTGCTGGCATAATATTTTATAAGGCTTGRGTGCCTGTGGCACATAATATTTTA
- SMILLSKQMVMHL*LKGSCHWL
- R*YCQRWNYTYD*KVRVTGY
- DDIKVDKVDLTMIERFVSLAI

FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con't
FIG. 11 Con’t
26881 - GACCTGCAAAAGAGATCATCTGTGGCCTACATCGAAGCGCTTCTTATTGAAATATAGA - 26940
- DLPKIEITVATSRTLSSYKLEG
- TCKKRSLWHLHERFILTN*E
- FAKRDHCAYSITNAFLQIERS
26941 - CGGTCGACGGGCTGATGACACTGACGATCGTCTTCGTCCCATCGAATCGGAGATGAGATGATGTATGAT - 27000
- ASQVRGFTDSGFAYNRYRI
- RRSVEAIIQVLLHTTATVLE
- VAAACRHR*FFRCECCIQQLPYPW
27001 - AACATAAAATTTAAATACAGACCAACCCCGGATAGAGACAAATATGTCTCTCTACACAG - 27060
- NYKLTNDHRAGSNNDNIALLLVQ
- TINRIQPHTPVATITLLCYS
- L*IKYRPRR*QRQYCFASTV
27061 - TAAAGCTGCAACAGATGGTTACCTCACTTGTGCTTCTCGCTACGTAAAATGCAGAGATGAGATGATGT - 27120
- VTTVSSCLPGYNRSRDID
- KQMQFHLDVDQVTIAEILI
- SDRNCFILLTSRLOQ*QRY*L
27121 - TATCATATTAGAGCAGACCCAGCTTCCAGATGTTGAAATATCTGACGATGAGATGAGATGAGATGAGATGAG - 27180
- YHYEDFQDYLESRNYKNF
- IIMRTFRIAIWNLDVIISSI
- SL*GLSGLLFGILTL**VQ*
27181 - AGTGGAGGAGATTTATATTGGATGCTTATACAGAGGAAATATATCACGGAGGAGATGAGATGAGATGAGATGAGATGAG - 27240
- SETIIASNEELELFQVR**R
- VQLFKPLTCKKNYESEDITORED
- *DNYLSL*LRRIRSIMKKN
27241 - ACCTAAGGAGATTTATATTGGATGCTTATACAGAGGAAATATATCACGGAGGAGATGAGATGAGATGAGATGAGATGAG - 27300
- TGYVRLISKIRT*KLFSSSH*
- PMLDYPNEHENYSLPDID
- LWS*IHKTNNMKILFLTLI
27301 - TGTCATATTACCTTCGGCAGATGCTTATACAGAGGAAATATATCACGGAGGAGATGAGATGAGATGAGATGAGATGAG - 27360
- LYLHLASITYTIRSVLEVRLY
- CIYILRAISLSGVC*RYDC
- VFTSCELYHYQECVRGTTVL
27361 - TACTAAAAGAGCCTGCGCAGGAGACATACAGGGGCGTACACACATCTCTAGCTGCTGAGAGATGAGATGAGATGAGATGAGATGAG - 27420
- Y*KNLAHEHTRAIHHFTLL
- TKTLPINHERQFETSPPSC
- LKEPFCPSGTYEWSNPFHPLA
27421 - CGGACCATAAATTTTGACACTTATGCTGTGACACACAGCTCTTCGCTTTCTGTTGCTTCAGAG - 27480
- LTINLHL*LLLALAAHTLLLVLTV
- *Q*ICTIONIHHTLCFCLC*R
- DNKFALTCTSTHFAFACADG
27481 - GACTCTGACATACATCATCGCTGTCAGAAGCTAAGCTCTCCTACATCTCTCTACATCTTCTTCGCTGCTCTGTATAT - 27540
- VLDIPISCVQDQHFQHNFSSD
- IYSTYLSAAACRISKSTFKTFTFHQST
- TRHYQLRARSVPFLKFLIRQ
27541 - AAGGAGGAGCTTACCAAGAAGAGATACGTCCTACTCCACACTCTCTACATCTCTCTACATCTTCTTCGCTGCTCTGTATAT - 27600
- KRRNFNKSSTRHFSLLLVL*Y
- RGGSTRALLATFSSHCCCSSI
- EEYQELYSLPSLLFLLIVAALLVF
27601 - TTTAATATCTTTGCTCAGCTTAATAGAGAAGAAGAGAATGATGACTGTCCTTTTATTATGAGA - 27660
- FYFASFLREQRNE*AHHF*N
- FNTLLLHH*ERDOKMELTLDLILFIKREKTE*MSL*YL
27661 - CTTCTATTGGCTTCCTTTAGCTTCTCTTGTCATCCTCTTGCTTTAAAAATATGCTTGATATATTAT - 27720
- LLFLFVLSLSLSTAIPCFNNRAYY
- FYLCFLAFLLLFLVILLMLILIF
- SICAF*FFGYSLF*+CLLYF

FIG. 11 Con’t
FIG. 11 Con’t
FIG. 11 Con’t
FIG. 11 Con’t
1 - TTTTTTTTTTTTTTGTCACTTCTCCTAAAGAAGCTATTAAATAATCACAATGGAATAGCACTA - 60
   - FFFFFFVILLRSY*NHMGIAL
   - FFFFFLSFS*EAIKITWGH*HY
   - FFFFFCHSPKLLKLKSHGDSTT

61 - CTAATAATTTTTTACATACAAATTAGGCTCTCCATCATAAGCAGCTCTCTCCATGATTATTC - 120
   - LKLILHIRALPYRQLSLALF
   - *N*FYTLGLFHIGSSP*HY
   - KINFTH*GSSIAALPSIIH

121 - ACTGTACCTGTGATCATTACCTGGCTTTAGCTGATGAAATAATTGCTGAGTTAAGATCATAACCTA - 180
   - TYPTRSARMTPKCGGSK
   - CTLDRTPRGLDENUVALSRS

181 - CTCCTAAATGTCTACAGTAAATTGCTATAGATAAAAGATTTACATGTGAGTATAAGTTAAACTAAACCTA - 240
   - LPNVTH*LKIAM*DS*LNL
   - SMLHID*RLLCIEIKVNTY
   - P*CYTLKDCYVRKLKTLKPT

241 - CTTGCTGTTTATGTTAGGAAATATTCCATGCAACAGAGTAGACTGATATGCATGFAACG - 300
   - LVLFSYSNFSCTRVDYVT*T
   - LCCVLTRIHSAQ*TMYRKR
   - CAV*LREFILKHSLRCIVNG

301 - GAATTCCGAAGAAGTTTACATAGACCATGGCCTTGTTGTTGCTGATGAAATGCATTGGTC - 360
   - ERLKRKLHPSALCGHKHECLC
   - NENVYIAHLPCVVMYSYVA
   - IAKTFT*PICLVWSS*VFMP

361 - CTAGTGTAAATGCAGGAAATTCACATGCTATGGAATTGGTGAATGGTGGGAGAAATCGTC - 420
   - LSV*ISRSTTHGILKLSGEII
   - *VESAEAPMELMFC*SCLEGKSS
   - ELNQQKHLHSWNEFVVVRNHNP

421 - CATGCTACCGCCAGGAAAGAGACACAATGTTGGGCTCTCTTTTGTCTCTGCGCCAGAAGG - 480
   - HVSSRRKSHSGGLLSSLLSRQR
   - MSAAAGRRVTVGCFCCLCGCWK
   - CQPQEEESQWAASFVSAAKA

481 - CTAGCTTACACATTACCTCTTGTCTTTGTCCTTTTTTTGTTGGCAGCTGTGGTGAGTTTTTT - 540
   - LSFISSLFLVFLFLRCLWWECF
   - *ASSVFHLSFGLGSVGGNVL
   - ELHQSFSFCFF*ALVGMFC

541 - GTATGGGCTAATGTTGCTGTTCATAGACGATGTTGGCTCTTTGTGAGTTTTTCGTGTGTGCTC - 600
   - VCVNVLVLQQYDVDFELLWIFV
   - YASMCLFSSMTLSLNCGLSL
   - MRQCACSAV*RCL*IVDLCH

601 - ATCCAAATTAATGCGTCTACATGAAATGCGCAACACATGCTTCCAGGATTGTGACTTCCATGGCC - 660
   - IQFNGSMMISQPCSCRRCDPHA
   - SNLMAP***VSHVPEGVTSMMP
   - PIPWLHDKSMFPPKV*LPCQ

661 - AATTGCCTGACATCTCCAAAGATATGCAAGGCAACATGCTTCCAGAATTGGCTGAAATGGCCCA - 720
   - NASHKCECRGTWSKLCNLRP
   - MARDPKNAELGANCAICYCGQ
   - CVTFQORMQHRLEQUIQVFAAN

721 - ATGTTTGAAATCAGTCTCCATTGCTAGAGTTGGCTGCTGGCTCCAGAAATTTCTCTGGGTTTTG - 780
   - MFFVIISSLDS*VVLVPESILGL
   - CL*SVPCILIRSWSPKFPWVVC
   - VCNQFPLV*LGLGPLPRNFLG

781 - TTCTGCAGACAGCTCTCCCAAATGCTGAGCTGTACGTACTTTGTGGGAGTAGCTTTT - 840
   - FWTTSPKCLKSDDVLFLFCCGSTF
   - SGPRLPLNA*VTLYCFVAVRFL
   - LDHVSQMLE*RCTVWLWQYVF

FIG. 12
FIG. 12 Con't
FIG. 12 Con't
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FIG. 12 Con't
FIG. 12 Con't
FIG. 12 Cont'

7561 - AATTGTGATATCTTATATATGAGGAGTCACTTAAAAATAGGTTTCAAAAGTGTTAAACACAG - 7620
- N L * C * Y Q E A T * K * V S K C * N Q
- I C N V N T K R Q L K N R F Q S V K T R
- F V M L I P R G N L K I G F K V L K P E
7621 - AAGGTGAGTACGAGATCTACATGCTATGGTGATAGCCCTTATATAAACCAGATAGAGAACCCAT - 7680
- K V D H E L H L * V D S P Y K H R E T H
- R * I T N I Y I V R L I A L I N I E K P I
- G R S T T S I G * * P L * T A R N P S
7681 - CTTTATTTTTTACACACAAACACCTCCTCTGTAAGGTGTTAAAAATACCTGACTTTTCTCAGGACCATC - 7740
- L Y F * T Q T L V S V * N Y L T L F K H
- F I K H K L S * V F K I T * L F * N I
- L F L N T N S R K C L K L P D F S E T S
7741 - CAAGCGAAAAGGCCATCCAGATATGTAATCCTGCAAAGTGCATATTTAAATGCTATATGTAATCA - 7800
- Q A K R H Q I C T R K C N * M H Y R I S
- K R K G I R Y V L E S A I K C I I E Y H
- S E K A S D M Y S K V Q L N A L S N I I
7801 - TAGATGTCCTGCTCTGTAACCATACTGTTGGAGGAAAGAGGAAGGTGTTTGGCACAACACTG - 7920
- * Y V S V Y P V W * K Q R K G C H T I
- S M L C T H G F R N S K R V R V T Q F
- V C V C V P M G L E T A K K G L S H N S
7861 - CAAAATGGTCTCCTGCTATATAAACAACATTAGTGAAATTGTTTTAATAAATACTACCAGCAGTG - 7920
- Q S Y M L V * Q H * * N C * * * S P T V
- K V T C S Y N N I S R I V N N N H R L *
- K L H A R I T T L V E L L I I T D C D
7921 - ACTGTGTGGTTATGGTAGACCAAAACACCAACGCGACAACTTGGATTTCTCTGTGG - 7980
- T C C S W * N Q K P N H G Q H L I S L W
- L V V H G R T K N P T T D N I * F L C G
- L L F M V E F P K T Q P R T T F D S V A
7981 - CAGCAAAATATAATAACCCATCCTTTAAAGGATATGACAGAGGTGACGTTCCAACACGTATGATTAATAG - 8040
- Q Q N K Y H P * K V * Q G C Q T Y D * *
- S K I N T I L K R Y D R V A K R M I N S
- A K * I P S L K G M T G L P N V * L I V
8041 - ATATGAAACCCTATGTACATAGATTAAAATGGAAGAATAATAATTCTGAGTTAAATAGAAG - 8100
- Y E T L * H N K M E I N P E L N K E
- * N P V T L E N G R N K S * V K * R V
8101 - TGCTGATCTAAAATTTCACTGAGATATACACCCCCCTCCTCTAGATGAGTATGATG - 8160
- C L I * K F H Q D S K P P S * M K Y V E
- V * S K N F I R I V N P P H R * S M L S
- S D L K I S S G * * * P L I D E V C * V
8161 - TGTAATTAGGAGCTGTGGAATCATCACAAAATGTTGTGACCCCGGTCTAGGTACACTACCAGTA - 8220
- C N * E L E H H Q K W C T G Q G Q H Y H *
- V I R S L N I I K S G A P V K V T T T S
8221 - TGAAGAGTTAGAAATATATAGAATAAAATTAAATGATAGTTGCTTTTGTGTTTAACCAAGATAATGCA - 8280
- * E * E I I I A K * T C S F S C * Q E Y H
- E S K K * E N K H V R L V V V N K N I T
- R V R N N K K I N M F V * L L T R I S L
8281 - TGAGAGCACCACACTCTGCTTGGTTTTTCTCTAATGATAAGCCCTACCTTTTCCAGAGAGAAGAT - 8340
- L K P Q L C C F L * * * A Y L F P E E N
- * N H N S V V F S N D K P T F F Q K R I
- E T T T I L F S L M I S L P S F S R E *
8341 - AAATCTATATCTTATTGATTCCTCTCTTAAGAGACATTACACAGGCTTTCTCCTATATTTTA - 8400
- K S Y H I * F D S P * E T L Q Q F L L I *
- N H I I D L L K R H Y S S S * F K
- I I S L I * F S L R D I T A V P L N L R
FIG. 12 Con't

8401 - GAGGAAATTTGCTTACATGCAAGAGGTGAAATGAGAAGACACACTTGAGATAGGATTTTGTTGTCC - 8460
- E E I C S C Q R V N R K T G * D L C S
- G N L L M S K S E * E D N W I G F V F L
8461 - TCCAGAAAATGTCAGGACATGCTGATATGACACATATTATGCTCTTCTGCTGCTGAA - 8520
- S R K C S * H A W Y S H Q F V P S A C Q
- P E N V V S M H G I A I N L F L R L R L K
- Q K M * L A C M V * P S I C S F G L P R
8521 - GATAGTGTTAATGGCCATATCAAATATGCGTCTGATGTGATGCTCTTACATTGTAAACAAAA - 8580
- D S * P Q L K M L P M M M H L H L * Q K
- I V S P N * K C F R * C I Y I C N K S
- L A P I K N A S D D D A T F V T K A
8580 - CGTGCACCAGATGAAATGGCCATATAATCTGTTGCAAGTCAGAAGATCGTCTGAGATC - 8640
- L S T M R N G P * A C K G Q H S K N A L
- C P P * E M A H K L V K V S I P R M L C
- V H H E K W P I S L * R S A F Q E C S V
8641 - TTATCTTTTACAGCTATAGACACCCAGGCTGATTTTGTCTTATAAATCCACACAGAT - 8700
- L S L Q * H P G L V F A L * I H T D
- Y L Y S Y R T T Q G * F L L Y K S T Q I
- I F T A I E P F P R A S F C I N P H R *
8700 - AAAGTGAAAACCCCTTCTCTAAGATGCTCTTCTTTCTGTACACATTTGGTTTGCTAGGTCT - 8760
- K * K T L L * S H L S L S H V W S * G H
- S E K P F F R V I L F C H M F G P R V I
- V K N P S L E S F S F V T C L V L G S Y
8761 - ACATATCCGTGTTATATAATAGGTGCGTCTGTTGATATGCTGATGCTCTCACAACAA - 8820
- T Y R * * G P I Y * F V V L L H S L Q
- H I A N N K V P F I S R M Y C C T V S N
- I S L I I R S H L L L A V C T V A Q S P I
8820 - TTAAAGTGAATACCTCGGTCCAGGACAGGATCTAAAGATCGATCTGCCAGACATGCTG - 8880
- L K * N L R R R R S H * D L N R Q V V C
- S R I C V G D E V I K I * I D K * C A
- K V E S A T K S L R S E S T S S V P
8880 - CAGTTTGCCACCCCTTGTGCTGACTCAGGCTGATCTGGAGAATCGCAGACAGATGCTG - 8940
- Q L A T I V * A Q L Y L V Q L L Y Q S Q
- S W Q P L S E H S C T W C N S F I R A S
- V G N H C L S T A V P P G A T P L S E P A
8940 - CACCAAGAGTCAGATAACTCTACCTGATGGTACGATCTAAAGATGAAGTGGTAAGTAAATGTAT - 9000
- H Q S E * L S C R C V Q L K * V Y L S I
- T K V N N S H V V G Y S * S K C I * V L
- P K * I T L M L * G T A K V S V F K Y *
9001 - GACACAGTTGATGATATTTTGCGCACATTCAATTATTCTCTTTTGTGATATACGATTGAAAT - 9060
- D T V E Y T L R H S L S L F L V * Q H F
- T Q L S I L C D I H H Y S F W Y N S I F
- H S * V Y F A T F I I I I P F G I T A F S
9061 - CACCCATAATTGTGAGGCACACTTCTTTAAGAAGGACTCTTTTGCACTTTTGACAAAGTATAG - 9120
- H H N S E G H F Q E A F F A S C T S *
- T I L K V T L F K K H S L H L V L V Q V R
- P * F * R S H F S R S I L C I L Y K L G
9120 - GACATCGAACAACCTCGGTGCTCAGGCTGCTTTGTTTGTAAGTAGCCCTGCAAA - 9180
- A S Q H L V V A T L D L L L L V L V L G R F Q
- H R N T W L P R L T C L * F W V E G F N
- I A T P G C H A * L A C S F G * K V S T
9180 - CATGTCATCTCTCTAACACAAAGGATGATGAAATTCTCGGACATCTATGCAATGACAACATCA - 9240
- H V H P Y T K A * M K F Q H S Q L * P *
- M S I L T P K H E * N F S I V N C N L D
- C P S L H Q S M N E I S A * S I V T L
FIG. 12 Con't
FIG. 12 Con't
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FIG. 12 Con't
FIG. 12 Con't
FIG. 12 Con't
19321 - ACCACATGTACCATTAAAGGAAAGAGACCTTTATTATGTTACGATTTAGGCCTCACAGCAGACTG - 19380
   - TT* SIKERTFGIHRSMHTGL
   - PHDPRLRKEPELMV* LGLMMAH*
   - HMHI*KGNL* WYD* VSWHTD
19381 - ATAAACACCAAGATGTTAAGAACATCTATGGCTAAGAACATACGAAATATGGTTGACCCAGTTG - 19440
   - ITRW* TIVAC*N KCLTRL
   - TPDGPLEL* HARTENV* PGW
   - KHQMVNHCMSLHLMFLKMDQV
19441 - GATACGACGCAAAATTATCTCCTTGGTCAATCTGGATGATCTCATTATAAAGCTCTAAG - 19500
   - DTDFKILGCRLVRSDINFKPK
   - TIRNLYLGVGLGLEYSTLSLS
   - YGQIYTWVS* G*KYQL*A*A
19501 - CAGACACATTTCGATGAATGGCCTAATACGAGGGTGCTATGACCTGCTGGAGAAAG - 19560
   - QTLIRMANNNTKLNIALSNK
   - RQFCIEWPITRSA*TLPATR
   - DNFA*NGQ*HEVHCOPEQE
19561 - AAACGCTAGTTGGACGATCAGATCTCTGGGAATGACGCAATCTCACTTCAATGATGAAGTACGATCTTC - 19620
   - KAMVGFAANEQIFVIRKHKVF
   - KLWLDLRSRSSLGLLSMSSS
   - SYGWCIE*AADLHS*D*ACLL
19621 - TGGCTGTGGCCAAATGGACATGTCTCTGGGAATGACGCAATCTCACTTCAATGATGAAGTACGATCTTC - 19680
   - CCANDSAWMSTVCTCVIPQPSIK
   - AQVMTCLGQYTVSSNHNPLR
   - LCKHVLDSILCHPTTIIHE
19681 - AGTTGTGATCCACGCGTCTCTGGGACGCTTTAGCCTGTGACGCAATTACGATCTTC - 19740
   - SCSSTGTLYLHAPFNFAG*REC
   - VVVFQPVTCTMHPSTLQPSEGNA
   - L*FHRLLVLPCTLQLCIFTGM
19741 - CATTTGCCATTAAACGAACACTCGAGAAAGCAGAAAGTGATTGATCTGGCTGCTGGGTA - 19800
   - HFPTKKTTLQNSRSD*CLWWLVT
   - IFKPLCRTAEVIDVCNGGW*
   - FSNHSAEQQKLMSVVVRG
19801 - GAGAACACTACGCCAAGATGCTCTGGGAATGACGCAATCTCACTTCAATGATGAAGTACGATCTTC - 19860
   - EINSTVAVKVI*SLC*VAAS
   - ERTSAPELLLKSFRAFAKWQQA
   - EHQLHSC*SHEPLELSSGSKL
19861 - TGCTCTACGACTATGCTGGTAATCTATGGCTACCTGAAATATTCTGTACCTTGATATATAG - 19920
   - CFTIAGSIA*GSTEIULVVLV
   - ASR*LVSVSKAPLKYLYLLYR
   - LHDSWYLRLHLNTCTCYIE
19921 - AGCAGAGATACCTGGTATACCTGGTAAAGTCGACAGTGCTCGCTACGCAATTATGGTTA - 19980
   - SKIPVILCCKWQQCQLATQF*V
   - ARLLLYCVSVGNSSVSLRNRFRY
   - QDTCYTVD*VATVSRAYAILG
19981 - CATTTCTGTTGGACGAAATTACAAACGAGCGCAGCTCCAGGCAGGACTAAGGTGAC - 20040
   - HFLVEQKGTQSSLLEGTKCN
   - ISLSSKKVHKAASSKSVKLNVNT
   - FPC*AKRYTKQPFRY*RML
20041 - TCCATATACGTGACTTCTTCTCTGAATGTTTTAAAGAACCCTTGGCTGCTGCAG - 20100
   - SIKHDSPFPKIVVKEEPMAMQLQ
   - PLNMTLFLR*LLKNQWQCFR
   - H*T*LSFSDSC*RTNGSASE
20101 - AGAAAAATGGGAAATCACTGATGCTGCTACGTAAAGGAACCAATGAGGAAAGAATCCGGC - 20160
   - RNYTEYIDCCYPKRHRNRKRKHG
   - EIQNT*IAVIQKGGTIGENMA
   - KYRIRIHLRLSSKKAQ*EKTWQ

