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<td><strong>Author(s)</strong></td>
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Hedgehog/Notch-induced premature gliogenesis represents a new disease mechanism for Hirschsprung disease in mice and humans

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Hirschsprung (HSCR) disease is a complex genetic disorder attributed to a failure of the enteric neural crest cells (ENCCs) to form ganglia in the hindgut. Hedgehog and Notch are implicated in mediating proliferation and differentiation of ENCCs. Nevertheless, how these signaling molecules may interact to mediate gut colonization by ENCCs and contribute to a primary etiology for HSCR are not known. Here, we report our pathway-based epistasis analysis of data generated by a genome-wide association study on HSCR disease, which indicates that specific genotype constellations of Patched (PTCH1) (which encodes a receptor for Hedgehog) and delta-like 3 (DLL3) (which encodes a receptor for Notch) SNPs confer higher risk to HSCR. Importantly, deletion of Ptc1 in mouse ENCCs induced robust Dll1 expression and activation of the Notch pathway, leading to premature gliogenesis and reduction of ENCC progenitors in mutant bowels. Dll1 integrated Hedgehog and Notch pathways to coordinate neuronal and glial cell differentiation during enteric nervous system development. In addition, Hedgehog-mediated gliogenesis was found to be highly conserved, such that Hedgehog was consistently able to promote gliogenesis of human neural crest–related precursors. Collectively, we defined PTCH1 and DLL3 as HSCR susceptibility genes and suggest that Hedgehog/Notch-induced premature gliogenesis may represent a new disease mechanism for HSCR.

Introduction
Hirschsprung (HSCR) disease is the most common neurocris-topathy in humans, affecting 1:5,000 newborns. It is a complex genetic disorder attributed to a failure of the enteric neural crest cells (ENCCs) to form ganglia in the hindgut. Colonization of the gut by ENCCs depends on synchronization and balance of the signaling networks involved in the proliferation, migration, and differentiation of ENCCs (1). Hedgehog (Hh) and Notch are implicated in mediating proliferation and differentiation of ENCCs (1). Hedgehog (Hh) and Notch are known signaling pathways implicated in these processes.

Hh signal transduction is mediated by 2 transmembrane proteins: Smoothened (Smo) is the obligatory signal transducer, and the Hh-binding receptor Patched (Ptc1) constitutively blocks Smo activity in the absence of Hh. Hh binding to Ptc1 unleashes Smo activity and activates the downstream cascade by blocking the formation of the repressor form of the GLI-Kruppel family member GLI1 (Gli1) family of zin-c-finger transcription factor, Gli1Δ, and promoting the formation of Gli1 activators, Gli1Δ (2, 3). It is known that Sonic Hh (Ssh) signaling is essential for diverse developmental processes and implicated in various diseases, particularly, those affecting the nervous system and cancer (4–8). Both graded activity and stepwise regulation of Hh signaling are important for its actions. Importantly, mice lacking either the Indian Hh– or Shh-secreted proteins show partial intestinal aganglionosis, accompanied by megacolon or ectopic ganglia formation, respectively (9), suggesting the biological relevance of these molecules in enteric nervous system (ENS) development.

Similarly, Notch signaling also plays a key role in various aspects of neurogenesis of ENCCs. Activation of Notch signaling is required not only for the maintenance of ENCCs (10) but also for the switching of these cells from a program of neurogenesis to gliogenesis in vitro (11) and in vivo (12). Notch receptors, Notch1 and Notch2, and their ligands, delta-like 1 and delta-like 3 (Dll1 and Dll3), are the main forms expressed in the ENCCs (ref. 10 and our unpublished observations). Notch ligands activate Notch signaling in neighboring cells by inducing a series of proteolytic cleavages of the receptor. The resulting Notch intracellular domain (NICD) translocates to the nucleus in which it associates with the DNA-binding protein CSL (RBP-Jκ) to initiate the transcription of target genes, such as hairy and enhancer of split-1 (Hes1) (13), which in turn, represses the expression of proneural proteins (e.g., Math1) to switch differentiation from neurogenesis to gliogenesis.

Genetic alterations in these signaling molecules may therefore interfere with the colonization process and potentially represent a primary etiology for HSCR. Theoretically, HSCR could be caused by deleterious mutations in a critical signaling component and/or cumulative effects of multiple less severe mutations in these pathways. Nevertheless, so far, none of the genes involved in Hh and
Notch pathways were found to be associated with HSCR, neither in a genome-wide association study (GWAS), nor in a mutation screening conducted on patients with HSCR (14). In this study, we further exploited the data initially generated by our GWAS by applying canonical correlation analysis (CCA) to assess whether interactions among gene members of the Hh and Notch signaling pathways could contribute to HSCR. In addition, the coordination mechanism between the Hh and Notch pathways in ENS development has also been examined using EIGENSTRAT, the P score of which was still highly significant (P = 0.008). Correlations among genes for each group are illustrated in Supplemental Table 2. Given the relevance of ret proto-oncogene (RET) SNPs in HSCR (14), we investigated whether the epistasis identified was dependent on RET using partial CCA. The P value was 0.014, indicating that the epistasis was RET independent.

