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PIV Characterisation of Flocculation Dynamics and Floc Structure in Water Treatment

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Running Head: Particle flocculation and coagulation, Water treatment

1 **Abstract**

2 Particle flocculation with chemical flocculant addition is an essential step in water treatment.
3 The performance of flocculation and the property of the flocs formed affect the overall results
4 of the treatment process. In addition to particulate impurities, the presence of organic matter
5 in water, such as natural organic materials (NOM), also influence the effectiveness of
6 chemical flocculation. In this paper, the PIV system was employed to investigate the
7 flocculation dynamics for different flocculants in different model waters. With the PIV and
8 image analysis, the change in particle size distribution could be well recorded. Using the
9 sequence of flocculation, shear breakage and re-flocculation on a jar-test device together with
10 the PIV system, the rate of floc formation, the strength of the flocs, the recovery of broken
11 flocs, and the morphological and structural features of the flocs were characterized. The
12 results indicated that the adsorption of HA on the particle will stabilize the particles, hence
13 hindered the flocculation process. Sweep flocculation using a higher chemical coagulant
14 dosage was an effective means of process enhancement for the removal of particulates and
15 associated organic matter. The dynamics of A-B-R process was characterized by particle size
16 distribution (PSD) measurement with PIV setup. The particle strength and reversibility
17 capability were examined. Strength index showed the HA flocs have comparable strength,
18 while recovery index indicated a less recovery capability with the increasing of HA
19 concentration after exposure to a higher shear, especially for ferric HA flocs. It appears that
20 the bonds holding HA flocs together are not purely physical bonds given the limited regrowth
21 seen. Finally, evolution of floc structure during A-B-R process was analyzed by investigating
22 the fractal dimension D_b . The results were generally consistent with previous PSD
23 measurements. It suggested that the structure of flocs in breakage became more compact with
24 little permeability. An increase in floc compaction provides a further explanation for the
25 limited regrowth for most of flocs. According to the performances of alum and ferric, it can

26 be noticed that HA flocs have different properties dependent on which chemical coagulant is
27 used. Alum produced larger HA flocs which endured a higher recovery capability after
28 exploring higher shear, hence, compared to ferric, it could be preferred to using in the
29 practical enhanced coagulation unit.

30

31 *Keywords:* Aggregation-breakage-re-flocculation; humic acid; hydrodynamics; particle image
32 velocimetry (PIV).

33

34 **1. Introduction**

35 Humic acids (HAs) are one of the main constitute of natural organic matter (NOM) in
36 most water sources, resulting from the weathering and biodegradation of dead plants and
37 animals [1, 2]. The presence of NOM in the water is a major concern not only to form the
38 disinfection by-products, such as trihalomethanes (THMs), but to reduce the effectiveness of
39 filtration processes, such as membrane fouling [3, 4]. Since United States Environmental
40 Protection Agency has proposed that enhanced coagulation is a best available technology for
41 NOM removal [5, 6], extensive studies are addressed on the performance of humic acid
42 coagulation [7-9], however little thought is given to the physiochemical characteristics of HA
43 flocs. It includes floc size, compaction, strength and the potential to regrow after being
44 broken.

45 Enhanced coagulation is still a shear-induced flocculation, which means the shear
46 flow is a main reason to result in collisions which cause the flocs to grow. However, they can
47 still be subjected to higher shear rates where the flocs have to resist the corresponding
48 stresses [10]. By use of model spherical particles, such as latex beads, it has been previously
49 shown that particle suspensions destabilized with an ionic salt (i.e. NaCl) will reform to their
50 initial size if the original velocity gradient is subsequently reapplied [11]. This behaviour is
51 known as reversible breakage. However, in most instances where conventional metal
52 coagulants and polymers are used for the aggregation of small particle suspensions (such as
53 precipitated solids), irreversible breakage is usually seen, such that the initial floc size is
54 never subsequently achieved after breakage.

55 The irreversibility of aggregates during cycled shear is most likely the result of
56 particle-flocculant bond breakage during fragmentation, hence the reorganisation and
57 restructuring can both occur. Experimental shear-induced coagulation-fragmentation
58 processes [11-14] found that the fractal dimension shifts to a larger value compared to the

59 initial one, which showed a more compact structure formed during the cycled shear.
60 However, there has been no previous work showing the regrowth potential of HA flocs with
61 different coagulants and the structure restructuring and reorganisation during a cycled shear
62 schedule. Thus, an understanding of the reversible potential for natural organic flocs may
63 provide an important addition to the well-studied field of HA enhanced flocculation.

64 The structure of the HA containing flocs is non-homogeneous and should be
65 described by the fractal scaling law [15-17]. This fractal structure would affect the properties
66 of the flocs during aggregation-breakage-regrowth process. Although, more research effort
67 has been made to specify the properties of HA containing flocs, there is no study to report the
68 exact influence of the fractal structure on the hydrodynamic behaviours of flocs formed in
69 water treatment. Therefore, it merits more efforts to investigate the hydrodynamic properties
70 of the HA containing flocs and their relationship with the fractal structure of the flocs.

71 In present work, a series of the standard jar-tests were carried out to predict the
72 overall result of water flocculation and sedimentation. ζ -potential, turbidity removal and HA
73 reduction were determined for the particle suspension at various coagulant dosage. An
74 improved experimental facility for characterization of the PSD dynamics was developed by
75 making use of particle image velocimetry (PIV) coupled with image-analysis system. This
76 non-intrusive measurement of PSD was performed to characterize the HA flocs during
77 aggregation-breakage-regrowth (A-B-R) process. Morphology evolution indicated by surface
78 fractal dimension, breakage and regrowth potential of different HA flocs were evaluated with
79 different coagulants.

80

81 **2. Materials and Methods**

82

83 *2.1 Model waters and jar-test flocculation experiments*

84 Humic acid (HA) (Florida peat humic acid reference: 1R103H-2) was obtained from
85 International Humic Substances Society, and the stock HA solution was made by dissolving
86 the HA into de-ionized water. Kaolin (Aldrich, Milwaukee, WI) with a mean size of around
87 2.6 μm was used for making particle suspensions. Three types of the model waters that
88 contained 10 mg/L kaolin with an initial turbidity of 12 NTU and different HA contents were
89 prepared for the experimental study, including (1) HA0 – no HA addition, (2) HA3 - 3 mg/L
90 HA in water measured in terms of dissolved organic carbon (DOC), and (3) HA10 - 10 mg
91 DOC/L of HA. Two flocculants – alum ($\text{Al}_2(\text{SO}_4)_3 \cdot 14\text{H}_2\text{O}$) (BDH Chemicals, England) and
92 ferric chloride ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$) (UNI-Chem, Mumbai, India) - were tested for the flocculation
93 performance.

94 Standard jar-test flocculation and sedimentation experiments were conducted at room
95 temperature ($\sim 22^\circ\text{C}$) the model waters with a jar-test device (ZR4-6, Zhongrun, Shenzhen,
96 China). The jar-tester consisted of six 1-L rectangular beakers, each was filled with 500 mL
97 water, and the mixing was provided by with flat paddle mixers ($5.0 \times 4.0 \text{ cm}^2$). For each
98 model water, a flocculant, alum or ferric, was added at various dosages from 0 to 50 mg/L
99 into the six beakers. Throughout a flocculation experiment, the water pH was monitored by a
100 pH meter (420A, Orion, Boston, MA). 1 M NaHCO_3 was used to adjust the solution pH to a
101 proper range for the different flocculant dosages, i.e. pH \sim 7.0 for alum flocculation and
102 pH \sim 6.5 for ferric flocculation.