FIG. 12 Con't
20161 - AAAACATTGAGGTGAAGCAGAACAAATGACATATTGAGTGAATGAATGTCAAGTCAA - 20220
   - KPLKRSVEK* NITIGEIIECQVQ
   - NH*R RAKNEETSLVK*NVVKYY
   - TIEGEPRKMHHW*NRRMSTS

20221 - GTAAAAGAACGTGCTAGCTCACTCGCAGCAGAAGGCTCTACTACGTCACTATGTACGAGTG
   - VKD* VDSRQVAWSWYQTEYS
   - *KTE*TFRGKLAAGTRQSV
   - KRLSRLPASCEKLVPDR)+

20281 - GAAGAACATCAAAACAAATTGAGCTACAGCAAGCAACATATTGTGCTTCTACACACAAAC - 20340
   - ERHQQKQKCEISNNMVMVLT
   - KDITKNKSALATATTWLYS
   - KTSKTKVH*QQQHGCTH</code>

20341 - ACGTCCATTATTCAATAAAGTACGTGAGCCACACGACATCACAATGACTCTACACACAC
   - TSEIFKLYGTVSTHGNTHY
   - RLNF*IAAQVNTMMAIP
   - V*IS*SSRQHKSPIW*YHQ

20401 - AGGCACTACTGAGACAGACAGAGATCTTACATCGACCATCCAGAAGGAGATTACAGAT
   - SYH*SRRHI*SSTHRKLKRSK
   - ATEADESKAPCTGCTRGEVM
   - FLLQKQTHLKHQPVQAEE

20461 - GTTAGCCTAGTAGCTGCACTGATCAGCAGCAAAATCTCCATGTAGACCGCTGTAGT
   - SYE1HINTENSSSIVNV
   - LAMRFIASFTQPQKTPDRALC
   - *L*DSSQHHRKLLIELCNA

20521 - CTCAATTATTTAGACAGCCCTACTGCTGGATAGCTAGGGAATACCCTCTCTCTGTGAC
   - LIIKINSTTG*RANTFY*PF
   - SSLRTHLPLVDRQIPSTDSL
   - HY*EPIYHWW*IGKYLLLTFR

20581 - GCAATGCAACTCTCTGTACAAGACATCCCATGAGAACATGTTTCTCTTAGCTACACGACAC
   - ACTMSTVLSIKSCYSSNRTL
   - HVPCLQYSAASKVVTTLTPE
   - MYHYYYSTQHQLKKL*QNNP

20641 - CAGTTAAGGTTAGAAAGCAGATGTGAGCAGACATCCATAGCACATTACAGGAGGTCTG
   - QSVKLYDGTIHKHITSVW
   - **RLGNCMMEPISTG
   - GVKG*ETV*WNHP*AHNECLD

20701 - GCAAGACATCTACTATAGAAGACATAGCTGCAGAATGATATGCTCTACACATGAGACCAC
   - TKLTINRRLQISIVITWH
   - RSSLEIEEPSKSLVSQYGT
   - EAHYYKK*NPAN*CHNNMAQ

20761 - AGCTTTGGCCGCTAGCCTATAAAATGTCACCTACACAGGAAAGGAGGAGATCAGGAGGT
   - RFAHSILKNCTLSKSKNASRGG
   - GLLPIALSKIKVIIASAARATQAEV
   - VCP*HFP*KLYTQQQERKQRR

20821 - AGCGAATACATTTACATAGAACCTAGTGTGCTGAATGCTTTAGCCAGCAGACAGAGCGAGAG
   - S KittLNEFGRCVANVANST
   - AGLSLSLEAEGV*QMPLTA
   - QNHITQ*VWKVCSSKCCQH

20881 - AAAAACACAGAGTATAGAAATGCCAAGAATGCTACCGGAGATAGCTCCAGCAAACATCGG
   - KNTR*RMQREVTDSCSQHSTRR
   - KTRGRCCKSSPILALSTVPG
   - KHEINARSHHLLLMSAIPV

20941 - TAAGCGACACATTGAGAACATCTCTCTGATGATAGAATNGCAGCTGACTACAGGCCACG
   - *ARKHYESNLS-CNDSSTYYRNAA
   - KFEKLNKPISSLVMIMAATTGQL
   - SQAL*NQSSL**QLQLQGSSF

FIG. 12 Con't
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FIG. 12 Con't
HUMAN VIRUS CAUSING SEVERE ACUTE RESPIRATORY SYNDROME (SARS) AND USES THEREOF

[0001] This application is a division of U.S. patent application Ser. No. 10/808,121, filed Mar. 24, 2004; which claims priority benefit to U.S. provisional application No. 60/457,031, filed Mar. 24, 2003; U.S. provisional application No. 60/457,730, filed Mar. 26, 2003; U.S. provisional application No. 60/459,931, filed Apr. 2, 2003; U.S. provisional application No. 60/460,357, filed Apr. 3, 2003; U.S. provisional application No. 60/461,265, filed Apr. 8, 2003; U.S. provisional application No. 60/462,805, filed Apr. 14, 2003; and U.S. provisional application No. 60/464,886 filed Apr. 23, 2003, each of which is incorporated herein by reference in its entirety.

[0002] The instant application contains a lengthy Sequence Listing which is being concurrently submitted via triplicate CD-R in lieu of a printed paper copy, and is hereby incorporated by reference in its entirety. Said CD-R, recorded on Jun. 5, 2007, are labeled "CRF," "Copy 1" and "Copy 2", respectively, and each contains only one identical 1.58 MB file (V96611027.APP).

1. INTRODUCTION

[0003] The present invention relates to an isolated novel virus causing Severe Acute Respiratory Syndrome (SARS) in humans ("SARS virus"). The SARS virus is identified to be morphologically and phylogenetically similar to known members of Coronavirusae. The present invention relates to a nucleotide sequence comprising the complete genomic sequence of the SARS virus. The invention further relates to nucleotide sequences comprising a portion of the genomic sequence of the SARS virus. The invention also relates to the deduced amino acid sequences of the complete genome of the SARS virus. The invention further relates to the nucleic acids and peptides encoded by and/or derived from these sequences and their use in diagnostic methods and therapeutic methods, such as for immunogens. The invention further encompasses chimeric or recombinant viruses encoded by said nucleotide sequences and antibodies directed against polypeptides encoded by the nucleotide sequence. Furthermore, the invention relates to vaccine preparations comprising the SARS virus, including recombinant and chimeric forms of said virus as well as protein extracts and subunits of said virus.

2. BACKGROUND OF THE INVENTION

[0004] Recently, there has been an outbreak of atypical pneumonia in Guangdong province in mainland China. Between November 2002 and March 2003, there were 792 reported cases with 31 fatalities (WHO. Severe Acute Respiratory Syndrome (SARS) Weekly Epidemiol. Rev. 2003; 78: 86). In response to this crisis, the Hospital Authority in Hong Kong has increased the surveillance on patients with severe atypical pneumonia. In the course of this investigation, a number of clusters of health care workers with the disease were identified. In addition, there were clusters of pneumonia incidents among persons in close contact with those infected. The disease was unusual in its severity and its progression in spite of the antibiotic treatment typical for the bacterial pathogens that are known to be commonly associated with atypical pneumonia. The present inventors were one of the groups involved in the investigation of these patients.

[0005] All tests for identifying commonly recognized viruses and bacteria were negative in these patients. The disease was given the acronym Severe Acute Respiratory Syndrome ("SARS"). The etiologic agent responsible for this disease was not known until the isolation of SARS virus from the SARS patients by the present inventors as disclosed herein. Namely, the present invention discloses a novel human virus that has been isolated and identified from the patients suffering from SARS. The invention is useful in both clinical and scientific research applications.

3. SUMMARY OF INVENTION

[0006] The present invention is based upon the inventor's isolation and identification of a novel virus causing Severe Acute Respiratory Syndrome in humans ("SARS virus"). The virus was isolated from the patients suffering from SARS in the recent outbreak of severe atypical pneumonia in China. The isolated virus is an enveloped, single-stranded RNA virus of positive polarity which belongs to the order, Nidovirales, of the family, Coronavirusae. Accordingly, the invention relates to the isolated SARS virus that morphologically and phylogenetically relates to known members of Coronavirusae. In a specific embodiment, the isolated SARS virus is that which was deposited with China Center for Type Culture Collection (CCTCC) on Apr. 2, 2003 and recorded an accession number, CCTCC-V200030, as described in Section 7, infra. In another specific embodiment, the invention provides complete genomic sequence of the SARS virus. In a preferred embodiment, the virus comprises a nucleotide sequence of SEQ ID NO:15. In another specific embodiment, the invention provides nucleic acids isolated from the virus. The virus preferably comprises a nucleotide sequence of SEQ ID NO:1, 11 and/or 13 in its genome. In a specific embodiment, the present invention provides isolated nucleic acid molecules comprising or, alternatively, consisting of the nucleotide sequence of SEQ ID NO:1, a complement thereof or a portion thereof, preferably at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:1, or a complement thereof. In another specific embodiment, the present invention provides isolated nucleic acid molecules comprising or, alternatively, consisting of the nucleotide sequence of SEQ ID NO:11, a complement thereof or a portion thereof, preferably at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:11, or a complement thereof. In yet another specific embodiment, the present invention provides isolated nucleic acid molecules comprising or, alternatively, consisting of the nucleotide sequence of SEQ ID NO:13, a complement thereof or a portion thereof, preferably at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200, 2,000,
3,000, 4,000, 5,000, 6,000, 7,000, 8,000, 9,000, 10,000, 11,000, 12,000, 13,000, 14,000, 15,000, 16,000, 17,000, 18,000, 19,000, 20,000, 21,000, 22,000, 23,000, 24,000, 25,000, 26,000, 27,000, 28,000, 29,000, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:15, or a complement thereof. Furthermore, in another specific embodiment, the invention provides isolated nucleic acid molecules which hybridize under stringent conditions, as defined herein, to a nucleic acid molecule having the sequence of SEQ ID NO:1, 11, 13, 15, 16, 240, 737, 737, 1108, 1590 or 1965 or a complement thereof. In one embodiment, the invention provides an isolated nucleic acid molecule which is antisense to the coding strand of a nucleic acid of the invention. In another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising or, alternatively consisting of a nucleotide sequence that is at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:1, or a complement thereof. In yet another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising or, alternatively consisting of a nucleotide sequence that is at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200 or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:11, or a complement thereof. In yet another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising or, alternatively consisting of a nucleotide sequence that is at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200, 2,000, 3,000, 4,000, 5,000, 6,000, 7,000, 8,000, 9,000, 10,000, 11,000, 12,000, 13,000, 14,000, 15,000, 16,000, 17,000, 18,000, 19,000, 20,000, 21,000, 22,000, 23,000, 24,000, 25,000, 26,000, 27,000, 28,000, 29,000, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:15, or a complement thereof. In yet another embodiment, the invention provides polypeptides or proteins that are isolated from the hSARS virus, including viral proteins isolated from cells infected with the virus but not present in comparable uninfected cells. The invention further provides proteins or polypeptides of SEQ ID NO:2, 12 and 14 and those shown in FIGS. 11 (SEQ ID NO:S:17-239, 241-736 and 738-1107) and 12 (1109-1589, 1591-1964, 1966-2470). The polypeptides or the proteins of the present invention preferably have a biological activity of the protein (including antigenicity and/or immunogenicity) encoded by a nucleotide sequence that is at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200, 2,000, 3,000, 4,000, 5,000, 6,000, 7,000, 8,000, 9,000, 10,000, 11,000, 12,000, 13,000, 14,000, 15,000, 16,000, 17,000, 18,000, 19,000, 20,000, 21,000, 22,000, 23,000, 24,000, 25,000, 26,000, 27,000, 28,000, 29,000, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:15, or a complement thereof. In another embodiment, the invention relates to the use of the isolated hSARS virus for diagnostic and therapeutic methods. In a specific embodiment, the invention provides a method of detecting in a biological sample an antibody immunospecific for the hSARS virus using the isolated hSARS virus or any proteins or polypeptides thereof. In another specific embodiment, the invention provides a method of screening for an antibody which immunospecifically binds and neutralizes hSARS. Such an antibody is useful for a passive immunization or immunotherapy of a subject infected with hSARS.

[0007] The invention further relates to the use of the sequence information of the isolated virus for diagnostic and therapeutic methods. In a specific embodiment, the invention provides nucleic acid molecules which are suitable for use as primers consisting of or comprising the nucleotide sequence of SEQ ID NO:1, 11, 13, or 15, a complement thereof, or at least a portion of the nucleotide sequence thereof. In another specific embodiment, the invention provides nucleic acid molecules which are suitable for hybridization to hSARS nucleic acid, including but not limited to, as PCR primers, Reverse Transcriptase primers, probes for Southern analysis or other nucleic acid hybridization analysis for the detection of hSARS nucleic acids, e.g., consisting of or comprising the nucleotide sequence of SEQ ID NO:1, 11, 13, or 15, a complement thereof, or a portion thereof. The invention further encompasses chimeric or recombinant viruses encoded in whole or in part by said nucleotide sequences.

[0008] The invention further provides antibodies that specifically bind a polypeptide of the invention encoded by the nucleotide sequence of SEQ ID NO:1, 11, 13, 16, 240, 737, 737, 1108, 1590 or 1965, or a fragment thereof, or encoded by a nucleic acid comprising a nucleotide sequence that hybridizes under stringent conditions to the nucleotide sequence of SEQ ID NO:1, 11, or 13, and/or any hSARS epitope, having one or more biological activities of a polypeptide of the invention. The invention further provides antibodies that specifically bind polypeptides of the invention encoded by the nucleotide sequence of SEQ ID NO:15 or a complement thereof, or a fragment thereof. These polypeptides include those shown in FIGS. 11 (SEQ ID NO:17-239, 241-736 and 738-1107) and 12 (SEQ ID NO:S:1109-1589, 1591-1964 and 1966-2470). The invention further provides antibodies that specifically bind polypeptides of the invention encoded by a nucleic acid comprising a nucleotide sequence that hybridizes under stringent conditions to the nucleotide sequence of SEQ ID NO:15, and/or any hSARS epitope, having one or more biological activities of a polypeptide of the invention. Such antibodies include, but are not limited to polyclonal,
monoclonal, bispecific, multispecific, human, humanized, chimeric antibodies, single chain antibodies, Fab fragments, F(ab')2 fragments, disulfide-linked Fv's, intrabodies and fragments containing either a V.L. or V.H domain or even a complementary determinant region (CDR) that specifically binds to a polypeptide of the invention.