To identify the DLL3 and PTC11 SNPs driving the correlation, we then used multifactor-dimensionality reduction (MDR). This revealed that 2 out of the 12 PTC11 SNPs (rs357552 [A/G] and rs10512248 [A/C]; throughout, underlining indicates a risk allele) and 1 out of the 4 SNPs in DLL3 (rs2354225 [A/G]) were the most relevant. These SNPs map to putative gene-regulatory regions. Specific genotype constellations of these 3 SNPs conferred higher risk to disease (odds ratio [OR] = 2.582 under an additive model and OR = 2.220 under a dominant model [P = 0.0003]) (Table 1). Indeed, our patient population was enriched with individuals with risk constellations (68.2%) when compared with controls (46.1%).

Notch-dependent Hh activity in ENCCs. In light of our genetic and expression data, we proceed to investigate the potential functional interaction of these 2 pathways in ENS development. We and others have previously demonstrated that ENCCs express Hh and Notch signaling components, including Ptc1, Dll1, and Dll3, and that Hh and Notch signaling are required for the maintenance of the ENCCs by promoting growth and attenuating premature neuronal differentiation (10, 15).

In this study, we further examined the potential functional cross-talk between Hh and Notch signaling in ENS development. Like in other systems, Shh or its agonist (purrmorphamine) activated the Ptc1-Smo-Gli1 pathway to promote the proliferation of ENCCs (Ret+/neuron-specific class III β-tubulin/glial fibrillary acidic protein: [Ret-/Tuj1-/GFAP −]) (Supplemental Figure 1). To assess whether induction of proliferation by Shh is caused by activation of Notch signaling, ENCCs isolated from wild-type mice were treated with Shh in presence or absence of an inhibitor of γ-secretase, DAPT, which suppresses proteolysis-dependent activation of endogenous Notch receptors (16). Consistent with the above finding, Shh could significantly promote the proliferation of ENCCs, while DAPT treatment counteracted the Shh-induced proliferation (Figure 1, A and B). Thus, Hh-induced proliferation is Notch dependent. In concordance with this data, RT-PCR analysis demonstrated that Shh promotes the expression of a Notch ligand, Dll1, and upregulates the expression of a principle effector of Notch pathway, Hes1 (Figure 1, C–E). Cotreatment of ENCCs with DAPT and Shh caused a similar induction of Dll1 expression as that seen with Shh alone; by contrast, Shh-mediated induction of Hes1 was suppressed (Figure 1C). This observation suggests that Shh directly upregulates Dll1, while induction of Hes1 expression is the concomitant of Notch activation. Consistently, as illustrated in the quantitative RT-PCR analyses, Shh-induced expression of Dll1 and Hes1 could be mimicked by the Shh agonist, purmorphamine (Figure 1, D and E). It is noteworthy that upregulation of Dll3 was not observed in mouse ENCCs (data not shown). Besides the chemical inhibition, we also suppressed the Dll1 expression in ENCCs by siRNA knockdown. Knockdown of Dll1, but not Hes1, with siRNA in ENCCs significantly abolished Hh-induced proliferation, further indicating that Hh activates the Notch pathway by upregulating Dll1 expression in ENCCs (Figure 1F and Supplemental Figure 2).

Deletion of Ptc1 activates Hh signaling and promotes proliferation of ENCCs. Concordant results were independently obtained from Ptc1 conditional mice. At E11.5, ENCCs were isolated from embryonic guts of floxed Ptc1 mice (Ptc1f/f mice), which have Cre recombinase

Table 1
Genetic interaction between PTC11 and DLL3 SNPs and summary of subject statistics

<table>
<thead>
<tr>
<th>PTC11 (Chr9q22.32)</th>
<th>DLL3 (Chr19q13.2)</th>
<th>Case/control frequency (%)</th>
<th>OR (95% CI)</th>
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<tr>
<td>rs357552 (A/G)</td>
<td>rs2354225 (A/G)</td>
<td></td>
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</tr>
<tr>
<td>AG or AA</td>
<td>AA</td>
<td>24.85/15.40</td>
<td>2.78 (1.717–4.514)</td>
</tr>
<tr>
<td>AG or AA</td>
<td>A/C + C</td>
<td>24.28/15.60</td>
<td>2.64 (1.627–4.281)</td>
</tr>
<tr>
<td>GG</td>
<td>AA</td>
<td>9.25/6.19</td>
<td>2.53 (1.277–5.021)</td>
</tr>
<tr>
<td>AG or AA</td>
<td>AA</td>
<td>9.82/9.17</td>
<td>1.82 (0.958–3.440)</td>
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MDR identified that 2 SNPs in PTC11, rs357552 and rs10512248, and 1 SNP in DLL3, rs2354225, were the most relevant on driving the correlation of Hh and Notch pathways. Shown are specific genotype constellations of PTC11 and DLL3 SNPs, with risk alleles (for rs357552, A; for rs10512248, C; for rs2354225, A) denoted underlined. Case/control frequencies and corresponding risks for HSCR are also shown; genotype constellations are listed in descending order according to risk. Other low-risk genotypes returned a case/control frequency of 31.80/53.90.
recognition (loxP) sites flanking exon 3 of the \textit{Ptch1} gene (ref. 17 and Figure 2A). \textit{Ptch1}\textsuperscript{f/f} ENCCs were then infected with Cre-encoding adenovirus (Ad-GFP-Cre) to delete exon 3 of the \textit{Ptch1} gene; the expression of the mutant \textit{Ptch1} and other Hh target genes was analyzed by RT-PCR. The majority of ENCCs were expressing GFP 72 hours after the infection with the control virus (Ad-GFP) and Ad-GFP-Cre (Figure 2A). Consistently, RT-PCR also indicated that Cre adeno virus can completely delete exon 3 of the \textit{Ptch1} gene in ENCCs to generate the null mutant form of \textit{Ptch1} (Figure 2B). Given that \textit{Ptch1} is a negative regulator of Smo, inactivation of \textit{Ptch1} constitu-