103 For a jar-test run, the water after the chemical addition was mixed rapidly at 100 rpm
104 for 60 sec. A sample of 10 mL was then withdrawn from each beaker for particle ζ -potential
105 measurement by a laser ζ -potential analyzer (Delsa 440SX, Coulter, Amherst, MA).
106 Following the rapid mixing, the water in the jar-test beakers was mixed for flocculation at a
107 slower rate of 30 rpm for 40 min followed by 30 min of sedimentation. The supernatant was
108 collected, for which the turbidity and HA residues were analyzed. The turbidity was

109 measured with a Turbidimeter (2100N, HACH, Loveland, CO), and the DOC in water was
110 measured by a TOC analyzer (5000A, Shimadzu, Kyoto, Japan). In addition, based on the ζ -
111 potential measurement and jar-test results, the optimal dose of a flocculant for a model water
112 could be determined. The optimal dose was then used for the flocculation-breakage-
113 reflocculation experiment on the model water characterized by the PIV technique.

114

115 *2.2 Particle image velocimetry (PIV) for particle size distribution measurement*

116 A PIV system was employed to track the change in particle size distribution (PSD)
117 during a flocculation experiment. PIV is an advanced and powerful flow visualization and
118 particle tracking technique [18]. As a non-intrusive optical setup, the PIV was able to capture
119 the image of particles in a jar-test beaker within a millisecond (Figure 1). The PIV system
120 consists of a laser illumination setup, a high-speed CCD video camera and a process control
121 and image processing software package. A pulsed laser beam generated from the source
122 (Coherent, Inc., Santa Clara, USA) was expanded to a thin laser light sheet by a combination
123 of a cylindrical and a spherical lenses. The laser sheet illuminated a planar region of the water
124 for visualization of the particles and flocs in the flocculation tank (beaker). The images of
125 laser-illuminated particles could be captured by a high speed CCD camera (PCO.imaging
126 1200 with a resolution of 1280×1024 pixels). The PIV system was controlled by a computer
127 with dedicated software (PCO.camware) for laser flushing, CCD recording, image acquisition
128 and storage. Images were processed with an image analysis system (Scion Image, Frederick,
129 MD) for PSD determination. For a floc of irregular shape, its size, d , was calculated in terms
130 of the equivalent diameter by $d = (4A/\pi)^{1/2}$, where A is the projected area of the floc. Based on
131 calibration, the PIV system had a resolution of around 9 μm for particle tracking and imaging
132 in the present flocculation study. More than 40 consecutive images within a minute were

133 analyzed to produce a size distribution of the particles, and the result was presented as either
134 a number-based discrete PSD or a volume-based discrete PSD.

135 In addition to size measurement, particle image analysis also can provide more
136 information about the morphological and structural feature of the aggregate flocs. For the 2-D
137 projected particle images, a boundary fractal dimension was used to characterize the fractal
138 property of the flocs. The boundary fractal dimension defines how the projected areas of the
139 particles scale up with the length of the perimeter [14, 16]. Accordingly, the boundary fractal
140 dimension, D_b , may be determined from the correlation [16, 19] as follows,

$$141 \quad A \propto P^{2/D_b} \quad (3)$$

142 where P is the perimeter of an aggregate. D_b ranges from 1 to 2, and there is no
143 straightforward relationship between D_b and the mass fractal dimension for a particle
144 population [15]. Nonetheless, a higher D_b value often suggests a more fractal structure of the
145 objects with a less spherical shape and irregular or rough surface [16].

146

147 *2.3 Flocculation-breakage and reflocculation test for floc strength and re-growth*

148 A single beaker jar-test device was used together with the PIV for characterization of
149 the flocculation dynamics (Figure 1). The jar-tester included a glass rectangular tank
150 ($L \times W \times H = 80 \times 80 \times 200$ mm) equipped with a flat paddle mixer that was driven by a DC
151 power supply. The flocculation procedure on a model water was the same as previous
152 described, i.e., after the chemical addition a pre-determined dose, the water was stirred
153 rapidly at 100 rpm for 60 s followed by a slow mixing at 30 rpm for 30 min. Upon the
154 completion of flocculation, the shear breakage and then re-flocculation experiment was
155 carried out. The breakage of flocs was conducted at 100 rpm for 15 min, which was followed
156 by re-flocculation with slow mixing at 30 rpm for 30 min. During the course of flocculation-

157 breakage-reflocculation, the images of particles and flocs in water was recorded and analyzed
158 by the PIV system. The PSDs at different phases of the process were therefore obtained.

159 The inter-particle bonds that hold aggregate flocs together are considered as the
160 cohesive strength of the flocs. A size ratio method [20] is used here with an index (σ) to
161 express the strength of particle flocs, i.e.,

$$162 \quad \sigma = d_2 / d_1 \quad (1)$$

163 where d_1 and d_2 are the mean sizes of the flocs before and after the shear breakage,
164 respectively. A higher value of the σ index indicates a higher strength of the flocs to resist
165 breakage when exposed to an elevated fluid shear.

166 When the shear intensity was reduced after the breakage phase, re-flocculation of the
167 particles could take place. A reversibility factor is used here to measure the re-flocculation
168 potential of the particles when the shear is reinstated to its original level. A modified size
169 ratio approach may be applied to calculate the reversibility (γ) by

$$170 \quad \gamma = \frac{d_3 - d_2}{d_1 - d_2} \quad (2)$$

171 where d_3 is the mean size of the particle flocs after re-flocculation at the original shear rate. A
172 higher reversibility index suggests a greater flocculation and re-growth capability of the flocs
173 after the shear breakage.

174

175 **3. Results and Discussion**

176

177 *3.1 Coagulation performance and water treatment results*

178 *3.1.1 ξ -potentials of the kaolin particles at different flocculant doses*

179 The clean kaolin was negatively charged with an average ζ -potential of around -30
180 mV (Figure 2). The presence of HA apparently increased the intensity of negative charges on

181 the particles, and the ζ -potential became more negative to -46 mV or below. Hence, HA
182 would cause further stabilization of particles in water. This is likely due to the steric or elastic
183 repulsion between particles brought about by the humic substances [21-23]. As anticipated,
184 addition of the flocculants, alum or ferric, could effectively reduce the surface charge of the
185 particles in all types of the model waters, resulting in particle destabilization. As the
186 flocculant dose increased, the particle ζ -potentials in the model waters approached zero.
187 Further increase in flocculant dose caused a certain extent of charge reversal of the particles
188 (Figure 2).

189 For pure kaolin with no HA in water, a small amount of the flocculants (5 mg/L or
190 lower) would eliminate the ζ -potentials and destabilize the particles completely. As the HA
191 content increased, the amount of alum or ferric required to achieve the same level of ζ -
192 potential reduction increased considerably. In comparison, alum appeared to be more
193 effective than ferric for reducing the ζ -potential of kaolin particles (Figure 2). For example,
194 with alum flocculation, the dose to neutralize the surface charge of kaolin was about 20 mg/L
195 for the HA3 water and 30 mg/L for the HA10 water. In ferric chloride flocculation, the
196 corresponding dose for charge neutralization was around 25 mg/L for HA3 and 30 mg/L for
197 HA10.

198

199 *3.1.2 Jar-test results of the flocculation and turbidity removal*

200 The jar-test flocculation and sedimentation results for turbidity and HA removals
201 from the model waters were in general agreement with would be expected from the ζ -
202 potential analysis. For clean kaolin without HA, a low flocculant dose of 5 mg/L was
203 sufficient to bring about flocculation for turbidity removal (Figure 3). With the presence of
204 HA in water, the flocculant demand for particle flocculation and turbidity removal increased
205 significantly. Alum appeared to be slightly more effective than ferric chloride for particle

206 flocculation. Nonetheless, a low alum dose below 20 mg/L still left a high level of HA
207 residue in water after flocculation and sedimentation. For the HA10 water with a high HA
208 content, at least 20 mg/L of alum or 25 mg/L of ferric chloride was needed to have
209 satisfactory flocculation and turbidity removal.