In one embodiment, the invention provides methods for detecting the presence, activity or expression of the hSARS virus of the invention in a biological material, such as cells, blood, saliva, urine, and so forth. The increased or decreased activity or expression of the hSARS virus in a sample relative to a control sample can be determined by contacting the biological material with an agent which can detect directly or indirectly the presence, activity or expression of the hSARS virus. In a specific embodiment, the detecting agents are the antibodies or nucleic acid molecules of the present invention. Antibodies of the invention may also be used to treat SARS.

In another embodiment, the invention provides vaccine preparations, comprising the hSARS virus, including recombinant and chimeric forms of said virus, or protein subunits of the virus. In a specific embodiment, the vaccine preparations of the present invention comprise live but attenuated hSARS virus with or without adjuvants. In another specific embodiment, the vaccine preparations of the invention comprise an inactivated or killed hSARS virus. Such attenuated or inactivated viruses may be prepared by a series of passages of the virus through the host cells or by preparing recombinant or chimeric forms of virus. Accordingly, the present invention further provides methods of preparing recombinant or chimeric forms of hSARS. In another specific invention, the vaccine preparations of the present invention comprise a nucleic acid or fragment of the hSARS virus, e.g., the virus having accession no. CCTCC-V2000303, or nucleic acid molecules having the sequence of SEQ ID NO. 1, 11, 13, or 15, or a fragment thereof. In another embodiment, the invention provides vaccine preparations comprising one or more polypeptides isolated from or produced from nucleic acid of hSARS virus, for example, of deposit accession no. CCTCC-V2000303. In a specific embodiment, the vaccine preparations comprise a polypeptide of the invention encoded by the nucleotide sequence of SEQ ID NO:1, 11, 13, 16, 240, 737, 1108, 1590 or 1665, or a fragment thereof. In a specific embodiment, the vaccine preparations comprise polypeptides of the invention as shown in FIGS. 11 (SEQ ID NOS:17-239, 241-736 and 738-1107) and 12 (SEQ ID NOS:1109-1589, 1591-1964 and 1966-2470) or encoded by the nucleotide sequence of SEQ ID NO:15, or a fragment thereof. Furthermore, the present invention provides methods for treating, ameliorating, managing or preventing SARS by administering the vaccine preparations or antibodies of the present invention alone or in combination with adjuvants, or other pharmaceutically acceptable excipients.

In another aspect, the present invention provides pharmaceutical compositions comprising anti-viral agents of the present invention and a pharmaceutically acceptable carrier. In a specific embodiment, the anti-viral agent of the invention is an antibody that immunospecifically binds hSARS virus or any hSARS epitope. In another specific embodiment, the anti-viral agent is a polypeptide or protein of the present invention or nucleic acid molecule of the invention. The invention also provides kits comprising a pharmaceutical composition of the present invention.

3.1 Definitions

The term "an antibody or an antibody fragment that immunospecifically binds a polypeptide of the invention" as used herein refers to an antibody or a fragment thereof that immunospecifically binds to the polypeptide encoded by the nucleotide sequence of SEQ ID NO:1, 11, 13 or 15, or a fragment thereof, and does not non-specifically bind to other polypeptides. An antibody or a fragment thereof that immunospecifically binds to the polypeptide of the invention may cross-react with other antigens. Preferably, an antibody or a fragment thereof that immunospecifically binds to a polypeptide of the invention does not cross-react with other antigens. An antibody or a fragment thereof that immunospecifically binds to the polypeptide of the invention can be identified by, for example, immunoassays or other techniques known to those skilled in the art.

An "isolated" or "purified" polypeptide or protein is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the protein is derived, or substantially free of chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of a polypeptide/protein in which the polypeptide/protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. Thus, a polypeptide/protein that is substantially free of cellular material includes preparations of the polypeptide/protein having less than about 30%, 20%, 10%, 5%, 2.5%, or 1%, (by dry weight) of contaminating protein. When the polypeptide/protein is recombinantly produced, it is also preferably substantially free of culture medium, i.e., culture medium represents less than about 20%, 10%, or 5% of the volume of the protein preparation. When polypeptide/protein is produced by chemical synthesis, it is preferably substantially free of chemical precursors or other chemicals, i.e., it is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. Accordingly, such preparations of the polypeptide/protein have less than about 30%, 20%, 10%, 5%, (by dry weight) of chemical precursors or compounds other than polypeptide/protein fragment of interest. In a preferred embodiment of the present invention, polypeptides/proteins are isolated or purified.

An "isolated" nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid molecule. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. In a preferred embodiment of the invention, nucleic acid molecules encoding polypeptides/proteins of the invention are isolated or purified. The term "isolated" nucleic acid molecule does not include a nucleic acid that is a member of a library that has not been purified away from other library clones containing other nucleic acid molecules.

The term "portion" or "fragment" as used herein refers to a portion of a nucleic acid molecule containing at least about 25, 30, 35, 40, 45, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050, 1100, 1150, 1200, 1250, 1300, 1350, 2,000,
3,000, 4,000, 5,000, 6,000, 7,000, 8,000, 9,000, 10,000, 11,000, 12,000, 13,000, 14,000, 15,000, 16,000, 17,000, 18,000, 19,000, 20,000, 21,000, 22,000, 23,000, 24,000, 25,000, 26,000, 27,000, 28,000, 29,000, or more contiguous nucleic acids in length of the relevant nucleic acid molecule and having at least one functional feature of the nucleic acid molecule (or the encoded protein has one functional feature of the protein encoded by the nucleic acid molecule); or a fragment of a protein or a polypeptide containing at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 90, 100, 120, 140, 160, 180, 200, 220, 240, 260, 280, 300, 320, 340, 360, 400, 500, 600, 700, 800, 900, 1,000, 1,500, 2,000, 2,500, 3,000, 3,500, 4,000, 4,500, 5,000, 5,500, 6,000, 6,500, 7,000, 7,500, 8,000, 8,500, 9,000, 9,500, 10,000, 10,500, 11,000, 11,500, 12,000, or more residues of the relevant protein or polypeptide and having at least one functional feature of the protein or polypeptide.

[0017] The term “having a biological activity of the protein” or “having biological activities of the polypeptides of the invention” refers to the characteristics of the polypeptides or proteins having a common biological activity similar or identical structural domain and/or having sufficient amino acid identity to the polypeptide encoded by the nucleotide sequence of SEQ ID NO:1, 11, 13, 15, 16, 240, 737, 1108, 1590 or 1965. Such common biological activities of the polypeptides of the invention include antigenicity and immunogenicity.

[0018] The term “under stringent condition” refers to hybridization and washing conditions under which nucleotide sequences having at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95% identity to each other remain hybridized to each other. Such hybridization conditions are described in, for example but not limited to, Current Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6.; Basic Methods in Molecular Biology, Elsevier Science Publishing Co., Inc., N.Y. (1986), pp. 75-78, and 84-87; and Molecular Cloning, Cold Spring Harbor Laboratory, N.Y. (1982), pp. 387-389, and are well known to those skilled in the art. A preferred, non-limiting example of stringent hybridization conditions is hybridization in 6x sodium chloride/sodium citrate (SSC), 0.5% SDS at about 68°C followed by one or more washes in 2xSSC, 0.5% SDS at room temperature. Another preferred, non-limiting example of stringent hybridization conditions is hybridization in 6xSSC at about 45°C followed by one or more washes in 0.2xSSC, 0.1% SDS at about 50-65°C.

[0019] The term “variant” as used herein refers either to a naturally occurring genetic mutant of hSARS or a recombinantly prepared variation of hSARS each of which contain one or more mutations in its genome compared to the hSARS of CCTCC-V200303. The term “variant” may also refers either to a naturally occurring variation of a given peptide or a recombinantly prepared variation of a given peptide or protein in which one or more amino acid residues have been modified by amino acid substitution, addition, or deletion.

4. DESCRIPTION OF THE FIGURES

[0020] FIG. 1 shows a partial DNA sequence (SEQ ID NO:1) and its deduced amino acid sequence (SEQ ID NO:2) obtained from the SARS virus that has 57% homology to the RNA-dependent RNA polymerase protein of known Coronaviruses.

[0021] FIG. 2 shows an electron micrograph of the novel hSARS virus that has similar morphological characteristics of coronaviruses.

[0022] FIG. 3 shows an immunofluorescent staining for IgG antibodies that are specifically bound to the H4HK-4 cells infected with the novel human respiratory virus of Coronavirus.

[0023] FIG. 4 shows an electron micrograph of ultra-centrifuged deposit of hSARS virus that was grown in the cell culture and negatively stained with 3% potassium phosphotungstate at pH 7.0.

[0024] FIG. 5A shows a thin-section electron micrograph of lung biopsy of a patient with SARS; and FIG. 5B shows a thin section electron micrograph of hSARS-infected cells.

[0025] FIG. 6 shows the result of phylogenetic analysis for the partial protein sequence (215 amino acids: SEQ ID NO:2) of the hSARS virus (GenBank accession number: KY260708). The phylogenetic tree is constructed by the neighbor-joining method. The horizontal-line distance represents the number of sites at which the two sequences compared are different. Bootstrap values are deduced from 500 replicates.

[0026] FIG. 7A shows an amplification plot of fluorescence intensity against the PCR cycle in a real-time quantitative PCR assay that can detect a hSARS virus in samples quantitatively. The copy numbers of input plasmid DNA in the reactions are indicated. The X-axis denotes the cycle number of a quantitative PCR assay and the Y-axis denotes the fluorescence intensity (FI) over the background. FIG. 7B shows the result of a melting curve analysis of PCR products from clinical samples. Signals from positive (+ve) samples, negative (−ve) samples and water control (water) are indicated. The X-axis denotes the temperature (°C) and the Y-axis denotes the fluorescence intensity (FI) over the background.

[0027] FIG. 8 shows another partial DNA sequence (SEQ ID NO:11) and its deduced amino acid sequence (SEQ ID NO:12) obtained from the SARS virus.

[0028] FIG. 9 shows yet another partial DNA sequence (SEQ ID NO:13) and its deduced amino acid sequence (SEQ ID NO:14) obtained from the SARS virus.

[0029] FIG. 10 shows the entire genomic DNA sequence (SEQ ID NO:15) of the SARS virus.

[0030] FIG. 11 shows the deduced amino acid sequences obtained from SEQ ID NO:15 in three frames (see SEQ ID NO:16, 240 and 737). An asterisk (*) indicates a stop codon which marks the end of a peptide. The first-frame amino acid sequences: SEQ ID NO:17-239; the second-frame amino acid sequences: SEQ ID NO:241-736; and the third-frame amino acid sequences: SEQ ID NO:738-1107.

[0031] FIG. 12 shows the deduced amino acid sequences obtained from the complement of SEQ ID NO:15 in three frames (see SEQ ID NO:1108, 1590 and 1965). An asterisk (*) indicates a stop codon which marks the end of a peptide. The first-frame amino acid sequences: SEQ ID NO:1109-1589; the second-frame amino acid sequences: SEQ ID NO:1591-1964; and the third-frame amino acid sequences: SEQ ID NO:1966-2470.

5. DETAILED DESCRIPTION OF THE INVENTION

[0032] The present invention relates to the isolated hSARS virus that morphologically and phylogenetically relates to known Coronaviruses. In a specific embodiment, the isolated hSARS virus is that of CCTCC-V200303. In another specific embodiment, the virus comprises a nucleotide sequence of SEQ ID NO:1, 11, 13, and/or 15. In a specific embodiment, the present invention provides isolated nucleic acid molecules of the hSARS virus, comprising, or, alternatively, con-
sisting of the nucleotide sequence of SEQ ID NO:1, 11, 13, and/or 15, a complement thereof or a portion thereof. In another specific embodiment, the invention provides isolated nucleic acid molecules which hybridize under stringent conditions, as defined herein, to a nucleic acid molecule having the sequence of SEQ ID NO:1, 11, 13, or 15, or specific genes of known member of Coronavirus, or a complement thereof. In another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising a nucleotide sequence that is at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, or more contiguous nucleotide sequences of the nucleotide sequence of SEQ ID NO:1, or a complement thereof. In another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising a nucleotide sequence that is at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200, or more contiguous nucleotide sequences of the nucleotide sequence of SEQ ID NO:11, or a complement thereof. In yet another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising a nucleotide sequence that is at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:13, or a complement thereof. In yet another specific embodiment, the invention provides isolated polypeptides or proteins that are encoded by a nucleic acid molecule comprising or, alternatively consisting of a nucleotide sequence that is at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 100, 150, 200, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1,000, 1,050, 1,100, 1,150, 1,200, 2,000, 3,000, 4,000, 5,000, 6,000, 7,000, 8,000, 9,000, 10,000, 11,000, 12,000, 13,000, 14,000, 15,000, 16,000, 17,000, 18,000, 19,000, 20,000, 21,000, 22,000, 23,000, 24,000, 25,000, 26,000, 27,000, 28,000, 29,000, or more contiguous nucleotides of the nucleotide sequence of SEQ ID NO:15, or a complement thereof. The polypeptides include those shown in FIGS. 11 (SEQ ID NOS: 17-239, 241-736 and 738-1107) and 12 (SEQ ID NOS:1-239, 1591-1589, 1591-1589 and 1591-2470). The polypeptides or the proteins of the present invention preferably have one or more biological activities of the proteins encoded by the sequence of SEQ ID NO:1, 11, 13, 15, or the native viral proteins containing the amino acid sequences encoded by the sequence of SEQ ID NO:1, 11, 13, or 15, or those shown in FIGS. 11 (SEQ ID NOS:17-239, 241-736 and 738-1107) and 12 (SEQ ID NOS:1109-1589, 1591-1589 and 1591-2470).

The present invention also relates to a method for propagating the hSARS virus in host cells.

[0034] The invention further relates to the use of the sequence information of the isolated viruses for diagnostic and therapeutic methods. In a specific embodiment, the invention provides the entire nucleotide sequence of hSARS virus, CTCVC-200305, SEQ ID NO:15, or fragments, or complement thereof. Furthermore, the present invention relates to a nucleic acid molecule that hybridizes any portion of the genome of the hSARS virus, CTCVC-200305, SEQ ID NO:15, under the stringent conditions. In a specific embodiment, the invention provides nucleic acid molecules which are suitable for use as primers consisting of or comprising the nucleotide sequence of SEQ ID NO:1, 11, 13, or 15, or a complement thereof, or a portion thereof. In a non-limiting embodiment, the invention provides the primers consisting of or comprising the nucleotide sequence of SEQ ID NOS:3 and/or 4. In another specific embodiment, the invention provides nucleic acid molecules which are suitable for use as hybridization probes for the detection of nucleic acids encoding a polypeptide of the invention, consisting of or comprising the nucleotide sequence of SEQ ID NO:1, 11, 13, or 15, a complement thereof, or a portion thereof. The invention further encompasses chimeric or recombinant viruses or viral proteins encoded by said nucleotide sequences.
[e.g., amantadine, rimantadine, gancyclovir, acyclovir, ribavirin, penciclovir, oseltamivir, foscamet zidovudine (AZT), didanosine (ddI), lamivudine (3TC), zalcitabine (ddC), stavudine (d4T), nevirapine, delавirdine, indinavir, ritonavir, vidarabine, nelfinavir, saquinavir, relenza, tamiflu, pleconaril, interferons, etc.], steroids and corticosteroids such as prednisone, cortisone, fluticasone and glucocorticoid, antibiotics, analgesics, bronchodilators, or other treatments for respiratory and/or viral infections.

Furthermore, the present invention provides pharmaceutical compositions comprising anti-viral agents of the present invention and a pharmaceutically acceptable carrier. The present invention also provides kits comprising pharmaceutical compositions of the present invention.

In another aspect, the present invention provides methods for screening anti-viral agents that inhibit the infectivity or replication of hSARS virus or variants thereof.

5.1 Recombinant and Chimeric hSARS Viruses

The present invention encompasses recombinant or chimeric viruses encoded by viral vectors derived from the genome of hSARS virus or natural variants thereof. In a specific embodiment, a recombinant virus is one derived from the hSARS virus of deposit accession no. CCTCC-V200303.

In a specific embodiment, the virus has a nucleotide sequence of SEQ ID NO:15. In another specific embodiment, a recombinant virus is one derived from a natural variant of hSARS virus. A natural variant of hSARS has a sequence that is different from the genome sequence (SEQ ID NO:15) of the hSARS virus, CCTCC-V200303, due to one or more naturally occurring mutations, including, but not limited to, point mutations, rearrangements, insertions, deletions etc., to the genome sequence that may or may not result in a phenotypic change. In accordance with the present invention, a viral vector which is derived from the genome of the hSARS virus, CCTCC-V200303, is one that contains a nucleic acid sequence that encodes at least a part of one ORF of the hSARS virus. In a specific embodiment, the ORF comprises or consists of a nucleotide sequence of SEQ ID NO:11 or 13, or a fragment thereof. In a specific embodiment, there are more than one ORF within the nucleotide sequence of SEQ ID NO:15 or a complement thereof, as shown in FIGS. 11 (SEQ ID NOS:16, 240 and 737) and 12 (SEQ ID NOS:1108, 1550 and 1965), or a fragment thereof. In another embodiment, the polypeptide encoded by the ORF comprises or consists of an amino acid sequence of SEQ ID NO:2, 12, or 14, or a fragment thereof, or shown in FIGS. 11 (SEQ ID NOS:17-239, 241-736 and 738-1107) and 12 (SEQ ID NOS: 1109-1589, 1591-1964 and 1966-2470), or a fragment thereof. In accordance with the present invention these viral vectors may or may not include nucleic acids that are non-native to the viral genome.

In another specific embodiment, a chimeric virus of the invention is a recombinant hSARS virus which further comprises a heterologous nucleotide sequence. In accordance with the invention, a chimeric virus may be encoded by a nucleotide sequence in which heterologous nucleotide sequences have been added to the genome or in which endogenous or native nucleotide sequences have been replaced with heterologous nucleotide sequences.