210 Enhanced flocculation with a high flocculant dose was effective to remove humic
211 substances from water (Figure 3). However, further increase of alum beyond 40 mg/L did not
212 bring about a notable improvement in turbidity and HA removals. For ferric chloride
213 flocculation, a dose of more than 40 mg/L actually worsened kaolin flocculation, particularly
214 for the HA0 and HA3 waters, resulting in poor turbidity removal. Judging from the ζ -
215 potential changes and jar-test results, the optimal alum doses chosen for the PIV-flocculation
216 experiments on the HA0, HA3 and HA10 waters were 10, 20 and 30 mg/L at pH~7.0,
217 respectively, and the optimal ferric chloride doses were 10, 25, and 30 mg/L at pH~6.5,
218 respectively.

219

220 *3.2 PSD dynamics, floc strength and re-flocculation capability*

221 *3.2.1 PIV characterisation of the flocculation dynamics*

222 The PIV technique is shown to be a powerful tool for obtaining the particle size
223 distributions in a dynamic fluid system. The PIV is a true non-intrusive particle tracking
224 system that is able to perform real-time *in-situ* particle imaging acquisition for determination
225 of the PSD dynamics during shear flocculation. The PSD of the flocs was expressed as the
226 volume-based discrete PSD, i.e. the percentages of the total particle volume observed against
227 a series of size sections (Figure 4).

228 The PIV results showed the continuous floc formation and growth in the model waters
229 during the chemical flocculation process. The PSD maintained a constant unimodal shape
230 with an apparent peak. Accordingly, the peak size, the particle size section corresponding to

231 the peak of the PSD curve, was used here as the mean size of the particle population observed
232 by the PIV. The change in the peak size of PSD with time illustrated well the flocculation-
233 breakage dynamics for a particle system. For both alum and ferric flocculation at the
234 respective optimal doses, flocs were well formed with a peak size of 1000 μm or larger. The
235 PSDs became rather stable in shape and position by the end of 30 min slow flocculation at 30
236 rpm. Shear breakage at a high stirring rate (100 rpm) caused a rapid and remarkable shift of
237 the PSDs to smaller sizes, and re-flocculation took place when the fluid shear was reduced
238 (Figure 4).

239 The PSD evolution showed effective alum and ferric flocculation on the jar-test
240 device (Figure 4). For either one of the flocculants, the HA0 water with pure kaolin and no
241 HA had the largest flocs formed, followed by the HA3 water and then the HA10 water. The
242 humic substances in water reduced the effectiveness of the flocculants in forming larger flocs.
243 However, with the higher doses used for HA3 and HA10 than that for HA0, floc formation in
244 the HA3 and HA10 waters occurred at a faster rate in the early phase of flocculation than that
245 in the HA0 water. In comparison, flocculation by alum produced larger flocs than ferric
246 chloride for the same water samples (Table 1). Nonetheless, compared to alum, ferric
247 chloride flocculation took place more rapidly after the chemical addition. For both
248 flocculants, the growth of particle flocs became much slower approaching the end of 30 min
249 slow flocculation.

250

251 *3.2.2 Breakage of the flocs and their re-flocculation*

252 A sudden increase in shear rate in the jar-test led to dramatic breakage of the
253 aggregate flocs in all water samples. Within 1 min or so, the peak sizes of the PSDs were
254 more than halved, according to the PIV observations (Figure 5). In agreement with previous
255 findings [19, 24], the large alum or ferric flocs formed by slow flocculation were rather

256 fragile and vulnerable to shear breakage. After the initial break-up, the breakage of the flocs
257 became much slower in the later phase of the breakage step. Thus, as suggested by others
258 [25-27], a shear breakage process may be classified into two phases. In the first phase, rapid
259 fragmentation of the flocs was the dominant phenomenon. In the following phase, particle
260 erosion could be the main cause for the slower decrease of floc sizes. Between the two
261 flocculants, ferric flocs appeared to be broken more easily to smaller flocs than alum flocs
262 (Figure 5).

263 As the shear was reduced to the original level, re-flocculation of the particles took
264 place, but to different extents in different model waters. Alum flocs were seen to have
265 considerable re-growth in size by flocculation (Figure 5). In comparison, ferric flocs were
266 more difficult to be recovered after breakage. The HA0 water showed the highest level of re-
267 flocculation for both flocculants, followed by HA3 and HA10. The humic matter in flocs
268 apparently decreased their potential of aggregation. In general, re-flocculation of the broken
269 particle flocs was a much slower process than the original flocculation after the flocculant
270 addition. Meanwhile, except for the alum flocs in HA0, the flocs recovered by re-flocculation
271 were much smaller in size than the flocs before breakage.

272

273 *3.2.3 Strength and recoverability of the alum and ferric flocs*

274 Based on the change in peak size of the PSD, the strength and reversibility of the
275 particle flocs formed in different model waters were determined (Table 1). After 15 min of
276 shear breakage, the alum flocs in HA0 showed the highest strength index at 32%. Other types
277 of flocs, including the alum flocs in HA3 and HA10 and all of ferric flocs had the strength
278 indexes that were rather similar to each other. In re-flocculation at a slower shear rate, the
279 alum flocs after breakage generally had a higher potential of recovery than the ferric flocs.
280 Pure kaolin flocs formed by alum flocculation had a of recovery index of 46%. As the HA

281 content in water increased, the recovery indexes of the broken flocs decreased significantly.
282 The alum flocs in HA10 had a low reversibility of only 10%. In agreement with the PIV
283 observations, the ferric flocs in HA10 after breakage almost could not be re-flocculated with
284 a recovery index as low as 4%.

285

286 *3.3 Morphology and Fractal Dimension of the Flocs*

287 *3.3.1 Morphology and boundary fractal dimension of the flocs*

288 The high-quality PIV images (Figure 6) also allow detailed analysis of the
289 morphology and structural features of the flocs suspended in water. According to equation 3,
290 the boundary fractal dimension of the particle flocs can be approximated from the slope of
291 the log-log regression of a series of projected areas versus perimeters of the particles. The D_b
292 of the flocs ranged from 1.11 to 1.22 after 30 min of slow flocculation (Figure 7). This is
293 somewhat lower than the value of from 1.1 to 1.4 reported for the aggregates of polystyrene
294 spheres [19]. For alum flocculation, flocs in the HA3 and HA10 waters had a slightly higher
295 D_b than that in HA0. For ferric chloride flocculation, flocs in HA10 had the highest D_b ,
296 followed by HA3 and then HA0.

297 A higher D_b normally indicates a more irregular and/or elongated shape and a rougher
298 surface for the particles, whilst a lower D_b suggests a more spherical shape and a smoother
299 surface of the particles. Based on the D_b values, the flocs formed in the HA0 water at a low
300 alum or ferric chloride dose were less fractal with a more regular shape and smooth surface.
301 In comparison, the alum and ferric flocs formed by enhanced flocculation at higher flocculant
302 doses were more fractal with an elongated shape and a rougher surface.

303

304 *3.3.2 Change of the floc morphology during breakage and re-flocculation*

305 After the shear breakage, flocs in all of the model waters were smaller and became
306 less fractal in shape with reduced D_b values (Figure 7). When exposed to a higher shear, it is
307 expected that the flocs would break at their weak points and rearrange into more stable
308 structures [11, 28]. Fragmentation of the elongated flocs would break up the flocs into
309 smaller pieces that were more close to spherical objects than the original flocs. As described
310 previously, re-flocculation at a reduced shear rate resulted in regrowth of the floc sizes.
311 Meanwhile, the fractal structure of the particle flocs was recovered partially as indicated by
312 an increase in fractal dimension. Nonetheless, similar to the PSDs, the D_b of the flocs could
313 not be fully recovered to their original levels (Figure 7). After re-flocculation, the D_b values
314 of the alum flocs generally were somewhat higher than those of the ferric flocs, which was in
315 agreement with the indication of the recovery index of the broken flocs (Table 1).

316

317 *3.3.3 Effect of the humic content on particle flocculation and floc strength.*

318 The addition of chemical coagulants would impose mainly two aspects of impact on
319 particle flocculation in water. One effect is to destabilize particles in a suspension, which
320 enhances particle flocculation. The other effect is to form hydrolyzing metal salts and their
321 precipitates that adsorb particle colloids. [23, 29] pointed out that flocs formed following
322 charge neutralization should have a high recoverability after breakage. In comparison, the
323 precipitates of hydrolyzed flocculants would have a much lower recoverability after breakage
324 [24, 30]. In the HA0 water with pure kaolin and no HA, the low dose of the coagulants could
325 destabilize kaolin in the suspension by charge neutralization prior to particle flocculation.
326 Nonetheless, the partially reversible breakage of the HA0 flocs suggests that the formation of
327 the hydrolyzed flocculant precipitates and the adsorption of kaolin by the flocs of the
328 precipitates also played an important role in removing particulate turbidity from water. For
329 the HA3 and HA10 waters, more flocculants had to be used for kaolin adsorption and

330 removal. At a high flocculant dose, larger flocs of the precipitates would be formed more
331 easily. In agreement with previous findings [24, 30], these types of the flocs of hydrolyzed
332 precipitates in HA3 and HA10 waters had a lower recoverability after breakage compared to
333 the HA0 flocs (Table 1).

334 The content of organic matter appeared to be an important factor to determine the
335 surface properties of formed flocs, such as adhesion, inter-particle interaction and floc
336 stability [9, 31-32]. The HA content also affects the surface fractal dimension of the particle
337 flocs [33]. In this study, the HA presence apparently facilitated the formation of a more
338 porous and more fractal structure corresponding with a higher D_b (Figure 7). These results are
339 well consistent with the findings of [33]. They found that untreated kaolin formed flocs with
340 a less fractal and more regular structure and the flos of kaolin with HA attained a more
341 irregular and more fractal structure.

342

343 **4. Conclusion**

344 A series of standard jar-test flocculation experiments were performed on the model
345 waters with kaolin and various amounts of humic acids, 0 (HA0), 3 mg/L (HA3) and 10 mg/L
346 (HA10). Judging from the ζ -potential changes and the jar-test results, the optimal alum doses
347 for the HA0, HA3 and HA10 waters were 10, 20 and 30 mg/L, respectively, and the optimal
348 ferric chloride doses were 10, 25, and 30 mg/L, respectively.

349 The PIV technique was employed successfully to record and characterize the PSD
350 dynamics during the flocculation process in water treatment. The PIV system together with
351 the image analysis technique is capable to track the change in PSD on a jar-test during
352 particle flocculation and floc breakage and re-flocculation. Based on the change in the peak
353 size of the PSD, the strength and reversibility of the particle flocs formed in different model
354 waters were determined. The results showed that the alum flocs were somewhat stronger than

355 the ferric flocs. As the HA content in water increased, the recovery index of the flocs after
356 breakage decreased significantly. The alum flocs in HA10 had a reversibility of only 10%,
357 while the ferric flocs in HA10 after breakage could hardly be re-flocculated with a recovery
358 index as low as 4%.

359 According to the PIV images, the flocs formed initially after the flocculant addition
360 were larger and more fractal with a higher value of boundary fractal dimension D_b . After
361 shear breakage, the flocs became smaller and less fractal with a lower D_b . With the re-
362 flocculation, the fractal structure of the flocs could be only partially recovered. The results
363 suggested that initially aggregates have a ramified, open structure that became more compact
364 as exposure to a higher shear. An increase in floc compaction would lead to a reduction in
365 floc size, which provides a further explanation for the limited regrowth of most flocs. The
366 broken flocs seemed to become somehow difficult to form previous porous and fractal
367 clusters. It indicated the chemical bonds or/and chemical adsorption formed in first
368 aggregation phase were broken and flocs become more stable and rearrange into more
369 compact structure during the recovery stage.

370

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376

377 **References**

378 [1] S. Chang, R.A. Berner, Humic substance formation via the oxidative weathering of coal.
379 Environ. Sci. Technol. 32 (1998) 2883-2886.

- 380 [2] W.P. Cheng, F.H. Chi, A study of coagulation mechanisms of polyferric sulfate reacting
381 with humic acid using a fluorescence-quenching method. *Water Res.* 36 (2002) 4583-
382 4591.
- 383 [3] A. Thekkedath, W.M. Naceur, K. Kecili, M. Sbai, A. Elana, L. Auret, H. Suty, C.
384 Machinal, M. Pontie, Macroscopic and microscopic characterizations of a cellulosic
385 ultrafiltration (UF) membrane fouled by a humic acid cake deposit: First step for
386 intensification of reverse osmosis (RO) pre-treatments. *C. R. Chimie* 10 (2007) 803-812.
- 387 [4] A. Rubia, M. Rodriguez, V.M. Leon, D. Prats, Removal of natural organic matter and
388 THM formation potential by ultra- and nanofiltration of surface water. *Water Res.* 42
389 (2008) 714-722.
- 390 [5] EPA, Enhanced coagulation and enhanced precipitative softening guidance manual. EPA
391 815-R-99-012, 1999.
- 392 [6] J.K. Edzwald, Enhanced coagulation: US requirements and a broader view. *Water Sci.*
393 *Technol.* 40 (1999) 63-70.
- 394 [7] D. Schmitt, H.E. Taylor, G.R. Aiken, D.A. Roth, F.H. Frimmel, Influence of natural
395 organic matter on the adsorption of metal ions onto clay minerals. *Environ. Sci. Technol.*
396 36 (2002) 2932-2938.
- 397 [8] Y. Wang, B. Du, J. Liu, B. Shi, H. Tang, Surface analysis of cryofixation-vacuum-
398 freeze-dried polyaluminum chloride–humic acid (PACl–HA) flocs. *J. Colloid Interface*
399 *Sci.* 316 (2007) 457-466.
- 400 [9] X. Y. Li, U. Passow, B. E. Logan, Fractal dimension of small (15-200 μm) particles in
401 Eastern Pacific coastal waters. *Deep-Sea Res.* 45 (1998) 115-118.
- 402 [10] N.D. Vassileva, van den D. Ende, F. Mugele, J. Mellema, Fragmentation and erosion of
403 two-dimensional aggregates in shear flow. *Langmuir* 23 (2007) 2352-2361.