According to the present invention, the chimeric viruses are encoded by the viral vectors of the invention which further comprise a heterologous nucleotide sequence. In accordance with the present invention a chimeric virus is encoded by a viral vector that may or may not include nucleic acids that are non-native to the viral genome. In accordance with the invention a chimeric virus is encoded by a viral vector to which heterologous nucleotide sequences have been added, inserted or substituted for native or non-native sequences. In accordance with the present invention, the chimeric virus may be encoded by nucleotide sequences derived from different strains or variants of hSARS virus. In particular, the chimeric virus is encoded by nucleotide sequences that encode antigenic polypeptides derived from different strains or variants of hSARS virus.

A chimeric virus may be of particular use for the generation of recombinant vaccines protecting against two or more viruses (Tao et al., J Virol. 72, 2955-2961; Durbin et al., 2000, J Virol. 74, 6821-6831; Skladopoulos et al., 1998, J Virol. 72, 1762-1768 (1998); Teng et al., 2000, J. Virol. 74, 9317-9321). For example, it can be envisioned that a virus vector derived from the hSARS virus expressing one or more proteins of variants of hSARS virus, or vice versa, will protect a subject vaccinated with such vector against infections by both the native hSARS and the variant. Attenuated and replication-defective viruses may be of use for vaccination purposes with live vaccines as has been suggested for other viruses. (See, PCT WO 02/057302, at pp. 6 and 23, incorporated by reference herein).

In accordance with the present invention the heterologous sequence to be incorporated into the viral vectors encoding the recombinant or chimeric viruses of the invention include sequences obtained or derived from different strains or variants of hSARS virus.

In certain embodiments, the chimeric or recombinant viruses of the invention are encoded by viral vectors derived from viral genomes wherein one or more sequences, intergenic regions, termini sequences, or portions or entire ORF have been substituted with a heterologous or non-native sequence. In certain embodiments of the invention, the chimeric viruses of the invention are encoded by viral vectors derived from viral genomes wherein one or more heterologous sequences have been inserted or added to the vector.

The selection of the viral vector may depend on the species of the subject that is to be treated or protected from a viral infection. If the subject is human, then an attenuated hSARS virus can be used to provide the antigenic sequences.

In accordance with the present invention, the viral vectors can be engineered to provide antigenic sequences which confer protection against infection by the hSARS and natural variants thereof. The viral vectors may be engineered to provide one, two, three or more antigenic sequences.

In accordance with the present invention the antigenic sequences may be derived from the same virus, from different strains or variants of the same type of virus, or from different viruses.

The expression products and/or recombinant or chimeric viruses obtained in accordance with the invention may advantageously be utilized in vaccine formulations. The expression products and chimeric viruses of the present invention may be engineered to create vaccines against a broad range of pathogens, including viral and bacterial antigens, tumor antigens, allergen antigens, and auto antigens involved in autoimmune disorders. In particular, the chimeric viruses of the present invention may be engineered to create vaccines for the protection of a subject from infections with hSARS virus and variants thereof.
In certain embodiments, the expression products and recombinant or chimeric virions of the present invention may be engineered to create vaccines against a broad range of pathogens, including viral antigens, tumor antigens and autoantigens involved in autoimmune disorders. One way to achieve this goal involves modifying existing hSARS genes to contain foreign sequences in their respective external domains. Where the heterologous sequences are epitopes or antigens of pathogens, these chimeric viruses may be used to induce a protective immune response against the disease agent from which these determinants are derived.

Thus, the present invention relates to the use of viral vectors and recombinant or chimeric viruses to formulate vaccines against a broad range of viruses and/or antigens. The present invention also encompasses recombinant viruses comprising a viral vector derived from the hSARS or variants thereof which contains sequences which result in a virus having a phenotype more suitable for use in vaccine formulations, e.g., attenuated phenotype or enhanced antigenicity. The mutations and modifications can be in coding regions, in intergenic regions and in the leader and trailer sequences of the virus.

The invention provides a host cell comprising a nucleic acid or a vector according to the invention. Plasmid or viral vectors containing the polymerase components of hSARS virus are generated in prokaryotic cells for the expression of the components in relevant cell types (bacteria, insect cells, eukaryotic cells). Plasmid or viral vectors containing full-length or partial copies of the hSARS genome will be generated in prokaryotic cells for the expression of viral nucleic acids in vitro or in vivo. The latter vectors may contain other viral sequences for the generation of chimeric viruses or chimeric virus proteins, may lack parts of the viral genome for the generation of replicative defective virus, and may contain mutations, deletions or insertions for the generation of attenuated viruses. In addition, the present invention provides a host cell infected with hSARS virus, for example, of deposit no. CCTCC-V200303.

Infectious copies of hSARS (being wild type, attenuated, replication-defective or chimeric) can be produced upon co-expression of the polymerase components according to the state-of-the-art technologies described above.

In addition, eukaryotic cells, transiently or stably expressing one or more full-length or partial hSARS proteins can be used. Such cells can be made by transfection (proteins or nucleic acid vectors), infection (viral vectors) or transduction (viral vectors) and may be useful for complementation of mentioned wild type, attenuated, replication-defective or chimeric viruses.

The viral vectors and chimeric viruses of the present invention may be used to modulate a subject's immune system by stimulating a humoral immune response, a cellular immune response or by stimulating tolerance to an antigen. As used herein, a subject means: humans, primates, horses, cows, sheep, pigs, goats, dogs, cats, avian species and rodents.

5.2 Formulation of Vaccines and Antivirals

In a preferred embodiment, the invention provides a proteinaceous molecule or hSARS virus specific viral protein or functional fragment thereof encoded by a nucleic acid according to the invention. Useful proteinaceous molecules are for example derived from any of the genes or genomic fragments derivable from the virus according to the invention, including envpol protein (E protein), integral membrane protein (M protein), spike protein (S protein), nucleocapsid protein (N protein), hemagglutinin esterase (HE protein), and RNA-dependent RNA polymerase. Such molecules, or antigenic fragments thereof, as provided herein, are for example useful in diagnostic methods or kits and in pharmaceutical compositions such as subunit vaccines. Particularly useful are polypeptides encoded by the nucleotide sequence of SEQ ID NO:1, 11, 13, or 15, or as shown in FIGS. 11 (SEQ ID NO:17-239, 241-736 and 738-1107) and 12 (SEQ ID NO: 1109-1589, 1591-1964 and 1966-2470), or antigenic fragments thereof for inclusion as antigen or subunit immunogen, but inactivated whole virus can also be used. Particularly useful are also those proteinaceous substances that are encoded by recombinant nucleic acid fragments of the hSARS genome, of course preferred are those that are within the preferred bounds and moles of ORFs, in particular, for eliciting hSARS specific antibody or T cell responses, whether in vivo (e.g. for protective or therapeutic purposes or for providing diagnostic antibodies) or in vitro (e.g. by plaque display technology or another technique useful for generating synthetic antibodies).

The invention provides vaccine formulations for the prevention and treatment of infections with hSARS virus. In certain embodiments, the vaccine of the invention comprises recombinant and chimeric viruses of the hSARS virus. In certain embodiments, the virus is attenuated.

In another embodiment of this aspect of the invention, inactivated vaccine formulations may be prepared using conventional techniques to "kill" the chimeric viruses. Inactivated vaccines are “dead” in the sense that their infectivity has been destroyed. Ideally, the infectivity of the virus is destroyed without affecting its immunogenicity. In order to prepare inactivated vaccines, the chimeric virus may be grown in cell culture or in the allantois of the chick embryo, purified by zonal ultracentrifugation, inactivated by formaldehyde or β-propiolactone, and pooled. The resulting vaccine is usually inoculated intramuscularly.

Inactivated vaccines may be formulated with a suitable adjuvant in order to enhance the immunological response. Such adjuvants may include but are not limited to mineral gels, e.g., aluminum hydroxide; surface active substances such as lysolecithin, pluronics polyols, polynions; peptides; oil emulsions; and potentially useful human adjuvants such as BCG and Corynebacterium parvum.

In another aspect, the present invention also provides DNA vaccine formulations comprising a nucleic acid or fragment of the hSARS virus, e.g., the virus having accession no. CCTCC-V200303, or nucleic acid molecules having the sequence of SEQ ID NO:1, 11, 13, or 15, or a fragment thereof. In another specific embodiment, the DNA vaccine formulations of the present invention comprise a nucleic acid or fragment thereof encoding the antibodies which immunospecifically bind hSARS viruses. In DNA vaccine formulations, a vaccine DNA comprises a viral vector, such as that derived from the hSARS virus, bacterial plasmid, or other expression vector, bearing an insert comprising a nucleic acid molecule of the present invention operably linked to one or more control elements, thereby allowing expression of the vaccinating proteins encoded by said nucleic acid molecule in a vaccinated subject. Such vectors can be prepared by recombinant DNA technology as recombinant or chimeric viral
vectors carrying a nucleic acid molecule of the present invention (see also Section 5.1, supra).


[0662] Many methods may be used to introduce the vaccine formulations described above. These include, but are not limited to, oral, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, and intranasal routes. Alternatively, it may be preferable to introduce the chimeric virus vaccine formulation via the natural route of infection of the pathogen for which the vaccine is designed. The DNA vaccines of the present invention may be administered in saline solutions by injections into muscle or skin using a syringe and needle (Wolff J. A. et al., 1990, Direct gene transfer into mouse muscle in vivo, Science 247:1465-1468; Raz, E., 1994, Intradermal gene immunization: The possible role of DNA uptake in the induction of cellular immunity to viruses, Proc. Natl. Acad. Sci. USA 91:9519-9523). Another way to administer DNA vaccines is called “gene gun” method, whereby microscopic gold beads coated with the DNA molecules of interest are fired into the cells (Tang, D. et al., 1992, Genetic immunization is a simple method for eliciting an immune response, Nature 356:152-154). For general reviews of the methods for DNA vaccines, see Robinson, H. L., 1999, DNA vaccines: basic mechanism and immune responses (Review), Int. J. Mol. Med. 4(5):549-555; Barber, B., 1997, Introduction: Emerging vaccine strategies, Seminars in Immunology 9(5):269-270; and Robinson, H. L. et al., 1997, DNA vaccines, Seminars in Immunology 9(5):271-283.

5.3 Attenuation of hSARS Virus or Variants Thereof

[0663] The hSARS virus or variants thereof of the invention can be genetically engineered to exhibit an attenuated phenotype. In particular, the viruses of the invention exhibit an attenuated phenotype in a subject to which the virus is administered as a vaccine. Attenuation can be achieved by any method known to a skilled artisan. Without being bound by theory, the attenuated phenotype of the viruses of the invention can be caused, e.g., by using a virus that naturally does not replicate well in an intended host species, for example, by reduction replication of the viral genome, by reduced ability of the virus to infect a host cell, or by reduced ability of the viral proteins to assemble to an infectious viral particle relative to the wild type strain of the virus.

[0664] The attenuated phenotypes of hSARS virus or variants thereof can be tested by any method known to the artisan. A candidate virus can, for example, be tested for its ability to infect a host or for the rate of replication in a cell culture system. In certain embodiments, growth curves at different temperatures are used to test the attenuated phenotype of the virus. For example, an attenuated virus is able to grow at 35° C., but not at 39° C. or 40° C. In certain embodiments, different cell lines can be used to evaluate the attenuated phenotype of the virus. For example, an attenuated virus may only be able to grow in monkey cell lines but not the human cell lines, or the achievable virus titers in different cell lines are different for the attenuated virus. In certain embodiments, viral replication in the respiratory tract of a small animal model, including but not limited to, hamsters, cattle, mice and guinea pigs, is used to evaluate the attenuated phenotypes of the virus. In other embodiments, the immune response induced by the virus, including but not limited to, the antibody titers (e.g., assayed by plaque reduction neutralization assay or ELISA) is used to evaluate the attenuated phenotypes of the virus. In a specific embodiment, the plaque reduction neutralization assay or ELISA is carried out at a low dose. In certain embodiments, the ability of the hSARS virus to elicit pathological symptoms in an animal model can be tested. A reduced ability of the virus to elicit pathological symptoms in an animal model system is indicative of its attenuated phenotype. In a specific embodiment, the candidate viruses are tested in a monkey model for nasal infection, indicated by mucous production.

[0665] The viruses of the invention can be attenuated such that one or more of the functional characteristics of the virus are impaired. In certain embodiments, attenuation is measured in comparison to the wild type strain of the virus from which the attenuated virus is derived. In other embodiments, attenuation is determined by comparing the growth of an attenuated virus in different host systems. Thus, for a non-limiting example, hSARS virus or a variant thereof is said to be attenuated when grown in a human host if the growth of the hSARS or variant thereof in the human host is reduced compared to the non-attenuated hSARS or variant thereof.

[0666] In certain embodiments, the attenuated virus of the invention is capable of infecting a host, is capable of replicating in a host such that infectious viral particles are produced. In comparison to the wild type strain, however, the attenuated strain grows to lower titers or grows more slowly. Any technique known to the skilled artisan can be used to
determine the growth curve of the attenuated virus and compare it to the growth curve of the wild type virus.

[0067] In certain embodiments, the attenuated virus of the invention (e.g., a recombinant or chimeric hSARS) cannot replicate in human cells as well as the wild type virus (e.g., wild type hSARS) does. However, the attenuated virus can replicate well in a cell line that lacks interferon functions, such as Vero cells.

[0068] In other embodiments, the attenuated virus of the invention is capable of infecting a host, of replicating in the host, and of causing proteins of the virus of the invention to be inserted into the cytoplasmic membrane, but the attenuated virus does not cause the host to produce new infectious viral particles. In certain embodiments, the attenuated virus infects the host, replicates in the host, and causes viral proteins to be inserted in the cytoplasmic membrane of the host with the same efficiency as the wild type hSARS. In other embodiments, the ability of the attenuated virus to cause viral proteins to be inserted into the cytoplasmic membrane into the host cell is reduced compared to the wild type virus. In certain embodiments, the ability of the attenuated hSARS virus to replicate in the host is reduced compared to the wild type virus. Any technique known to the skilled artisan can be used to determine whether a virus is capable of infecting a mammalian cell, of replicating within the host, and of causing viral proteins to be inserted into the cytoplasmic membrane of the host.

[0069] In certain embodiments, the attenuated virus of the invention is capable of infecting a host. In contrast to the wild type hSARS, however, the attenuated hSARS cannot be replicated in the host. In a specific embodiment, the attenuated hSARS virus can infect a host and can cause the host to insert viral proteins in its cytoplasmic membranes, but the attenuated virus is incapable of being replicated in the host. Any method known to the skilled artisan can be used to test whether the attenuated hSARS has infected the host and has caused the host to insert viral proteins in its cytoplasmic membranes.

[0070] In certain embodiments, the ability of the attenuated virus to infect a host is reduced compared to the ability of the wild type virus to infect the same host. Any technique known to the skilled artisan can be used to determine whether a virus is capable of infecting a host.

[0071] In certain embodiments, mutations (e.g., missense mutations) are introduced into the genome of the virus, for example, into the sequence of SEQ ID NO:1, 11, 13, or 15, or to generate a virus with an attenuated phenotype. Mutations (e.g., missense mutations) can be introduced into the structural gene and/or regulatory genes of the hSARS. Mutations can be additions, substitutions, deletions, or combinations thereof. Such variant of hSARS can be screened for a predicted functionality, such as infectivity, replication ability, protein synthesis ability, assembling ability, as well as cytopathic effect in cell cultures. In a specific embodiment, the missense mutation is a cold-sensitive mutation. In another embodiment, the missense mutation prevents a normal processing or cleavage of the viral proteins.

[0072] In other embodiments, deletions are introduced into the genome of the hSARS virus, which result in the attenuation of the virus.

[0073] In certain embodiments, attenuation of the virus is achieved by replacing a gene of the wild type virus with a gene of a virus of a different species, of a different subgroup, or of a different variant. In another aspect, attenuation of the virus is achieved by replacing one or more specific domains of a protein of the wild type virus with domains derived from the corresponding protein of a virus of a different species. In certain other embodiments, attenuation of the virus is achieved by deleting one or more specific domains of a protein of the wild type virus.

[0074] When a live attenuated vaccine is used, its safety must also be considered. The vaccine must not cause disease. Any techniques known in the art that can make a vaccine safe may be used in the present invention. In addition to attenuation techniques, other techniques may be used. One non-limiting example is to use a soluble heterologous gene that cannot be incorporated into the virion membrane. For example, a single copy of the soluble version of a viral transmembrane protein lacking the transmembrane and cytosolic domains thereof, can be used.

[0075] Various assays can be used to test the safety of a vaccine. For example, sucrose gradients and neutralization assays can be used to test the safety. A sucrose gradient assay can be used to determine whether a heterologous protein is inserted in a virion. If the heterologous protein is inserted in the virion, the virion should be tested for its ability to cause symptoms in an appropriate animal model since the virus may have acquired new, possibly pathological, properties.

5.4 Adjuvants and Carrier Molecules

[0076] hSARS-associated antigens are administered with one or more adjuvants. In one embodiment, the hSARS-associated antigen is administered together with a mineral salt adjuvants or mineral salt gel adjuvant. Such mineral salt and mineral salt gel adjuvants include, but are not limited to, aluminum hydroxide (ALUM/HYDROGEL, REHYDRAGEL), aluminum phosphate gel, aluminum hydroxyphosphate (ADJU-PHOS), and calcium phosphate.

[0077] In another embodiment, hSARS-associated antigen is administered with an immunostimulatory adjuvant. Such class of adjuvants, include, but are not limited to, cytokines (e.g., interleukin-2, interleukin-7, interleukin-12, granulocyte-macrophage colony stimulating factor (GM-CSF), interferon-y interleukin-1β (IL-1β), and IL-1β peptide or Sclavo Peptide), cytokine-containing liposomes, triplexoid glyco-sides or saponins (e.g., Quil-A and QS-21, also sold under the trademark STIMULON, ISOCREPEP), Muramyl Dipeptide (MDP) derivatives, such as N-acetyl-muramyl-L-threonyl-D-isoglutamine (Threonyl-MDP, sold under the trademark THERMUTIDE), GMDP, N-acetyl-nor-muramyl-L-alanyl-D-isoglutamate, N-acetyl muramyl-L-alanyl-D-isoglutaminyl-L-alanine-2′-β-dipalmitoyl-sn-glycero-3 hydroxy phosphorylcholesterol, muramyl tripeptide phosphatidylethanolamine (MTP-PE), unmethylated Cpg dinucleo-tides and oligonucleotides, such as bacterial DNA and fragments thereof, LPS, monophosphorylipid A (3D-MLA) sold under the trademark MPI), and polyphosphazenes.