- 404 [11]P.T. Spicer, S.E. Pratsinis, J. Raper, R. Amal, G. Bushell, G. Messters, The reversibility
405 of floc breakage. *Powder Technol.* 97 (1998) 26-34.
- 406 [12]V. Oles, Shear-induced aggregation and breakage of polystyrene latex particles. *J.*
407 *Colloid Interf. Sci.* 154 (1992) 351-358.
- 408 [13]K. A. Kusters, S. E. Pratsinis, S. G. Thoma, D. M. Smith, Ultrasonic fragmentation of
409 agglomerate powders. *Chem. Eng. Sci.* 48 (1993) 4119-4127.
- 410 [14]T. Serra, X. Casamitjana, Structure of the aggregates during the process of aggregation
411 and breakup under a sheer flow. *J. Colloid Interface Sci.* 206 (1998) 505-511.
- 412 [15]P. Meakin, Fractal aggregates. *Adv Colloid Interface Sci.* 28 (1988) 249-331.
- 413 [16]Q. Jiang, B.E. Logan, Fractal dimensions of aggregates determined from steady-state size
414 distributions. *Environ. Sci. Technol.* 25 (1991) 2031-2038.
- 415 [17]S.R. Grey, C.B. Ritchie, T. Tran, B.A. Bolto, Effect of NOM characteristics and
416 membrane type on microfiltration performance. *Water Res.* 41 (2007) 3833-3841.
- 417 [18]M. Raffel, C.E. Willert, J. Kompenhans, Particle image velocimetry: a practical guide.
418 Springer, Berlin, 1998
- 419 [19]P.T. Spicer, S.E. Pratsinis, Coagulation and fragmentation: Universal steady-state
420 particle-size distribution. *AIChE J.* 42 (1996) 1612-1620.
- 421 [20]P. Jarvis, B. Jefferson, J. Gregory, S.A. Parsons, A review of floc strength and breakage.
422 *Water Res.* 39 (2005) 3121-3137.
- 423 [21]M. Elimelech, J. Gregory, X. Jia, R. A. Williams, Particle deposition and aggregation:
424 measurement, modelling and simulation, Butterworth-Heinemann, Woburn, MA, 1995.
425 p9-63, 261-289.
- 426 [22]M. Stumm, J.J. Morgan, Aquatic Chemistry: Chemical Equilibrium and Rates in Natural
427 Waters. Wiley, New York, 1995.

- 428 [23]A. Amirtharajah, C.R. O'Melia, Coagulation processes: Destabilisation, Mixing and
429 Flocculation. In AWWA Water Quality and Treatment: a Handbook of Community
430 Suppliers. McGraw-Hill, New York, 1999.
- 431 [24]R.J. Francois, Ageing of aluminum hydroxide flocs. *Water Res.* 21 (1987) 523-531
- 432 [25]D.S. Parker, W.J. Kaufman, D. Jenkins, Floc breakup in turbulent flocculation processes.
433 *Journal of the Sanitary Engineering Division-ASCE* 98 (NSA1) (1976) 79-99.
- 434 [26]P.A. Shamlou, S. Stavriniades, N. Titchener-hooker M. Hoare, Growth-independent
435 breakage frequency of protein precipitates in turbulently agitated bioreactors. *Chem.*
436 *Eng. Sci.* 49 (1994) 2647-2656.
- 437 [27]C.A. Biggs, P.A. Lant, Activated sludge flocculation: On-line determination of floc size
438 and the effect of shear. *Water Res.* 34 (2000) 2542-2550.
- 439 [28]C. Selomulya, R. Amal, G. Bushell, T.D. Waite, Evidence of shear rate dependence on
440 restructuring and break-up of latex aggregates. *J. Colloid Interface Sci.* 236 (2001) 67-
441 77.
- 442 [29]V. Chaignon, B.S. Lartiges, A.El. Samrani, C. Mustin, Evolution of size distribution and
443 transfer of mineral particles between flocs in activated sludges: an insight into floc
444 exchange dynamics. *Water Res.* 36 (2002) 676-684.
- 445 [30]K. McCurdy, K. Carlson, D. Gregory, Floc morphology and cyclic shearing recovery:
446 comparison of alum and polyaluminum chloride coagulants. *Water Res.* 38 (2004) 486-
447 494.
- 448 [31]M.R. Jekel, The stabilization of dispersed mineral particles by adsorption of humic
449 substances. *Water. Res.* 20 (1996) 1543 – 1554.
- 450 [32]B. Janczuk, T. Białopiotrowicz, A. Zdziennicka, M. Hajnos, G. Jozefaciuk, The influence
451 of soil clay constituents on surface free energy of clay fractions. *J. Soil Sci.* 43 (1992)
452 27–35.

453 [33]Z. Sokolowska, S. Sokolowski, Influence of humic acid on surface fractal dimension of
454 kaolin: analysis of mercury porosimetry and water vapour adsorption data. Geodema 88
455 (1998) 233-249.

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457 **Figure captions**

458 Figure 1. Schematic diagram of the PIV coupling with an image analysis system.

459 Figure 2. ζ -potential of the three particle systems as a function of the coagulant dose.

460 Figure 3. Turbidity and TOC after the jar-test flocculation and sedimentation experiments as
461 a function of the flocculant dose for the three particle systems.

462 Figure 4. PSD profiles of the particles flocs for different HA contents in water during the A-
463 B-R process.

464 Figure 5. Change of the peak size of flocs for different HA contents in water during the A-B-
465 R tests.

466 Figure 6. Examples of the PIV images of particle flocs during a typical A-B-R process.

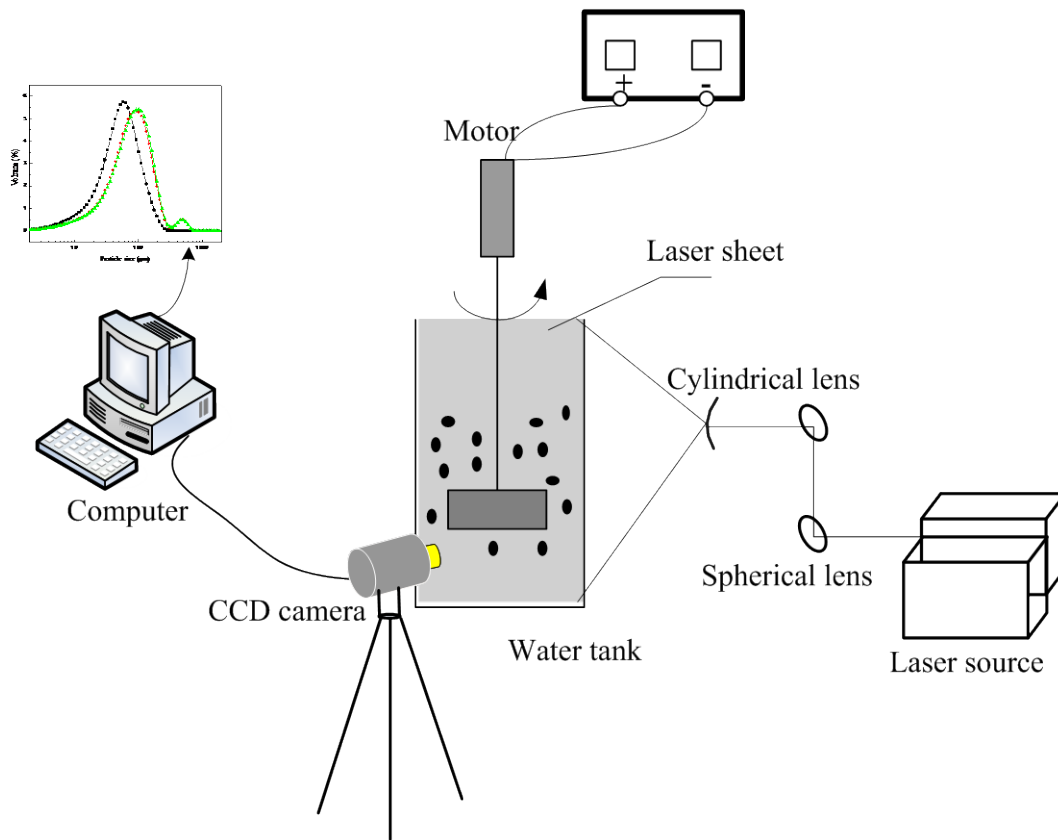
467 Figure 7. Change of the fractal dimension of the particle flocs with different HA contents in
468 water during the A-B-R tests.

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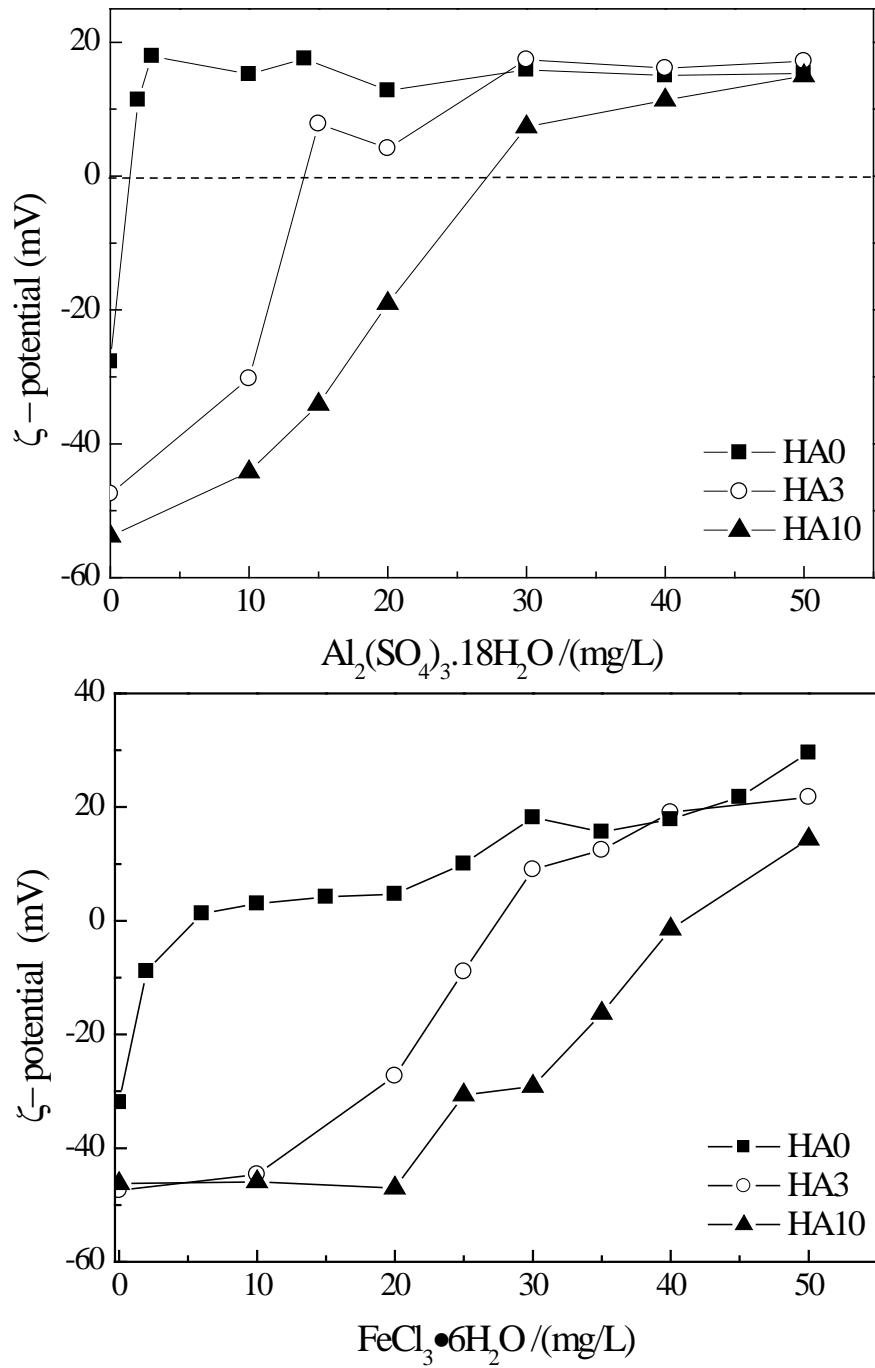


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475 **Figure 1.** Schematic diagram of the PIV coupling with an image analysis system.

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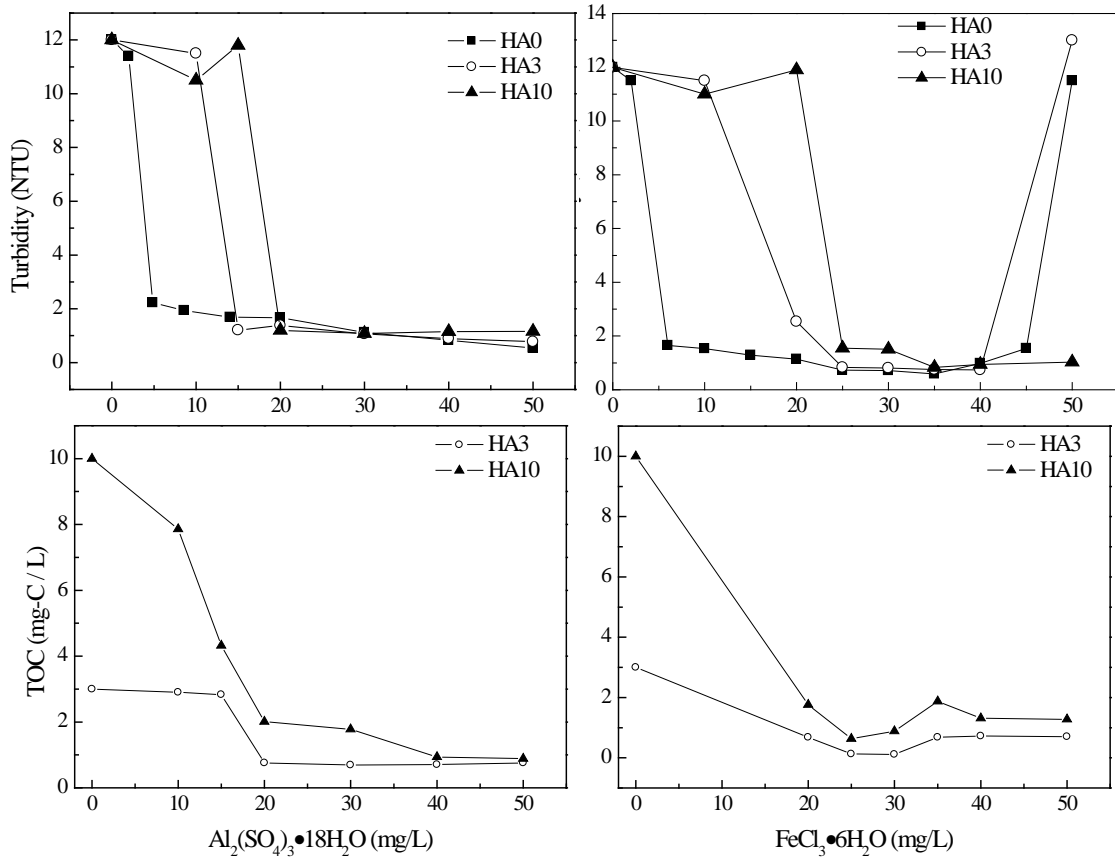
478 **Figure 2.** ζ -potential of the three particle systems as a function of the coagulant dose.