[0078] In another embodiment, the adjuvant used is a particular adjuvant, including, but not limited to, emulsions, e.g., Freund's Complete Adjuvant, Freund's Incomplete Adjuvant, squalene or squalane oil-in-water adjuvant formulations, such as SAF and MF59, e.g., prepared with block-copolymers, such as L-121 (polyoxypropylene/polyoxyethylene) sold under the trademark PLURONIC L-121, Liposomes, Virosomes, coacervates, and immune stimulating complex, which is sold under the trademark ISCOM.
In another embodiment, a microparticulate adjuvant is used. Microparticulate adjuvants include, but are not limited to biodegradable and biocompatible polyesters, homopolymers of lactide (PLA) and glycolic acid (PGA), polylactide-co-glycolides (PLGA) microparticles, polymers that self-assemble into particulates (poloxamer particles), soluble polymers (polysphosphazenes), and virus-like particles (VLPs) such as recombinant protein particulates, e.g., hepatitis B surface antigen (HbsAg).

Yet another class of adjuvants that may be used include mucosal adjuvants, including but not limited to heat-labile enterotoxin from Escherichia coli (LT) and cholera holotoxin (CT) and cholera Toxin B Subunit (CTB) from Vibrio cholerae, mutant toxins (e.g., LTK63 and LTK72), microparticles, and polymerized liposomes.

In other embodiments, any of the above classes of adjuvants may be used in combination with each other or with other adjuvants. For example, non-limiting examples of combination adjuvant preparations that can be used to administer the hSARS-associated antigens of the invention include liposomes containing immunomodulatory protein, cytokines, T-cell and/or B-cell peptides, or microbes with or without entrapped IL-2 or microparticles containing enterotoxin. Other adjuvants known in the art are also included within the scope of the invention (see Vaccine Design: The Subunit and Adjuvant Approach, Chap. 7, Michael F. Powell and Mark J. Newman (eds.), Plenum Press, New York, 1995, which is incorporated herein in its entirety).

The effectiveness of an adjuvant may be determined by measuring the induction of antibodies directed against an immunogenic polypeptide containing a hSARS polypeptide epitope, the antibodies resulting from administration of this polypeptide in vaccines which are also comprised of the various adjuvants.

The polypeptides may be formulated into the vaccine as neutral or salt forms. Pharmacologically acceptable salts include the acid addition salts (formed with free amino groups of the peptide) and which are formed with inorganic acids, such as, for example, hydrochloric or phosphoric acids, or organic acids such as acetic, oxalic, tartaric, maleic, and the like. Salts formed with free carboxyl groups may also be derived from inorganic bases, such as, for example, sodium potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, propanol and the like.

The vaccines of the invention may be multivalent or univalent. Multivalent vaccines are made from recombinant viruses that direct the expression of more than one antigen.

Many methods may be used to introduce the vaccine formulations of the invention:

these include but are not limited to oral, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal routes, and via scarification (scatching through the top layers of skin, e.g., using a bifurcated needle).

The patient to which the vaccine is administered is preferably a mammal, most preferably a human, but can also be a non-human animal including but not limited to cows, horses, sheep, pigs, fowl (e.g., chickens), goats, cats, dogs, hamsters, mice and rats.

5.5 Preparation of Antibodies

Antibodies which specifically recognize a polypeptide of the invention, such as, but not limited to, polypeptides comprising the sequence of SEQ ID NO: 2, 12, and 14, and polypeptides as shown in FIGS. 11 (SEQ ID NOS: 17-239, 241-736 and 738-1107) and 12 (SEQ ID NOS: 1109-1589, 1591-1964 and 1966-2470), or hSARS epitope or antigen-binding fragments thereof can be used for detecting, screening, and isolating the polypeptide of the invention or fragments thereof, or similar sequences that might encode similar enzymes from the other organisms. For example, in one specific embodiment, an antibody which immunospecifically binds hSARS epitope, or a fragment thereof, can be used for various in vitro detection assays, including enzyme-linked immunosorbent assays (ELISA), radioimmunoassays, Western blot, etc., for the detection of a polypeptide of the invention or, preferably, hSARS, in samples, for example, a biological material, including cells, cell culture media (e.g., bacterial cell culture media, mammalian cell culture media, insect cell culture media, yeast cell culture media, etc.), blood, plasma, serum, tissues, sputum, nasopharyngeal aspirates, etc.

Antibodies specific for a polypeptide of the invention or any epitope of hSARS may be generated by any suitable method known in the art. Polyclonal antibodies to an antigen-of-interest, for example, the hSARS virus from deposit no. CCTCC-V200303, or comprises a nucleotide sequence of SEQ ID NO: 15, can be produced by various procedures well known in the art. For example, an antigen can be administered to various host animals including, but not limited to, rabbits, mice, rats, etc., to induce the production of antisera containing polyclonal antibodies specific for the antigen. Various adjuvants may be used to increase the immunological response, depending on the host species, and include but are not limited to, Freund’s (complete and incomplete) adjuvant, mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyls, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful adjuvants for humans such as BCG (Bacille Calmette-Guerin) and Corynebacterium parvum. Such adjuvants are also well known in the art.

Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow et al., Antibodies: A Laboratory Manual, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988); Hammerling, et al., in: Monoclonal Antibodies and T-Cell Hybridomas, pp. 563-681 (Elsevier, N.Y., 1981) (both of which are incorporated by reference in their entirety). The term “monoclonal antibody” as used herein is not limited to antibodies produced through hybridoma technology. The term “monoclonal antibody” refers to an antibody that is derived from a single clone, including any eukaryotic, prokaryotic, or phage clone, and not the method by which it is produced.

Methods for producing and screening for specific antibodies using hybridoma technology are known well known in the art. In a non-limiting example, mice can be immunized with an antigen of interest or a cell expressing such an antigen. Once an immune response is detected, e.g., antibodies specific for the antigen are detected in the mouse serum, the mouse spleen is harvested and splenocytes isolated. The splenocytes are then fused by well known techniques to any suitable myeloma cells. Hybridomas are selected and cloned by limiting dilution. The hybridoma
clones are then assayed by methods known in the art for cells that secrete antibodies capable of binding the antigen. Ascites fluid, which generally contains high levels of antibodies, can be generated by inoculating mice intraperitoneally with positive hybridoma clones.

[0092] Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, Fab and F(ab)\textsubscript{2} fragments may be produced by proteolytic cleavage of immunoglobulin molecules, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab)\textsubscript{2} fragments). F(ab)\textsubscript{2} fragments contain the complete light chain, and the variable region, the CH1 region and the hinge region of the heavy chain.

[0093] The antibodies of the invention or fragments thereof can be also produced by any method known in the art for the synthesis of antibodies, in particular, by chemical synthesis or preferably, by recombinant expression techniques.

[0094] The nucleotide sequence encoding an antibody may be obtained from any information available to those skilled in the art (i.e., from Genbank, the literature, or by routine cloning and sequencing analysis). If a clone containing a nucleic acid encoding a particular antibody or an epitope-binding fragment thereof is not available, but the sequence of the antibody molecule or epitope-binding fragment thereof is known, a nucleic acid encoding the immunoglobulin may be chemically synthesized or obtained from a suitable source (e.g., an antibody cDNA library, or a cDNA library generated from, or nucleic acid, preferably poly A+ RNA, isolated from any tissue or cell expressing the antibody, such as hybridoma cells selected to express an antibody) by PCR amplification using synthetic primers hybridizable to the 3' and 5' ends of the sequence or by cloning using an oligonucleotide probe specific for the particular gene sequence to identify, e.g., a cDNA clone from a cDNA library that encodes the antibody. Amplified nucleic acids generated by PCR may then be cloned into replicable cloning vectors using any method well known in the art.

[0095] Once the nucleotide sequence of the antibody is determined, the nucleotide sequence of the antibody may be manipulated using methods well known in the art for the manipulation of nucleotide sequences, e.g., recombinant DNA techniques, site-directed mutagenesis, PCR, etc. (see, for example, the techniques described in Sambrook et al., supra; and Ausubel et al., eds., 1998, Current Protocols in Molecular Biology, John Wiley & Sons, NY, which are both incorporated by reference herein in their entirety), to generate antibodies having a different amino acid sequence by, for example, introducing amino acid substitutions, deletions, and/or insertions into the epitope-binding domain regions of the antibodies or any portion of antibodies which may enhance or reduce biological activities of the antibodies.

[0096] Recombinant expression of an antibody requires construction of an expression vector containing a nucleotide sequence that encodes the antibody. Once a nucleotide sequence encoding an antibody molecule or a heavy or light chain of an antibody, or portion thereof has been obtained, the vector for the production of the antibody molecule may be produced by recombinant DNA technology using techniques well known in the art as discussed in the previous sections. Methods which are well known to those skilled in the art can be used to construct expression vectors containing antibody coding sequences and appropriate transcriptional and translational control signals. These methods include, for example, in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. The nucleotide sequence encoding the heavy-chain variable region, light-chain variable region, both the heavy-chain and light-chain variable regions, an epitope-binding fragment of the heavy- and/or light-chain variable region, or one or more complementarity determining regions (CDRs) of an antibody may be cloned into such a vector for expression. Thus-prepared expression vector can be then introduced into appropriate host cells for the expression of the antibody. Accordingly, the invention includes host cells containing a polynucleotide encoding an antibody specific for the polypeptides of the invention or fragments thereof.

[0097] The host cell may be co-transfected with two expression vectors of the invention, the first vector encoding a heavy chain derived polypeptide and the second vector encoding a light chain derived polypeptide. The two vectors may contain identical selectable markers which enable equal expression of heavy and light chain polypeptides or different selectable markers to ensure maintenance of both plasmids. Alternatively, a single vector may be used which encodes, and is capable of expressing, both heavy and light chain polypeptides. In such situations, the light chain should be placed before the heavy chain to avoid an excess of toxic free heavy chain (Pandolf, Nature, 322:52, 1986; and Kohler, Proc. Natl. Acad. Sci. USA, 77:2197, 1980). The coding sequences for the heavy and light chains may comprise cDNA or genomic DNA.

[0098] In another embodiment, antibodies can also be generated using various plaque display methods known in the art. In plaque display methods, functional antibody domains are displayed on the surface of plaque particles which carry the polynucleotide sequences encoding them. In a particular embodiment, such plaque can be utilized to display antigen binding domains, such as Fab and Fv or disulfide-bond stabilized Fv, expressed from a repertoire or combinatorial antibody library (e.g., human or murine). Phage expressing an antigen binding domain that binds the antigen of interest can be selected or identified with antigen, e.g., using labeled antigen or antigen bound or captured to a solid surface or bead. Phage used in these methods are typically filamentous phage, including fd and M13. The antigen binding domains are expressed as a recombinantly fused protein on either the plaque gene III or gene VIII protein. Examples of plaque display methods that can be used to make the immunoglobulins, or fragments thereof, of the present invention include those disclosed in Brinkman et al., J. Immunol. Methods, 182:41-50, 1995; Ames et al., J. Immunol. Methods, 184:177-186, 1995; Kettleborough et al., Eur. J. Immunol., 24:952-958, 1994; Persie et al., Gene, 187:9-18, 1997; Burton et al., Advances in Immunology, 57:191-280, 1994; PCT application No. PCT/GB91/01374; PCT application No. PCT/GB91/0134; PCT publications WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20041; and U.S. Pat. Nos. 5,698,426; 5,223,403; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,608; 5,427,908; 5,516,637; 5,780,225; 5,658,727; 5,733,743 and 5,969,108; each of which is incorporated herein by reference in its entirety.

[0099] As described in the above references, after plaque selection, the antibody coding regions from the plaque can be isolated and used to generate whole antibodies, including human antibodies, or any other desired fragments, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, e.g., as described in detail below. For example, techniques to recombinantly
produce Fab', Fab" and F(ab')2 fragments can also be employed using methods known in the art such as those disclosed in PCT publication WO 92/23324; Mullinax et al., BioTechniques, 12(6):864-869, 1992; and Sawai et al., AJRI, 34:26-34, 1995; and Better et al., Science, 240:1041-1043, 1988 (each of which is incorporated by reference in its entirety). Examples of techniques which can be used to produce single-chain Fv's and antibodies include those described in U.S. Pat. Nos. 4,946,778 and 5,258,498; Huston et al., Methods in Enzymology, 203:46-88, 1991; Shu et al., PNAS, 90:7995-7999, 1993; and Skerra et al., Science, 240:1038-1040, 1988.

[0100] Once an antibody molecule of the invention has been produced by any methods described above, it may then be purified by any method known in the art for purification of an immunoglobulin molecule, for example, by chromatography (e.g., ion exchange, affinity, particularly by affinity for the specific antigen after Protein A or Protein G purification, and sizing column chromatography), centrifugation, differential solubility, or by any other standard techniques for the purification of proteins. Further, the antibodies of the present invention or fragments thereof may be fused to heterologous polypeptide sequences described herein or otherwise known in the art to facilitate purification.

[0101] For some uses, including in vivo use of antibodies in humans and in vitro detection assays, it may be preferable to use chimeric, humanized, or human antibodies. A chimeric antibody is a molecule in which different portions of the antibody are derived from different animal species, such as antibodies having a variable region derived from a murine monoclonal antibody and a constant region derived from a human immunoglobulin. Methods for producing chimeric antibodies are known in the art. See, e.g., Morrison, Science, 229:1202, 1985; Oi et al., BioTechniques, 4:214 1986; Gillies et al., J. Immunol., Methods, 125:191-202, 1989; U.S. Pat. Nos. 5,807,715; 4,816,567; and 4,816,397, which are incorporated herein by reference in their entirety. Humanized antibodies are antibody molecules from non-human species that bind the desired antigen having one or more complementarity determining regions (CDRs) from the non-human species and framework regions from a human immunoglobulin molecule. Often, framework residues in the human framework regions will be substituted with the corresponding residue from the CDR donor antibody to alter, preferably improve, antigen binding. These framework substitutions are identified by methods well known in the art, e.g., by modeling the interactions of the CDR and framework residues to identify framework residues important for antigen binding and sequence comparison to identify unusual framework residues at particular positions. See, e.g., Queen et al., U.S. Pat. No. 5,585,089; Riechmann et al., Nature, 332:323, 1988, which are incorporated herein by reference in their entirety. Antibodies can be humanized using a variety of techniques known in the art including, for example, CDR-grafting (EP 239,400; PCT publication WO 91/00967; U.S. Pat. Nos. 5,225,539; 5,530,101 and 5,585,089), veneering or resurfacing (EP 592,106; EP 519,596; Padian, Molecular Immunol., 28(4/5):489-498, 1991; Studnicka et al., Protein Engineering, 7(6):805-814, 1994; Roguska et al., Proc Natl. Acad. Sci. USA, 91:960-973, 1994), and chain shuffling (U.S. Pat. No. 5,565,332), all of which are hereby incorporated by reference in their entirety.

[0102] Completely human antibodies are particularly desirable for therapeutic treatment of human patients. Human antibodies can be made by a variety of methods known in the art including phage display methods described above using antibody libraries derived from human immunoglobulin sequences. See U.S. Pat. Nos. 4,444,887 and 4,716,111; and PCT publications WO 98/46645; WO 98/50433; WO 98/24893; WO 98/16654; WO 96/34096; WO 96/33735; and WO 91/10741, each of which is incorporated herein by reference in its entirety.

[0103] Human antibodies can also be produced using transgenic mice which are incapable of expressing functional endogenous immunoglobulins, but which can express human immunoglobulin genes. For an overview of this technology for producing human antibodies, see Lonberg and Huszar, Int. Rev. Immunol., 13:65-93, 1995. For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, e.g., PCT publications WO 98/24893; WO 92/01047; WO 96/34096; WO 96/33735; European Patent No. 0 598 877; U.S. Pat. Nos. 5,413,923; 5,625,126; 5,633,425; 5,569,825; 5,661,016; 5,545,806; 5,814,318; 5,885,793; 5,916,771; and 5,939,598, which are incorporated by reference herein in their entirety. In addition, companies such as Abgenix, Inc. (Fremont, Calif.), Medarex (N.J.) and Genpharm (San Jose, Calif.) can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above.

[0104] Completely human antibodies which recognize a selected epitope can be generated using a technique referred to as “guided selection.” In this approach a selected non-human monoclonal antibody, e.g., a mouse antibody, is used to guide the selection of a completely human antibody recognizing the same epitope. (Jespers et al., Bio/technology, 12:899-903, 1988).

[0105] Antibodies fused or conjugated to heterologous polypeptides may be used in in vitro immunoassays and in purification methods (e.g., affinity chromatography) well known in the art. See, e.g., PCT publication Number WO 93/21232; EP 439,005; Nummuru et al., Immunol. Lett., 39:91-99, 1994; U.S. Pat. No. 5,474,981; Gillies et al., PNAS, 89:1428-1432, 1992; and Fell et al., J. Immunol., 146:2446-2452, 1991, which are incorporated herein by reference in their entirety.

[0106] Antibodies may also be attached to solid supports, which are particularly useful for immunoassays or purification of the polypeptides of the invention or fragments, derivatives, analogs, or variants thereof, or similar molecules having the similar enzymatic activities as the polypeptide of the invention. Such solid supports include, but are not limited to, glass, cellulose, polyacrylamide, nylon, polystyrene, polyvinyl chloride or polypropylene.

5.6 Pharmaceutical Compositions and Kits

[0107] The present invention encompasses pharmaceutical compositions comprising anti-viral agents of the present invention. In a specific embodiment, the anti-viral agent is an antibody which immunospecifically binds and neutralizes the hSARS virus or variants thereof, or any proteins derived therefrom (see Section 5.5). In another specific embodiment, the anti-viral agent is a polypeptide or nucleic acid molecule of the invention (see, for example, Sections 5.1 and 5.2). The pharmaceutical compositions have utility as an anti-viral prophylactic agent and may be administered to a subject where the subject has been exposed or is expected to be exposed to a virus.
Various delivery systems are known and can be used to administer the pharmaceutical composition of the invention, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the mutant viruses, receptor mediated endocytosis (see, e.g., Wu and Wu, 1987, J. Biol. Chem. 262:4429 4432). Methods of introduction include but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidermal, and oral routes. The compounds may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local. In a preferred embodiment, it may be desirable to introduce the pharmaceutical compositions of the invention into the lungs by any suitable route. Pulmonary administration can also be employed, e.g., by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

In a specific embodiment, it may be desirable to administer the pharmaceutical compositions of the invention locally to the area in need of treatment; this may be achieved by, for example, and not by way of limitation, local infusion during surgery, topical application, e.g., in conjunction with a wound dressing after surgery, by injection, by means of a catheter, by means of a suppository, by means of nasal spray, or by means of an implant, said implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, or fibers. In one embodiment, administration can be by direct injection at the site (or former site) infected tissues.