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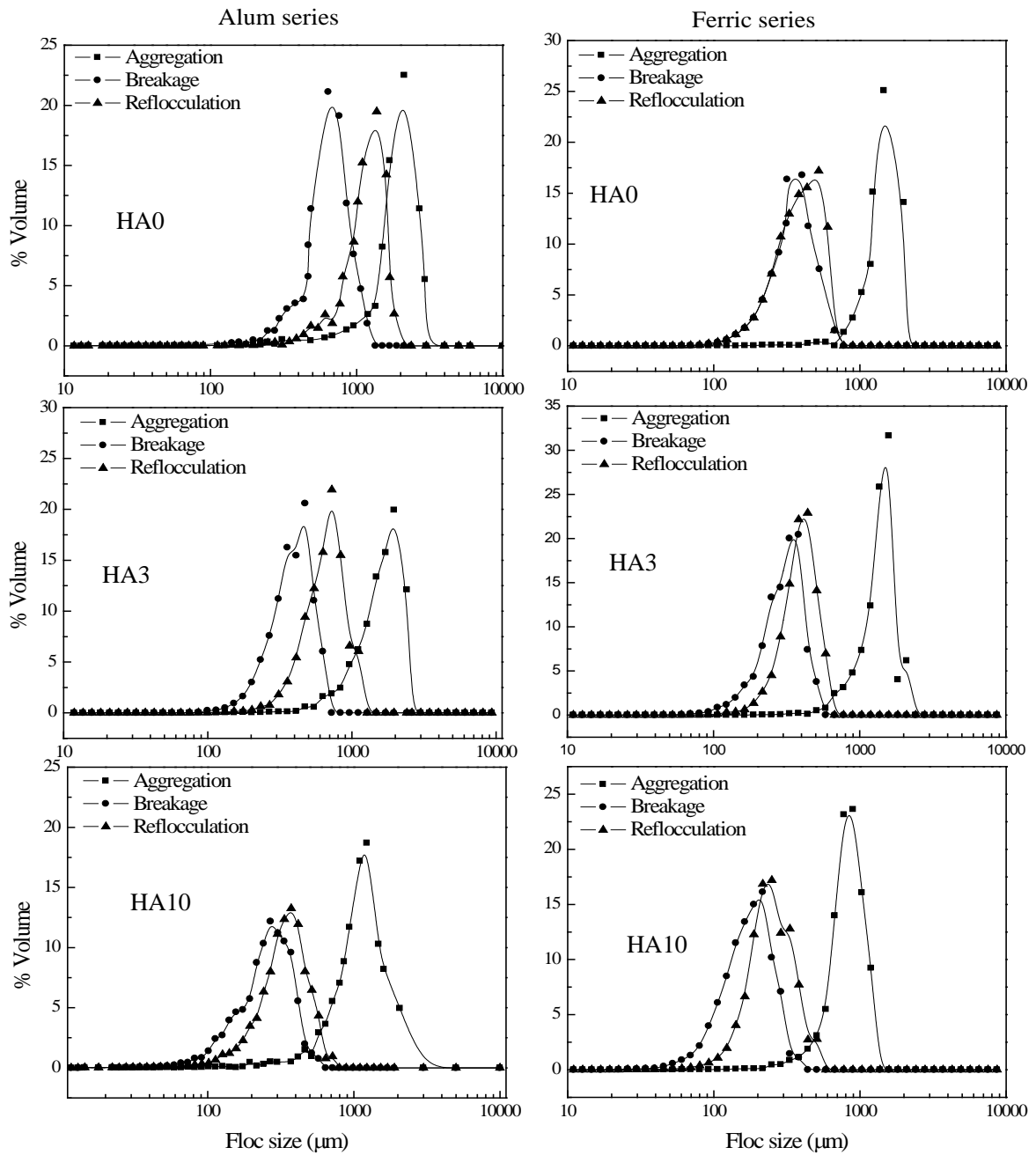
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485 a function of the flocculant dose for the three particle systems.

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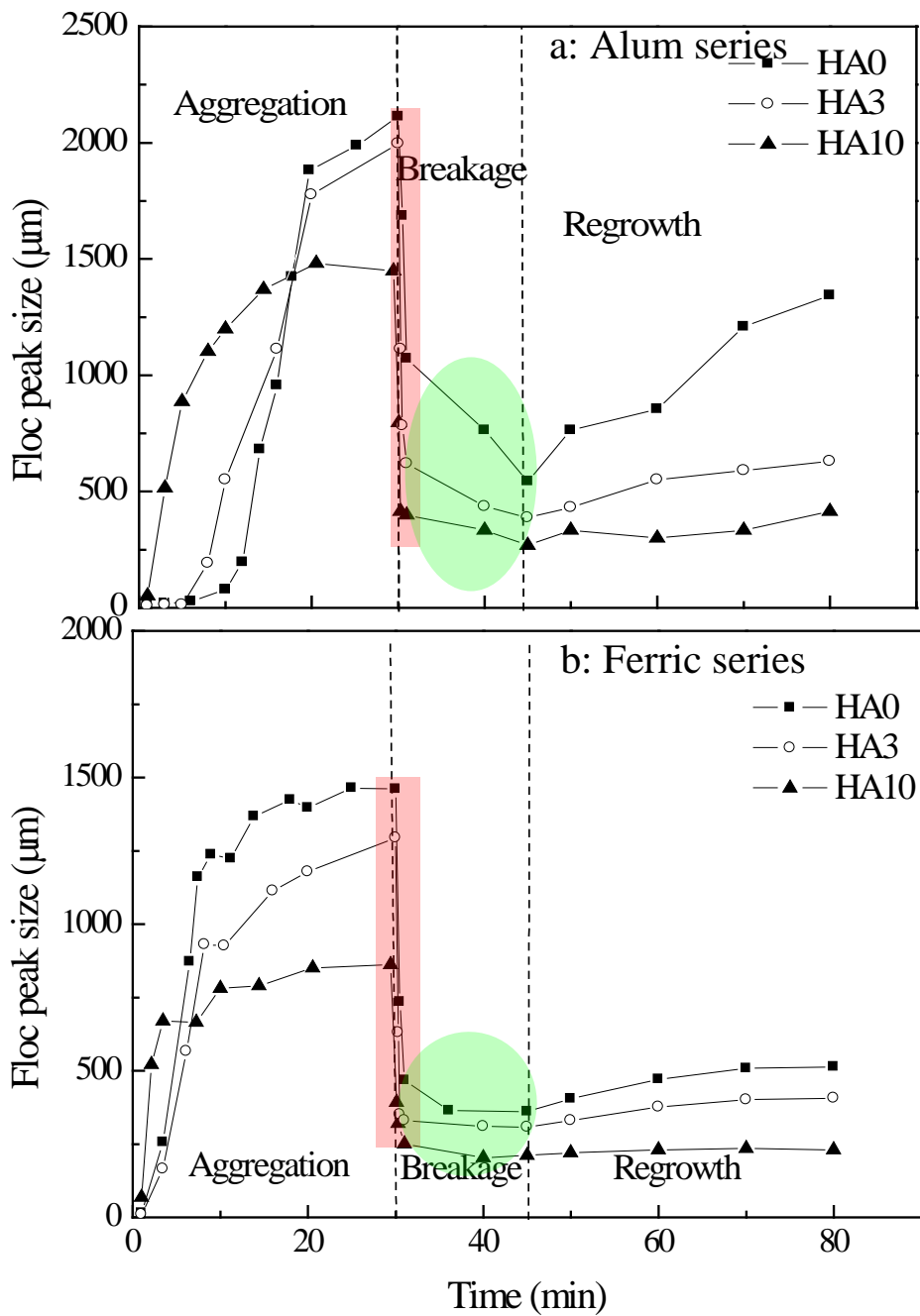
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491 **Figure 4.** PSD profiles of the particles flocs for different HA contents in water during the A-

492 B-R process.

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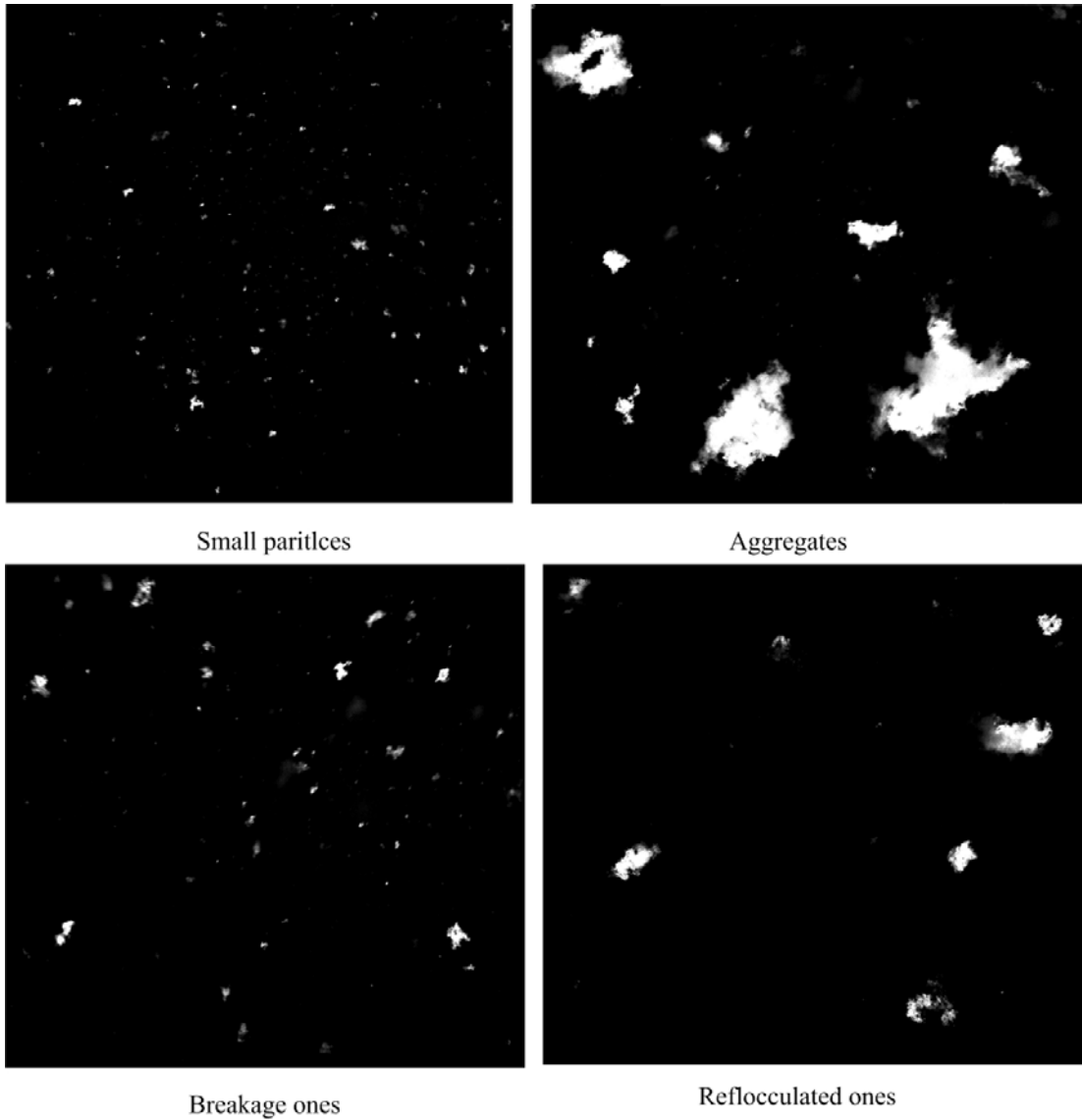
497 **Figure 5.** Change of the peak size of flocs for different HA contents in water during the A-B-
 498 R tests.

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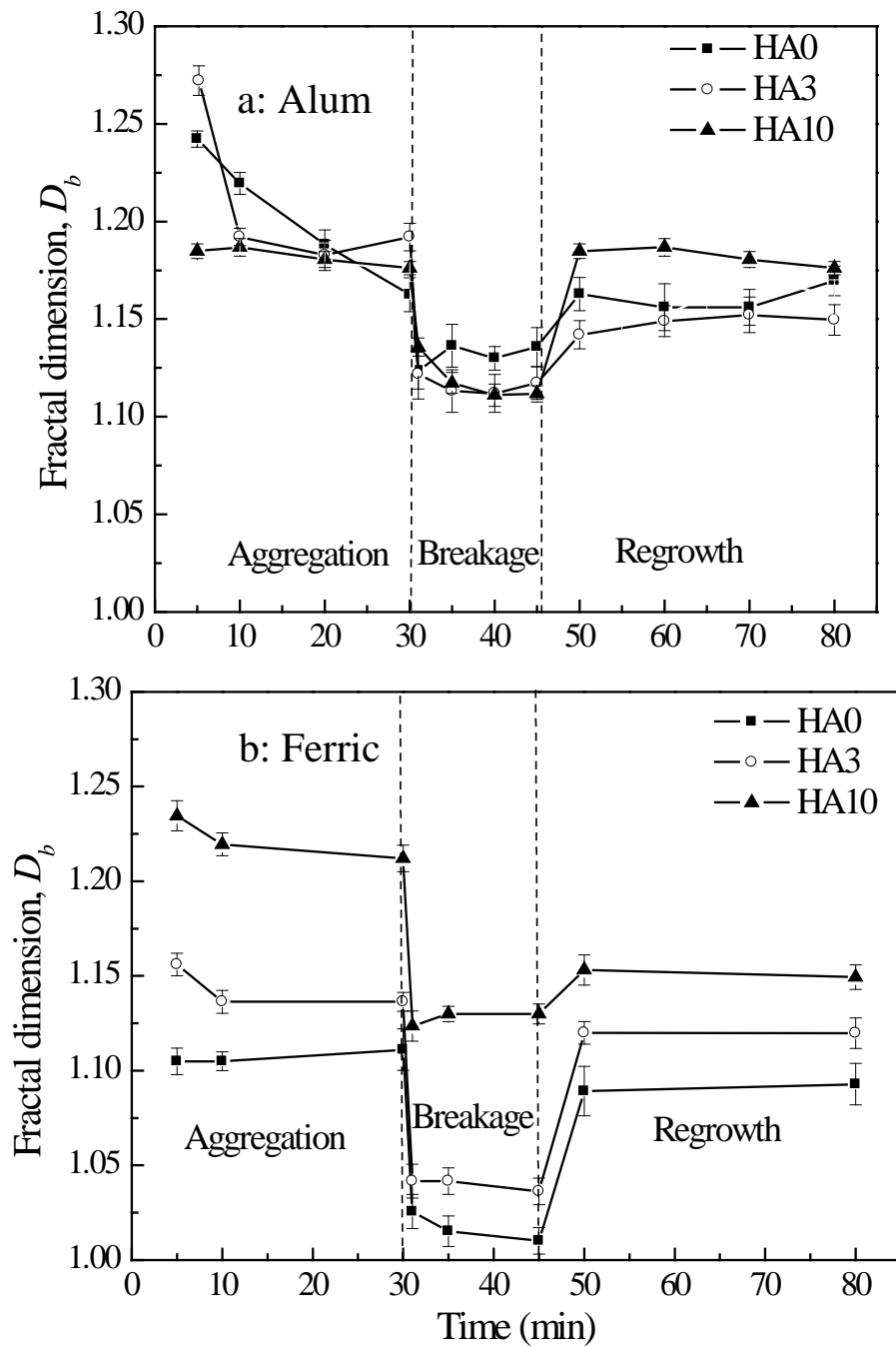
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Figure 6. Examples of the PIV images of particle flocs during a typical A-B-R process.



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