In another embodiment, the pharmaceutical composition can be delivered in a vesicle, in particular a liposome (see Langer, 1990, Science 249:1527-1533; Treat et al., 1990, in Liposomes in the Therapy of Infectious Disease and Cancer, Lopez Berestein and Fidler (eds.), Liss, New York, pp. 353-365 (1989); Lopez-Berestein, ibid., pp. 317-327; see generally ibid.).

In yet another embodiment, the pharmaceutical composition can be delivered in a controlled release system. In one embodiment, a pump may be used (see Langer, supra; Selkon, 1987, CRC Crit. Ref. Biomed. Eng. 14:201; Buchwald et al., 1980, Surgery 88:507; and Sandek et al., 1989, N. Engl. J. Med. 321:574). In another embodiment, polymeric materials can be used (see Medical Applications of Controlled Release, Langer and Wise (eds.), CRC Press, Boca Raton, Fla. (1974); Controlled Drug Bioavailability, Drug Product Design and Performance, Smolen and Ball (eds.), Wiley, New York (1984); Ranger and Peppas, J. Macromol. Sci. Rev. Macromol. Chem. 23:61 (1983); see also Levy et al., 1985, Science 228:190; During et al., 1989, Ann. Neurol. 25:351; Howard et al., 1989, J. Neurosurg. 71:105). In yet another embodiment, a controlled release system can be placed in proximity of the composition's target, i.e., the lung, thus requiring only a fraction of the systemic dose (e.g., Goodson, in Medical Applications of Controlled Release, supra, vol. 2, pp. 115-138 (1984)).

Other controlled release systems are discussed in the review by Langer (Science 249:1527-1533 (1990)).

The pharmaceutical compositions of the present invention comprise a therapeutically effective amount of an live attenuated, inactivated or killed HSARS virus, or recombinant or chimeric hSARS virus, and a pharmaceutically acceptable carrier. In a specific embodiment, the term “pharmaceutically acceptable” means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly in humans. The term “carrier” refers to a diluent, adjuvant, excipient, or vehicle with which the pharmaceutical composition is administered. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. Water is a preferred carrier when the pharmaceutical composition is administered intravenously. Saline solutions and aqueous dextrose and glycerol solutions can also be employed as liquid carriers, particularly for injectable solutions. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions, emulsions, tablets, pills, capsules, powders, sustained release formulations and the like. The composition can be formulated as a suppository, with traditional binders and carriers such as triglycerides. Oral formulation can include standard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, etc. Examples of suitable pharmaceutical carriers are described in “Remington’s Pharmaceutical Sciences” by E. W. Martin. The formulation should suit the mode of administration.

In a preferred embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for intravenous administration to human beings. Typically, compositions for intravenous administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocaine to ease pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the composition is to be administered by infusion, it can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

The pharmaceutical compositions of the invention can be formulated as neutral or salt forms. Pharmacologically acceptable salts include those formed with free amino groups such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with free carboxyl groups such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

The amount of the pharmaceutical composition of the invention which will be effective in the treatment of a particular disorder or condition will depend on the nature of the disorder or condition, and can be determined by standard clinical techniques. In addition, in vitro assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed in the formulation will also
depend on the route of administration, and the seriousness of the disease or disorder, and should be decided according to the judgment of the practitioner and each patient’s circumstances. However, suitable dosage ranges for intravenous administration are generally about 20 500 micrograms of active compound per kilogram body weight. Suitable dosage ranges for intranasal administration are generally about 0.01 pg/kg body weight to 1 mg/kg body weight. Effective doses may be extrapolated from dose response curves derived from in vitro or animal model test systems.

[0117] Suppositories generally contain active ingredient in the range of 0.5% to 10% by weight; oral formulations preferably contain 10% to 95% active ingredient.

[0118] The invention also provides a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions of the invention. Optionally associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of manufacture, use or sale for human administration. In a preferred embodiment, the kits contain an anti-viral agent of the invention, e.g., a polypeptide specific for the peptides encoded by a nucleotide sequence of SEQ ID NO:1, 11, 15, or 15, or as shown in FIGS. 1-1 (SEQ ID Nos. 17-239, 241-736 and 738-1107) and 12 (SEQ ID Nos: 1109-1589, 1591-1964 and 1966-2470), or any hSARS epitope, or a polypeptide or protein of the present invention, or a nucleic acid molecule of the invention, alone or in combination with adjuvants, antivirals, antibiotics, analgesics, bronchodilators, or other pharmaceutically acceptable excipients.

[0119] The present invention further encompasses kits comprising a container containing a pharmaceutical composition of the present invention and instructions for use.

5.7 Detection Assays

[0120] The present invention provides a method for detecting an antibody, which immunospecifically binds to the hSARS virus, in a biological sample, for example blood, serum, plasma, saliva, urine, etc., from a patient suffering from SARS. In a specific embodiment, the method comprising contacting the sample with the hSARS virus, for example, of deposit no. CCTCC-V200305, or having a genomic nucleic acid sequence of SEQ ID NO:15, directly immobilized on a substrate and detecting the virus-bound antibody directly or indirectly by a labeled heterologous anti-isotype antibody. In another specific embodiment, the sample is contacted with a host cell which is infected by the hSARS virus, for example, of deposit no. CCTCC-V200305, or having a genomic nucleic acid sequence of SEQ ID NO:15, and the bound antibody can be detected by immunofluorescent assay as described in Section 6.5, infra.

[0121] An exemplary method for detecting the presence or absence of a polypeptide or nucleic acid of the invention in a biological sample involves obtaining a biological sample from a patient suffering from SARS. In a specific embodiment, the sample is contacted with a compound or an agent capable of detecting an epitope or nucleic acid (e.g., mRNA, genomic DNA) of the hSARS virus such that the presence of the hSARS virus is detected in the sample. A preferred agent for detecting hSARS mRNA or genomic RNA of the invention is a labeled nucleic acid probe capable of hybridizing to mRNA or genomic RNA encoding the polypeptide of the invention. The nucleic acid probe can be, for example, a nucleic acid molecule comprising or consisting of the nucleotide sequence of SEQ ID NO:1, 11, 13, or 15, or a portion thereof, such as an oligonucleotide of at least 15, 20, 25, 30, 50, 100, 250, 500, 750, 1,000 or more contiguous nucleotides in length and sufficient to specifically hybridize under stringent conditions to a hSARS mRNA or genomic RNA.

[0122] In another preferred specific embodiment, the presence of hSARS virus is detected in the sample by an reverse transcription polymerase chain reaction (RT-PCR) using the primers that are constructed based on a partial nucleotide sequence of the genome of hSARS virus, for example, that of deposit accession no. CCTCC-V200305, or having a genomic nucleic acid sequence of SEQ ID NO:15, or based on a nucleotide sequence of SEQ ID NO:1, 11, 13, or 15. In a non-limiting specific embodiment, preferred primers to be used in a RT-PCR method are: 5'-TACACACCTCACGC-3' (SEQ ID NO:3) and 5'-CAGGACGTGAGCTAAT-3' (SEQ ID NO:4), in the presence of 2.5 mM MgCl₂ and the thermal cycles are, for example, but not limited to, 94° C. for 8 min followed by 40 cycles of 94° C. for 1 min, 50° C. for 1 min, 72° C. for 1 min (also see Section 6.7, infra). In more preferred specific embodiment, the present invention provides a real-time quantitative PCR assay to detect the presence of hSARS virus in a biological sample by subjecting the cDNA obtained by reverse transcription of the extracted total RNA from the sample to PCR reactions using the specific primers, such as those having nucleotide sequences of SEQ ID NO:3 and 4, and a fluorescence dye, such as SYBR® Green I, which fluoresces when bound non-specifically to double-stranded DNA. The fluorescence signals from these reactions are captured at the end of extension steps as PCR product is generated over a range of the thermal cycles, thereby allowing the quantitative determination of the viral load in the sample based on an amplification plot (see Section 6.7, infra).

[0123] A preferred agent for detecting hSARS is an antibody that specifically binds a polypeptide of the invention or any hSARS epitope, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab)2) can be used.

[0124] The term “labeled”, with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The detection method of the invention can be used to detect mRNA, protein (or any epitope), or genomic RNA in a sample in vitro as well as in vivo. For example, in vitro techniques for detection of mRNA include northern hybridizations, in situ hybridizations, RT-PCR, and RNase protection. In vitro techniques for detection of an epitope of hSARS include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. In vitro techniques for detection of genomic RNA include northern hybridizations, RT-PCR, and RNase protection. Furthermore, in vivo techniques for detection of hSARS include introducing into a subject organism a labeled antibody directed against the polypeptide.
example, the antibody can be labeled with a radioactive marker whose presence and location in the subject organism can be detected by standard imaging techniques, including autoradiography.

[0125] In a specific embodiment, the methods further involve obtaining a control sample from a control subject, contacting the control sample with a compound or agent capable of detecting hSARS, e.g., a polypeptide of the invention or mRNA or genomic RNA encoding a polypeptide of the invention, such that the presence of hSARS or the polypeptide or mRNA or genomic RNA encoding the polypeptide is detected in the sample, and comparing the absence of hSARS or the polypeptide or mRNA or genomic RNA encoding the polypeptide in the control sample with the presence of hSARS, or the polypeptide or mRNA or genomic DNA encoding the polypeptide in the test sample.

[0126] The invention also encompasses kits for detecting the presence of hSARS or a polypeptide or nucleic acid of the invention in a test sample. The kit, for example, can comprise a labeled compound or agent capable of detecting hSARS or the polypeptide or a nucleic acid molecule encoding the polypeptide in a test sample and, in certain embodiments, a means for determining the amount of the polypeptide or mRNA in the sample (e.g., an antibody which binds the polypeptide or an oligonucleotide probe which binds to DNA or mRNA encoding the polypeptide). Kits can also include instructions for use.

[0127] For antibody-based kits, the kit can comprise, for example: (1) a first antibody (e.g., attached to a solid support) which binds to a polypeptide of the invention or hSARS epitope; and, optionally, (2) a second, different antibody which binds to either the polypeptide or the first antibody and is conjugated to a detectable agent.

[0128] For oligonucleotide-based kits, the kit can comprise, for example: (1) an oligonucleotide, e.g., a detectably labeled oligonucleotide, which hybridizes to a nucleic acid sequence encoding a polypeptide of the invention or to a sequence within the hSARS genome or (2) a pair of primers useful for amplifying a nucleic acid molecule containing an hSARS sequence. The kit can also comprise, e.g., a buffering agent, a preservative, or a protein stabilizing agent. The kit can also comprise components necessary for detecting the detectable agent (e.g., an enzyme or a substrate). The kit can also contain a control sample or a series of control samples which can be assayed and compared to the test sample contained. Each component of the kit is usually enclosed within an individual container and all of the various containers are within a single package along with instructions for use.

5.8 Screening Assays to Identify Anti-Viral Agents

[0129] The invention provides methods for the identification of a compound that inhibits the ability of hSARS virus to infect a host or a host cell. In certain embodiments, the invention provides methods for the identification of a compound that reduces the ability of hSARS virus to replicate in a host or a host cell. Any technique well-known to the skilled artisan can be used to screen for a compound that would abolish or reduce the ability of hSARS virus to infect a host and/or to replicate in a host or a host cell.

[0130] In certain embodiments, the invention provides methods for the identification of a compound that inhibits the ability of hSARS virus to replicate in a mammal or a mammalian cell. More specifically, the invention provides methods for the identification of a compound that inhibits the ability of hSARS virus to infect a mammal or a mammalian cell. In certain embodiments, the invention provides methods for the identification of a compound that inhibits the ability of hSARS virus to replicate in a mammalian cell. In a specific embodiment, the mammalian cell is a human cell.

[0131] In another embodiment, a cell is contacted with a test compound and infected with the hSARS virus. In certain embodiments, a control culture is infected with the hSARS virus in the absence of a test compound. The cell can be contacted with a test compound before, concurrently with, or subsequent to the infection with the hSARS virus. In a specific embodiment, the cell is a mammalian cell. In an even more specific embodiment, the cell is a human cell. In certain embodiments, the cell is incubated with the test compound for at least 1 minute, at least 5 minutes at least 15 minutes, at least 30 minutes, at least 1 hour, at least 2 hours, at least 5 hours, at least 12 hours, or at least 1 day. The titer of the virus can be measured at any time during the assay. In certain embodiments, a time course of viral growth in the culture is determined. If the viral growth is inhibited or reduced in the presence of the test compound, the test compound is identified as being effective in inhibiting or reducing the growth of the hSARS virus. In a specific embodiment, the compound that inhibits or reduces the growth of the hSARS virus is tested for its ability to inhibit or reduce the growth rate of other viruses to test its specificity for the hSARS virus.

[0132] In one embodiment, a test compound is administered to a model animal and the model animal is infected with the hSARS virus. In certain embodiments, a control model animal is infected with the hSARS virus without the administration of a test compound. The test compound can be administered before, concurrently with, or subsequent to the infection with the hSARS virus. In a specific embodiment, the model animal is a mammal. In an even more specific embodiment, the model animal can be, but is not limited to, a cotton rat, a mouse, or a monkey. The titer of the virus in the model animal can be measured at any time during the assay. In certain embodiments, a time course of viral growth in the culture is determined. If the viral growth is inhibited or reduced in the presence of the test compound, the test compound is identified as being effective in inhibiting or reducing the growth or infection of the hSARS virus. In a specific embodiment, the compound that inhibits or reduces the growth of the hSARS virus in the model animal is tested for its ability to inhibit or reduce the growth rate of other viruses to test its specificity for the hSARS virus.

6. EXAMPLES

[0133] The following examples illustrate the isolation and identification of the novel hSARS virus. These examples should not be construed as limiting.

Methods and Results


6.1 Clinical Subjects

[0135] The study included all 50 patients who fitted a modified World Health Organization (WHO) definition of SARS and were admitted to 2 acute regional hospitals in Hong Kong Special Administrative Region (HK SAR) between Feb. 26 to Mar. 26, 2003 (WHO. Severe acute respiratory syndrome
(SARS) Weekly Epidemiol. Rec. 2003; 78: 81-83). A lung biopsy from an additional patient, who had typical SARS and was admitted to a third hospital, was also included in the study. Briefly, the case definition for SARS was: (i) fever of 38°C or more; (ii) cough or shortness of breath; (iii) new pulmonary infiltrates on chest radiograph; and (iv) either a history of exposure to a patient with SARS or absence of response to empirical antimicrobial coverage for typical and atypical pneumonia (beta-lactams and macrolides, fluoroquinolones or tetracyclines).

[0136] Nasopharyngeal aspirates and serum samples were collected from all patients. Paired acute and convalescent sera and feces were available from some patients. Lung biopsy tissue from one patient was processed for a viral culture, RT-PCR, routine histopathological examination, and electron microscopy. Nasopharyngeal aspirates, feces and sera submitted for microbiological investigation of other diseases were included in the study under blinding and served as controls.

[0137] The medical records were reviewed retrospectively by the attending physicians and clinical microbiologists. Routine hematological, biochemical and microbiological examinations, including bacterial culture of blood and sputum, serological study and collection of nasopharyngeal aspirates for virological tests, were carried out.

6.2 Cell Line

[0138] FRhK-4 (fetal rhesus monkey kidney) cells were maintained in minimal essential medium (MEM) with 1% fetal calf serum, 1% streptomycin and penicillin, 0.2% nystatin and 0.05% gentamycin.

6.3 Viral Infection

[0139] Two-hundred μl of clinical (nasopharyngeal aspirates) samples, from two patients (see the Result section, infra), in virus transport medium were used to infect FRhK-4 cells. The inoculated cells were incubated at 37°C for 1 hour. One ml of MEM containing 1 μg trypsin was then added to the culture and the infected cells were incubated at a 37°C incubator supplied with 5% carbon dioxide. Cytopathic effects were observed in the infected cells after 2 to 4 days of incubation. The infected cells were passaged into new FRhK-4 cells and cytopathic effects were observed within 1 day after the inoculation. The infected cells were tested by an immunofluorescent assay for influenza A, influenza B, respiratory syncytial virus, parainfluenza types 1, 2 and 3, adenovirus and human metapneumovirus (hMPV) and negative results were obtained for all cases. The infected cells were also tested by RT-PCR for influenza A and human metapneumovirus with negative results.

6.4 Virus Morphology

[0140] The infected cells prepared as described above were harvested, pelleted by centrifugation and the cell pellets were processed for thin-section transmission electron microscopic visualization. Viral particles were identified in the cells infected with both clinical specimens, but not in control cells which were not infected with the virus. Virions isolated from the infected cells were about 70-100 nanometers (FIG. 2). Viral capsids were found predominantly within the vesicles of the golgi and endoplasmic reticulum and were not free in the cytoplasm. Virus particles were also found at the cell membrane.

[0141] One virus isolate was ultracentrifuged and the cell pellet was negatively stained using phosphotungstic acid. Virus particles characteristic of Coronaviridae were thus visualized. Since the human Coronaviruses hitherto recognized are not known to cause a similar disease, the present inventors postulated that the virus isolates represent a novel virus that infects humans.

6.5 Antibody Response to the Isolated Virus

[0142] To further confirm that this novel virus is responsible for causing SARS in the infected patients, blood serum samples from the patients who were suffering from SARS were obtained and a neutralization test was performed. Typically diluted serum (×50, ×200, ×800 and ×1600) was incubated with acetone-fixed FRhK-4 cells infected with hSARS at 37°C for 45 minutes. The incubated cells were then washed with phosphate-buffered saline and stained with anti-human IgG-FITC conjugated antibody. The cells were then washed and examined under a fluorescent microscope. In these experiments, positive signals were found in 8 patients who had SARS (FIG. 3), indicating that these patients had an IgG antibody response to this novel human respiratory virus of Coronaviridae. By contrast, no signal was detected in 4 negative-control paired sera. The serum titers of anti-hSARS antibodies of the tested patients are shown in Table 1.

<table>
<thead>
<tr>
<th>Date</th>
<th>Lab No.</th>
<th>Anti-SARS</th>
</tr>
</thead>
<tbody>
<tr>
<td>25 Feb. 2003</td>
<td>S2728</td>
<td>&lt;50</td>
</tr>
<tr>
<td>6 Mar. 2003</td>
<td>S2728</td>
<td>1600</td>
</tr>
<tr>
<td>26 Feb. 2003</td>
<td>S2441</td>
<td>50</td>
</tr>
<tr>
<td>3 Mar. 2003</td>
<td>S2441</td>
<td>200</td>
</tr>
<tr>
<td>4 Mar. 2003</td>
<td>S3279</td>
<td>200</td>
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<tr>
<td>14 Mar. 2003</td>
<td>S3279</td>
<td>1600</td>
</tr>
<tr>
<td>6 Mar. 2003</td>
<td>M41043</td>
<td>&lt;50</td>
</tr>
<tr>
<td>11 Mar. 2003</td>
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<td>800</td>
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<tr>
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<td>M27124</td>
<td>&lt;50</td>
</tr>
<tr>
<td>1 Mar. 2003</td>
<td>M942268</td>
<td>&lt;50</td>
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<td>3 Mar. 2003</td>
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<td>&lt;50</td>
</tr>
<tr>
<td>7 Mar. 2003</td>
<td>KW103/2000</td>
<td>800</td>
</tr>
</tbody>
</table>

Blinded samples:

1a * Acute  <50
1b Acute  1600
2a * Acute  50
2b Convalescent  >1600
3a * Acute  <50
3b Convalescent  >1600
4a * Acute  <50
4b Convalescent  <50
5a * Acute  <50
5b Convalescent  <50
6a * Acute  <50
6b Convalescent  <50

Table 1: Titers of anti-hSARS antibodies.

[0143] These results indicated that this novel member of Coronaviridae is a key pathogen in SARS.

6.6 Sequences of the hSARS Virus

[0144] Total RNA from infected or uninfected FRhK-4 cells was harvested two days post-infection. One-hundred ng of purified RNA was reverse transcribed using Superscript® II reverse transcriptase (Invitrogen) in a 20 μl reaction mix-
ture containing 10 pg of a degenerated primer (5'-GCCGG-GAGGCTCTGAGAATCCTNNNNNNN-3'; SEQ ID NO:5; N=A, T, G or C) as recommended by the manufacturer. Reverse transcribed products were then purified by a QIAGen® PCR purification kit as instructed by the manufacturer and eluted in 30 μl of 10 mM Tris·HCl, pH 8.0. Three μl of purified cDNA products were added in a 25 μl reaction mixture containing 2.5 μl of 10x PCR buffer, 4 μl of 25 mM MgCl₂, 0.5 μl of 10 mM dNTP, 0.25 μl of AmpliTaq Gold® DNA polymerase (Applied Biosystems), 2.5 μl of [α-32P] CTP (Amer sham), 2 μl of 10 μM primer (5'-GCCGG-GAGGCTCTGAGAATCCTNNNNNNN-3'; SEQ ID NO:6). Reactions were thermal cycled through the following profile: 94°C for 5 min followed by 2 cycles of 94°C for 1 min, 40°C for 1 min, and 72°C for 2 min. This temperature profile was followed by 35 cycles of 94°C for 1 min, 60°C for 1 min, 72°C for 1 min. 6 μl of the PCR products were analyzed in a 5% denaturing polyacrylamide gel electrophoresis. Gel was exposed to X-ray film and the film was developed after an overnight exposure. Unique PCR products which were only identified in infected cell samples were isolated from the gel and eluted in a 50 μl of 1x TE buffer. Eluted PCR products were then re-amplified in 25 μl of reaction mixture containing 2.5 μl of 10x PCR buffer, 4 μl of 25 mM MgCl₂, 0.5 μl of 10 mM dNTP, 0.25 μl of AmpliTaq Gold® DNA polymerase (Applied Biosystems), 1 μl of 10 μM primer (5'-GCCGG-GAGGCTCTGAGAATCCTNNNNNNN-3'; SEQ ID NO:6). Reaction mixtures were thermal cycled through the following profile: 94°C for 8 min followed by 35 cycles of 94°C for 1 min, 60°C for 1 min, 72°C for 1 min. 5 μl of PCR products were cloned using a TOP10 TA Cloning® kit (Invitrogen) and ligated plasmids were transformed into Top10 E. coli competent cells (Invitrogen). PCR insert were sequenced by a BigDye cycle sequencing kit as recommended by the manufacturer (Applied Biosystems) and sequencing products were analyzed by an automatic sequencer (Applied Biosystems, model number 3770). The obtained sequence (SEQ ID NO:1) is shown in Fig. 1. The deduced amino acid sequence (SEQ ID NO:2) from the obtained DNA sequence showed 57% homology to the polymerase protein of identified coronaviruses. [0145] Similarly, two other partial sequences (SEQ ID NOS:11 and 13) and deduced amino acid sequences (SEQ ID NOS:12 and 14, respectively) were obtained from the hSARS virus and are shown in Figs. 8 (SEQ ID NOS:11 and 12) and 9 (SEQ ID NOS:13 and 14).

[0146] The entire genomic sequence of hSARS virus is shown in Fig. 10 (SEQ ID NO:15). The deduced amino acid sequences of SEQ ID NO:15 in all three frames are shown in Figs. 11 (nucleotide sequences shown in SEQ ID NOS:16, 240 and 737; for amino acid sequences, see SEQ ID NO:17-239, 241-736 and 738-1107). The deduced amino acid sequences of the complement of SEQ ID NO:15 in all three frames are shown in Fig. 12 (nucleotide sequences shown in SEQ NOS:1108, 1590 and 1965; for amino acid sequences, see SEQ ID NOS:1109-1589, 1591-1964 and 1966-2470).

6.7 Detection of hSARS Virus in Nasopharyngeal Aspirates

[0147] First, the nasopharyngeal aspirates (NPA) were examined by rapid immunofluorescent antigen detection for influenza A and B, parainfluenza types 1, 2 and 3, respiratory syncytial virus and adenovirus (Chan K H, Maldeis N, Pope W, Yung A, Ozinskias A, Gill J, Seto W H, Shortridge K F, Petriss J S M. Evaluation of Directigen Fly A+B test for rapid diagnosis of influenza A and B virus infections. J Clin Microbiol. 2002; 40: 1675-1680) and were cultured for conventional respiratory pathogens on Mardin Darby Canine Kidney, LLC-MK2, DLE-2 and MRC-5 cells (Wiedbrauk D L, Johnston S L. Manual of clinical virology. Raven Press, New York. 1993). Subsequently, fetal rhesus kidney (FRK-4) and A-549 cells were added to the panel of cell lines used. Reverse transcription polymerase chain reaction (RT-PCR) was performed directly on the clinical specimen for influenza A (Fouchier R A, Bestebroer T M, Herfst S, Van Der Kemp L, Rimmelzwaan G F, Osterhaus A D. Detection of influenza A virus from different species by PCR amplification of conserved sequences in the matrix gene. J Clin Microbiol. 2000; 38: 4096-4101) and human metapneumovirus (HMPV). The primers used for HMPV were: for first round, 5'-AARGT-SATGATCATCAGC-3' (SEQ ID NO. 7) and 5'-CAKATYTG-GCCTATGCCTTC-3' (SEQ ID NO:8); and nested primers: 5'-ACACCTGATCATACGACG-3' (SEQ ID NO:9) and 5'-GACCTTGGATCGCCCTAA-3' (SEQ ID NO:10). The size of the nested PCR product was 201 bp. An ELISA for mycoplasma was used to screen cell cultures (Roche Diagnostics GmbH, Roche, Indianapolis, USA).

RT-PCR Assay

[0148] Subsequent to culturing and genetic sequencing of the hSARS virus from two patients (see Section 6.6, supra), an RT-PCR was developed to detect the hSARS virus sequence from NPA samples. Total RNA from clinical samples was reverse transcribed using random hexamers and cDNA was amplified using primers 5'-TACACACCTGACGTTG-3' (SEQ ID NO:3) and 5'-ACACAACGTGACGAAAT-3' (SEQ ID NO:4), which are constructed based on the RNA-dependent RNA polymerase-encoding sequence (SEQ ID NO:1) of the hSARS virus in the presence of 2.5 mM MgCl₂ (94°C for 8 min followed by 40 cycles of 94°C for 1 min, 50°C for 1 min, and 72°C for 1 min).

[0149] The summary of a typical RT-PCR protocol is as follows:

[0150] 1. RNA Extraction

[0151] RNA from 140 μl of NPA samples is extracted by QIAqicuk viral RNA extraction kit and is eluted in 50 μl of elution buffer.

[0152] 2. Reverse Transcription

<table>
<thead>
<tr>
<th>Reaction</th>
<th>Volume</th>
</tr>
</thead>
<tbody>
<tr>
<td>RNA</td>
<td>11.5 μl</td>
</tr>
<tr>
<td>0.1 M DTT</td>
<td>2 μl</td>
</tr>
<tr>
<td>5x buffer</td>
<td>4 μl</td>
</tr>
<tr>
<td>10 mM dNTP</td>
<td>1 μl</td>
</tr>
<tr>
<td>Superscript II, 200 U/μl (Invitrogen)</td>
<td>1 μl</td>
</tr>
<tr>
<td>Random hexamers, 0.5 μM</td>
<td>0.5 μl</td>
</tr>
<tr>
<td>Reaction condition</td>
<td>42°C, 30 min</td>
</tr>
<tr>
<td>94°C, 3 min</td>
<td>4°C</td>
</tr>
</tbody>
</table>

[0153] 3. PCR

[0154] cDNA generated by random primers is amplified in a 50 μl reaction as follows:

<table>
<thead>
<tr>
<th>cDNA</th>
<th>2 μl</th>
</tr>
</thead>
<tbody>
<tr>
<td>10 mM dNTP</td>
<td>0.5 μl</td>
</tr>
<tr>
<td>10x buffer</td>
<td>5 μl</td>
</tr>
</tbody>
</table>
25 mM MgCl₂  5 μl
25 μM Forward primer  0.5 μl
25 μM Reverse primer  0.5 μl
AmpliTag Gold® polymerase, 5 U/μl (Applied Biosystems)  0.25 μl
Water  36.25 μl

[0155] Thermal-cycle condition: 95°C, 10 min, followed by 40 cycles of 95°C, 1 min; 50°C, 1 min; 72°C, 1 min.

[0156] 4. Primer Sequences

[0157] Primers were designed based on the RNA-dependent RNA polymerase encoding sequence (SEQ ID NO:1) of the hSARS virus.

Forward 5’ TAGACACCTCGCTGCTTG 3’ (SEQ ID NO:3)
Reverse 5’ GACGACGTACAT 3’ (SEQ ID NO:4)

[0158] Product size: 182 bps

[0159] Real-Time Quantitative PCR Assay

[0160] Total RNA from 140 μl of nasopharyngeal aspirate (NPA) was extracted by QIAamp® virus RNA mini kit (Qiagen) as instructed by the manufacturer. Ten μl of eluted RNA samples were reverse transcribed by 200 U of SuperScript® II reverse transcriptase (Invitrogen) in a 20 μl reaction mixture containing 0.15 μg of random hexamers, 10 mmol/L DTT, and 0.5 mmol/L dNTP, as instructed. Complementary DNA was then amplified in a SYBR® Green I fluorescence reaction (Roche) mixtures. Briefly, 20 μl reaction mixtures containing 2 μl of cDNA, 3.5 mmol/L MgCl₂, 0.25 mmol/L of forward primer (5’-TAGACACCTCGCTGCTTG-3’, SEQ ID NO:3) and 0.25 μmol/L of reverse primer (5’-GACGACGTACAT-3’, SEQ ID NO:4) were thermal-cycled by a Light-Cycler (Roche) with the PCR program, [95°C, 10 min followed by 50 cycles of 95°C, 30 sec; 72°C, 9 sec]. Plasmids containing the target sequence were used as positive controls. Fluorescence signals from these reactions were captured at the end of extension step in each cycle. (see FIG. 7A). To determine the specificity of the assay, PCR products (184 base pairs) were subjected to a melting curve analysis at the end of the assay (65°C to 95°C, 0.1°C per second; see FIG. 7B).

Clinical findings:

[0161] Clinical findings:

[0162] All 50 patients with SARS were ethnic Chinese. They represented 5 different epidemiologically linked clusters as well as additional sporadic cases fitting the case definition. They were hospitalized at a median of 5 days after the onset of symptoms. The median age was 42 years (range of 23 to 74) and the female to male ratio was 1.3. Seventeen (28%) were health care workers and five (10%) had a history of visit to a hospital experiencing a major outbreak of SARS. Thirteen (26%) patients had household contacts and 12 (24%) others had social contacts with patients with SARS. Four (8%) had a history of recent travel to mainland China.

[0163] The major complaints from most patients were fever (90%) and shortness of breath. Cough and myalgia was present in more than half the patients (Table 2). Upper respiratory tract symptoms such as rhinorrhea (24%) and sore throat (20%) were present in a minority of patients. Diarrhea (10%) and anorexia (10%) were also reported. At initial examination, auscultatory findings, such as cracking and decreased air entry, were present in only 38% of patients. Dry cough was reported by 62% of patients. All patients had radiological evidence of consolidation, at the time of admission, involving 1 zone (in 36), 2 zones (13) and 3 zones (1).

TABLE 2

<table>
<thead>
<tr>
<th>Clinical symptoms</th>
<th>Number (percentage)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fever</td>
<td>50 (100%)</td>
</tr>
<tr>
<td>Chill or rigor</td>
<td>37 (74%)</td>
</tr>
<tr>
<td>Cough</td>
<td>31 (62%)</td>
</tr>
<tr>
<td>Myalgia</td>
<td>27 (54%)</td>
</tr>
<tr>
<td>Malaise</td>
<td>25 (50%)</td>
</tr>
<tr>
<td>Running nose</td>
<td>12 (24%)</td>
</tr>
<tr>
<td>Sore throat</td>
<td>10 (20%)</td>
</tr>
<tr>
<td>Shortness of breath</td>
<td>10 (20%)</td>
</tr>
<tr>
<td>Anorexia</td>
<td>10 (20%)</td>
</tr>
<tr>
<td>Diarrhea</td>
<td>5 (10%)</td>
</tr>
<tr>
<td>Headache</td>
<td>10 (20%)</td>
</tr>
<tr>
<td>Dizziness</td>
<td>6 (12%)</td>
</tr>
</tbody>
</table>

* Truncal macropolipapular rash was noted in 1 patient.

[0164] In spite of the high fever, most patients (98%) had no evidence of a leukocytosis. Lymphopenia (68%), leukopenia (26%), thrombocytopenia (40%) and anemia (18%) were present in peripheral blood examination (Table 3). Parenchymal liver enzyme, alanine aminotransferase (ALT) and muscle enzyme, creatine kinase (CPK) were elevated in 34% and 26% respectively.

TABLE 3

<table>
<thead>
<tr>
<th>Laboratory parameter</th>
<th>Mean (range)</th>
<th>Percentage of abnormal</th>
<th>Normal range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haemoglobin</td>
<td>12.9 (8.9-15.9)</td>
<td>9 (18%)</td>
<td>11.5-16.5 g/dL</td>
</tr>
<tr>
<td>Anaemia</td>
<td>5.17 (1.1-11.4)</td>
<td>13 (26%)</td>
<td>4-11 × 10⁹/L</td>
</tr>
<tr>
<td>Leucopenia</td>
<td>0.78 (0.3-1.5)</td>
<td>34 (68%)</td>
<td>1.5-4.0 × 10⁹/L</td>
</tr>
<tr>
<td>Lymphocyte count</td>
<td>174 (98-351)</td>
<td>20 (40%)</td>
<td>150-400 x 10⁹/L</td>
</tr>
<tr>
<td>Thrombocytopenia</td>
<td>63 (31-350)</td>
<td>17 (34%)</td>
<td>150-400 x 10⁹/L</td>
</tr>
<tr>
<td>Elevated ALT</td>
<td>37 (26-50)</td>
<td>34 (68%)</td>
<td>42-54 g/L</td>
</tr>
<tr>
<td>Albumin</td>
<td>33 (21-42)</td>
<td>24 (46%)</td>
<td>24-36 g/L</td>
</tr>
<tr>
<td>Elevated globulin</td>
<td>244 (31-1379)</td>
<td>13 (26%)</td>
<td>34-138 U/L</td>
</tr>
</tbody>
</table>

[0165] Routine microbiological investigations for known viruses and bacteria by culture, antigen detection, and PCR were negative in most cases. Blood culture was positive for Escherichia coli in a 74-year-old male patient, who was admitted to intensive care unit, and was attributed to hospital acquired urinary tract infection. Klebsiella pneumoniae and Hemophilus influenzae were isolated from the sputum specimens of 2 other patients on admission.

[0166] Oral levofloxacin 500 mg q24 h was given in 9 patients and intravenous (1.2 g q8 h) oral (375 mg tid) amoxicillin-clavulanate and intravenous/oral clarithromycin 500 mg q12 h were given in another 40 patients. Four patients were given oral oseltamivir 75 mg bid. In one patient, intra-
ventous ceftriaxone 2 gm q24h, oral azithromycin 500 mg q24 h, and oral amantadine 100 mg bid were given for empirical coverage of typical and atypical pneumonia.

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[0167] Nineteen patients progressed to severe disease with oxygen desaturation and were required intensive care and ventilatory support. The mean number of days of deterioration from the onset of symptoms was 8.3 days. Intravenous ribavirin 8 mg/kg q8 h and steroid was given in 49 patients at a mean day of 6.7 after onset of symptoms.

[0168] The risk factors associated with severe complicated disease requiring intensive care and ventilatory support were older age, lymphopenia, impaired ALT, and delayed initiation of ribavirin and steroid (Table 4). All the complicated cases were treated with ribavirin and steroid after admission to the intensive care unit whereas all the uncomplicated cases were started on ribavirin and steroid in the general ward. As expected, 31 uncomplicated cases recovered or improved whereas 8 complicated cases deteriorated with one death at the time of writing. All 50 patients were monitored for a mean of 12 days at the time of writing.

10-hour household contact with a Chinese visitor who came from Guangzhou and later died from SARS. Two days after this exposure, he presented with fever, malaise, myalgia, and headache. Crepitations were present over the right lower zone and there was a corresponding alveolar shadow on the chest radiograph. Hematological investigation revealed lymphopenia of 0.7×10^9/L with normal total white cell and platelet counts. Both ALT (41 U/L) and CPK (405 U/L) were impaired. Despite a combination of oral azithromycin, amantadine, and intravenous ceftriaxone, there was increasing bilateral pulmonary infiltrates and progressive oxygen desaturation. Therefore, an open lung biopsy was performed 9 days after admission. Histopathological examination showed a mild interstitial inflammation with scattered alveolar pneumocytes showing cytomegaly, granular amorphophilic cytoplasm and enlarged nuclei with prominent nucleoli. No cells showed inclusions typical of herpesvirus or adenovirus infection. The patient required ventilation and intensive care after the operative procedure. Empirical intravenous ribavirin and hydrocortisone were given. He succumbed 20 days after

<table>
<thead>
<tr>
<th>TABLE 4</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Complicated case (n = 19)</strong></td>
</tr>
<tr>
<td>---------------------------------</td>
</tr>
<tr>
<td>Mean (SD) age (range)</td>
</tr>
<tr>
<td>Male:Female ratio</td>
</tr>
<tr>
<td>Underlying illness</td>
</tr>
<tr>
<td>Mode of contact</td>
</tr>
<tr>
<td>Travel to China</td>
</tr>
<tr>
<td>Health care worker</td>
</tr>
<tr>
<td>Hospital visit</td>
</tr>
<tr>
<td>Household contact</td>
</tr>
<tr>
<td>Social contact</td>
</tr>
<tr>
<td>Mean (SD) duration of symptoms to admission (days)</td>
</tr>
<tr>
<td>Mean (SD) admission temperature (°C)</td>
</tr>
<tr>
<td>Mean (SD) initial peripheral WBC count (×10^9/L)</td>
</tr>
<tr>
<td>Mean (SD) initial lymphocyte count (×10^12/L)</td>
</tr>
<tr>
<td>Presence of thrombocytopenia (&lt;30 ×10^9/L)</td>
</tr>
<tr>
<td>Impaired liver function test</td>
</tr>
<tr>
<td>CXR changes (number of zone affected)</td>
</tr>
<tr>
<td>Mean (SD) day of deterioration from the onset of symptoms</td>
</tr>
<tr>
<td>Mean (SD) day of initiation of Ribavirin &amp; steroid from the onset of symptoms</td>
</tr>
<tr>
<td>Initiation of ribavirin &amp; steroid after deterioration</td>
</tr>
<tr>
<td>Response to ribavirin &amp; steroid Outcome</td>
</tr>
<tr>
<td>Improved or recovered</td>
</tr>
<tr>
<td>Not improving</td>
</tr>
</tbody>
</table>

* Multi-varient analysis is not performed due to low number of cases;
  1 patients had diabetic mellitus, 1 had hypertrophic obstructive cardiomyopathy, 1 had chronic active hepatitis B, and 1 had brain tumour;
  2 patients had essential hypertension;
  3 deterioration requiring intensive care support;
  4 died.

[0169] Two virus isolates, subsequently identified as a member of Coronaviridae (see below), were isolated from two patients. One was from an open lung biopsy tissue of a 53-year-old Hong Kong Chinese resident and the other from a nasopharyngeal aspirate of a 42-year-old female with good previous health. The 53-year-old male had a history of admission. In retrospect, coronavirus-like RNA was detected in his nasopharyngeal aspirate, lung biopsy, and post-mortem lung. He had a significant rise in titer of antibodies against his own hSARS isolate from 1/200 to 1/1600.

[0170] The second patient from whom a hSARS virus was isolated, was a 42-year-old female with good past health. She
had a history of travel to Guangzhou in mainland China for 2
days. She presented with fever and diarrhea 5 days after her
return to Hong Kong. Physical examination showed crepita-
tion over the right lower zone which had a corresponding
alveolar shadow on the chest radiograph. Investigation
revealed leukocytosis (2.7×10^9/L), lymphopenia (0.6×10^9/L),
and thrombocytopenia (104×10^9/L). Despite the empirical
antimicrobial coverage with amoxicillin-clavulanate,
clarithromycin, and oseltamivir, she deteriorated 5 days after
admission and required mechanical ventilation and intensive
care for 5 days. She gradually improved without receiving
treatment with ribavirin or steroid. Her nasopharyngeal aspir-
ate was positive for the virus in the RT-PCR and she was
seroconverted from antibody titre <1:50 to 1:1600 against the
SARS isolate.

**[0171]** Virological Findings:

**[0172]** Viruses were isolated on FRhk-4 cells from the lung
biopsy and nasopharyngeal aspirate respectively, of two
patients described above. The initial cytopathic effect
appeared between 2 and 4 days after inoculation, but on
subsequent passage, cytopathic effect appeared in 24 hours.
Both virus isolates did not react with the routine panel of
reagents used to identify virus isolates including those for
influenza A, B parainfluenza types 1, 2, 3, adenovirus and
respiratory syncytial virus (DAKO, Glostrup, Denmark). They
also failed to react in RT-PCR assays for influenza A and
HMPV or in PCR assays for mycoplasma. The virus was ether
sensitive, indicating that it was an enveloped virus. Electron
microscopy of negatively stained (2% potassium phospha-
tungstate, pH 7.0) cell culture extracts obtained by ultracen-
trifugation showed the presence of pleomorphic enveloped
viral particles, of about 80-90 nm (ranging 70-130 nm) in
diameter, whose surface morphology appeared comparable to
members of Coronaviridae (FIG. 5A). Thin section electron
microscopy of infected cells revealed virus particles of
55-90 nm diameter within the smooth-walled vesicles in the
eytoplasm (FIGS. 5A and 5B). Virus particles were also seen at
the cell surface. The overall findings were compatible with in-
fecions in the cells caused by viruses of Coronaviridae.

**[0173]** A thin section electron micrograph of the lung
biopsy of the 53 year old male contained 60-90-nm viral
particles in the cytoplasm of desquamated cells. These viral
particles were similar in size and morphology to those
observed in the cell-cultured virus isolate from both patients
(FIG. 4).

**[0174]** The RT-PCR products generated in a random primer
RT-PCR assay were analyzed and unique bands found in the
virus infected specimen was cloned and sequenced. Of 30
clones examined, a clone containing 646 base pairs (SEQ ID
NO:1) of unknown origin was identified. Sequence analysis of
this DNA fragment suggested this sequence had a weak
homology to viruses of the family of Coronaviridae (data not
shown). Deduced amino acid sequence (215 amino acids:
SEQ ID NO:2) from this unknown sequence, however, had the
highest homology (57%) to the RNA polymerase of
bovine coronavirus and murine hepatitis virus, confirming
that this virus belongs to the family of Coronaviridae. Phylo-
genetetic analysis of the protein sequences showed that this
virus, though most closely related to the group II coronavi-
ruses, was a distinct virus (FIGS. 5A and 5B).

**[0175]** Based on the 646 by sequence of the isolate, specific
primers for detecting the new virus was designed for RT-PCR
detection of this SARS virus genome in clinical specimens.
Of the 44 nasopharyngeal specimens available from the 50
SARS patients, 22 had evidence of SARS RNA. Viral RNA
was detectable in 10 of 18 fecal samples tested. The specific-
ity of the RT-PCR reaction was confirmed by sequencing
selected positive RT-PCR amplified products. None of 40
nasopharyngeal and fecal specimens from patients with
unrelated diseases were reactive in the RT-PCR assay.

**[0176]** To determine the dynamic range of real-time quan-
titative PCR, serial dilutions of plasmid DNA containing the
target sequence were made and subjected to the real-time
quantitative PCR assay. As shown in FIG. 7A, the assay was
able to detect as little as 10 copies of the target sequence. By
contrast, no signal was observed in the water control (FIG. 7A).
Positive signals were observed in 23 out of 29 serologi-
cally confirmed SARS patients. In all of these positive cases,
a unique PCR product (T_m=82° C) corresponding to the
signal from the positive control was observed (FIG. 7B, and
data not shown). These results indicated this assay is highly
specific to the target. The copy numbers of the target sequence
in these reactions range from 4539 to less than 10. Thus, as
high as 6.48×10^6 copies of this viral sequence could be found
in 1 ml of NPA sample. In 5 of the above positive cases, it was
possible to collect NPA samples before seroconversion. Viral
RNA was detected in 3 of these samples, indicating that this
assay can detect the virus even at the early onset of infection.

**[0177]** To further validate the specificity of this assay, NPA
samples from healthy individuals (n=11) and patients suf-
f ered from adenovirus (n=11), respiratory syncytial virus
(n=11), human metapneumovirus (n=11), influenza A virus
(n=13) or influenza B virus (n=1) infection as negative
controls. All of these samples, except one, were
negative in the assay. The false positive case was negative in
a subsequent test. Taken together, including the initial false
positive case, the real-time quantitative PCR assay has
sensitivity of 79% and specificity of 98%.

**[0178]** Epidemiological data suggest that droplet transmis-
sion is one of the major route of transmission of this virus.
The detection of live virus and the detection of high copies of viral
sequence from NPA samples in the current study clearly
support that cough and sneeze droplets from SARS patients
might be the major source of this infectious agent. Interest-
ingly, 2 out of 4 available stool samples form the SARS
patients in this study were positive in the assay (data not
shown). The detection of the virus in feces suggests that there
might be other routes of transmission. It is relevant to note
that a number of animal coronaviruses are spread via the
focal-source route (McIntosh K., 1974, Coronaviruses: a com-
parative review. Current Top Microbiol Immunol. 63:
85-112). However, further studies are required to test whether
the virus in feces is infectious or not.

**[0179]** Currently, apart form this SARS virus, there are
two known serogroups of human coronaviruses (229E
and OC43) (Hruskova J. et al., 1990, Antibodies to human coro-
naviruses 229E and OC43 in the population of C.R., Acta
Viral. 34:346-52). The primer set used in the present assay
does not have homology to the strain 229E. Due to the lack
of available corresponding OC43 sequence in the Genbank,
it is not known whether these primers would cross-react with
this strain. However, sequence analyses of available sequences
in other regions of OC43 polymerase gene indicate that
the novel human virus associated with SARS is geneti-
cally distinct from OC43. Furthermore, the primers used in
this study do not have homology to any of sequences from
known coronaviruses. Thus, it is very unlikely that these
primers would cross-react with the strain OC43.
Apart from the novel pathogen, metapneumovirus was reported to be identified in some of SARS patients (Center for Disease Control and Prevention, 2003, *Morbidity and Mortality Weekly Report* 52: 269-272). No evidence of metapneumovirus infection was detected in any of the patients in this study (data not shown), suggesting that the novel hSARS virus of the invention is the key player in the pathogenesis of SARS.

**Immunofluorescent Antibody Detection:**

Thirty-five of the 50 most recent serum samples from patients with SARS had evidence of antibodies to the hSARS (see FIG. 3). Of 27 patients from whom paired acute and convalescent sera were available, all were seroconverted or had >4-fold increase in antibody titer to the virus. Five other pairs of sera from additional SARS patients from clusters outside this study group were also tested to provide a wider sampling of SARS patients in the community and all of them were seroconverted. None of 50 sera from patients with respiratory or other diseases as well as none of 200 normal blood donors had detectable antibody.

**When either seropositivity to HP-CV in a single serum or viral RNA detection in the NPA or stool are considered evidence of infection with the hSARS, 45 of the 50 patients had evidence of infection. Of the 5 patients without any virological evidence of Coronavirus viral infection, only one of these patients had their sera tested >14 days after onset of clinical disease.**

**Discussion**

The outbreak of SARS is unusual in a number of aspects, in particular, in the appearance of clusters of patients with pneumonia in health care workers and family contacts. In this series of patients with SARS, investigations for conventional pathogens of atypical pneumonia proved negative. However, a virus that belongs to the family Coronavirus was isolated from the lung biopsy and nasopharyngeal aspirate obtained from two SARS patients, respectively. Phylogenetically, the virus was not closely related to any known human or animal coronavirus or to porcine. The present analysis is based on a 646 by fragment (SEQ ID NO:1) of the polymerase gene and the entire genome of the isolated hSARS virus, which indicates that the virus relates to antigenic group 2 of the coronaviruses along with murine hepatitis and bovine coronavirus. However, viruses of the Coronavirus can undergo heterologous recombination within the virus family and genetic analysis of other parts of the genome needs to be carried out before the nature of this new virus is more conclusively defined (Holmes K V. Coronaviruses. Eds Knipe D M, Howley P M Fields Virology, 4th Edition, Lippincott Williams & Wilkins, Philadelphia, 1187-1203). The biological, genetic and clinical data, taken together, indicate that the new virus is not one of the two known human coronaviruses.

**Majority (90%) of patients with clinically defined SARS had either serological or RT-PCR evidence of infection by this virus. In contrast, neither antibody nor viral RNA was detectable in healthy controls. All 27 patients from whom acute and convalescent sera were available demonstrated rising antibody titers to hSARS virus, strengthening the contention that a recent infection with this virus is a necessary factor in the evolution of SARS. In addition, all five pairs of acute and convalescent sera tested from patients from other hospitals in Hong Kong also showed seroconversion to the virus. The five patients who has not shown serological or virological evidence of hSARS virus infection, need to have later convalescent sera tested to define if they are also seroconverted. However, the concordance of the hSARS virus with the clinical definition of SARS appears remarkable, given that clinical case definitions are not perfect.**

**No evidence of HMPV infection, either by RT-PCR or rising antibody titer against HMPV, was detected in any of these patients. No other pathogen was consistently detected in our group of patients with SARS. It is therefore highly likely that this hSARS virus is either the cause of SARS or a necessary pre-requisite for disease progression. Whether or not other microbial or other co-factors play a role in progression of the disease remains to be investigated.**


We describe for the first time the clinical presentation and complications of SARS. Less than 25% of patients with coronavirus pneumonia had upper respiratory tract symptoms. As expected in atypical pneumonia, both respiratory symptoms and positive auscultatory findings were very disproportional to the chest radiographic findings. Gastrointestinal symptoms were present in 10%. It is relevant that the virus RNA is detected in faeces of some patients and that coronaviruses have been associated with diarrhoea in animals and humans (Caul E O, Eggleston S J. Further studies on human enteric coronaviruses *Arch. Virol.* 1977; 54: 107-17).

The high incidence of deranged liver function test, leucopenia, significant lymphopenia, thrombocytopenia and subsequent evolution into adult respiratory distress syndrome suggests a severe systemic inflammatory damage induced by this hSARS virus. Thus immuno-modulation by steroid may be important to complement the antiviral therapy by ribavirin. In this regard, it is pertinent that severe human disease associated with the avian influenza subtype H5N1, another virus
that recently crossed from animals to humans, has also been postulated to have an immune-pathological component (Cheung C Y, Poon L M, Lau A S Y et al. Induction of proinflammatory cytokines in human macrophages by influenza A (H5N1) viruses: a mechanism for the unusual severity of human disease. *Lancet* 2002; 360: 1831-1837). In common with H5N1 disease, patients with severe SARS are adults, are significantly more lymphopenic and have parameters of organ dysfunction beyond the respiratory tract (Table 4) (Yuen K Y, Chan P K S, Peiris J S M, et al. Clinical features and rapid virologic diagnosis of human disease associated with avian influenza A H5N1 virus. *Lancet* 1998; 351: 467-471). It is important to note that a window of opportunity of around 8 days exists from the onset of symptoms to respiratory failure. Severe complicated cases are strongly associated with both underlying disease and delayed use of ribavirin and steroid therapy. Following our clinical experience in the initial cases, this combination therapy was started very early in subsequent cases which were largely uncomplicated cases at the time of admission. The overall mortality at the time of writing is only 2% with this treatment regimen. There were still 8 out of 19 complicated cases who had not shown significant response. It is not possible to a detailed analysis of the therapeutic response to this combination regimen due to the heterogeneous dosing and time of initiation of therapy.

[0190] Other factors associated with severe disease is acquisition of the disease through household contact which may be attributed to a higher dose or duration of virus exposure and the presence of underlying illnesses.

[0191] The clinical description reported here pertains largely to the more severe cases admitted to hospital. We presently have no data on the full clinical spectrum of the emerging Coronaviridae infection in the community or in an out-patient setting. The availability of diagnostic tests as described here will help address these questions. In addition, it will allow questions pertaining to the period of virus shedding (and communicability) during convalescence, the presence of virus in other body fluids and excreta and the presence of virus shedding during the incubation period, to be addressed.

[0192] The epidemiological data at present appears to indicate that the virus is spread by droplets or by direct and indirect contact although airborn spread cannot be ruled out in some instances. The finding of infectious virus in the respiratory tract supports this contention. Preliminary evidence also suggests that the virus may be shed in the feces. However, it is important to note that detection of viral RNA does not prove that the virus is viable or transmissible. If viable virus is detectable in the feces, this would be a potentially additional route of transmission that needs to be considered. It is relevant to note that a number of animal coronaviruses are spread via the fecal-oral route (Mcintosh K. Coronaviruses: a comparative review. *Current Top Microbiol Immunol.* 1974; 63: 85-112).

[0193] In conclusion, this report provides evidence that a virus in the Coronaviridae family is the etiological agent of SARS.

7. DEPOSIT

[0194] A sample of isolated hSARS virus was deposited with China Center for Type Culture Collection (CCTCC) at Wuhan University, Wuhan 430072 in China on Apr 2, 2003 in accordance with the Budapest Treaty on the Deposit of Microorganisms, and accession number CCTCC-V200303, which is incorporated herein by reference in its entirety.

8. MARKET POTENTIAL

[0195] The hSARS virus can now be grown on a large scale, which allows the development of various diagnostic tests as described hereinabove as well as the development of vaccines and antiviral agents that are effective in preventing, ameliorating or treating SARS. Given the severity of the disease and its rapid global spread, it is highly likely that significant demands for diagnostic tests, therapies and vaccines to battle against the disease, will arise on a global scale. In addition, this virus contains genetic information which is extremely important and valuable for clinical and scientific research applications.

9. EQUIVALENTS

[0196] Those skilled in the art will recognize, or be able to ascertain many equivalents to the specific embodiments of the invention described herein using no more than routine experimentation. Such equivalents are intended to be encompassed by the following claims.

[0197] All publications, patents and patent applications mentioned in this specification are herein incorporated by reference into the specification to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated herein by reference.

[0198] Citation or discussion of a reference herein shall not be construed as an admission that such is prior art to the present invention.

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**SEQUENCE LISTING**

The patent application contains a lengthy “Sequence Listing” section. A copy of the “Sequence Listing” is available in electronic form from the USPTO web site (http://seqdata.uspto.gov/?page=RequestDocDetail&DocID=US20100080824A1). An electronic copy of the “Sequence Listing” will also be available from the USPTO upon request and payment of the fee set forth in 37 CFR 1.19(b)(3).
18. An immunogenic formulation comprising an immunogenically effective amount of 1) a protein extract of an isolated hSARS virus having China Center for Type Culture Collection Deposit Accession No. CCTCC-V200303, or 2) an isolated hSARS virus comprising a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 11, 13 or 15 or a subunit thereof, and a pharmaceutically acceptable carrier.

19-23. (canceled)

24. A kit comprising a container containing the immunogenic formulation of claim 18.

25-39. (canceled)