Figure 1
Shh upregulates \textit{Dll1} expression to mediate proliferation of ENCCs. (A) BrdU proliferation assay showed that Shh (3 days) significantly (\(P < 0.001\)) increases the proliferation of ENCC progenitors (BrdU\(^+/\)Ret\(^+\)). This effect could be counteracted by treatments with DAPT (10 mM). Scale bar: 100 \(\mu\)m. Ctrl, control. (B) Proliferating Ret-expressing cells that incorporated with BrdU (for 16 hours) were counted. The values reported in bar charts represent the mean \(\pm\) SEM, and 3 independent experiments were performed. Data were analyzed by 1-way ANOVA, followed by the Tukey post-hoc test. \(P\) values of less than 0.05 are statistically different from those of the control. (C) Semiquantitative RT-PCR for the Notch pathway genes after treatment with Shh in presence or absence of the Notch inhibitor DAPT (10 mM) (3 days). (D and E) Quantitative RT-PCR (qRT-PCR) for the expression of \textit{Dll1} and \textit{Hes1} in ENCCs treated with Shh, Shh plus DAPT, and Hh agonist, purmorphamine (Pur, 1 mM) (3 days). (F) Knockdown of \textit{Dll1} significantly abrogated the Shh-mediated proliferation (3 days). Quantitative RT-PCR for the expression of \textit{Dll1} shows that siRNA can significantly downregulate \textit{Dll1} expression (3 days). Nonsilencing RNA (NS) was used as a negative control. Bars represent the mean \(\pm\) SEM, and 3 independent experiments were performed, each in triplicate. \(P\) values of less than 0.05 were considered to be statistically significant.
tively activates the Hh pathway in ENCCs similar to that observed in control ENCCs treated with Shh. Cre adenovirus-mediated deletion of Ptc1 (Ad-GFP-Cre) resulted in an elevated expression of Gli1, accompanied by an extensive proliferation of ENCCs. It was similar to the control virus–infected ENCCs with Shh treatment (Figure 2, B and C). Addition of Shh to Ptc1−/− ENCCs did not further increase the expression of Gli1 (Figure 2B) or their proliferation (Supplemental Figure 3), suggesting that Ptc1 deletion is sufficient to fully activate Hh signaling. A similar counteracting effect of DAPT was also observed in Ptc1−/− ENCCs, indicating that activation of Notch by Shh or by deletion of Ptc1 in ENCCs is required for the observed elevated proliferation (Figure 2C).
Deletion of Ptch1 in ENCCs inhibits neurogenesis and is accompanied by spontaneous gliogenesis. To examine whether deletion of Ptch1 affects the differentiation capacity of ENCCs, we induced the neuronal differentiation of ENCCs with the administration of glial cell–derived neurotrophic factor (GDNF). Neuronal and glial differentiation were monitored based on the expression of tyrosine hydroxylase (TH) or TuJ1 and S100β, respectively. The control virus–infected ENCCs gave rise to both neurons and glia in the presence of GDNF (Figure 2D and E). Conversely, deletion of Ptch1 in ENCCs resulted in a significant reduction of TH⁺ (Figure 2D) and TuJ1⁺ (Supplemental Figure 4) cells upon the GDNF treatment. In addition, spontaneous gliogenesis was observed in Ptch1⁻/⁻ ENCCs, and a
significantly greater percentage of cells were S100β- in the absence of GDNF (Figure 2E). GDNF treatment of these glia-committed ENCCs, on the other hand, slightly reduced the percentage of S100β- cells in the culture, suggesting that it may exhibit a negative regulatory effect on their spontaneous gliogenesis (Figure 2E). Nevertheless, GDNF treatment alone was not sufficient to fully retrieve the neuronal differentiation capacity of Ptch1+/− ENCCs (Figure 2E). Consistently, the expression of GFAP was significantly upregulated, specifically in the neural crest, including premigratory ENCCs (10).

To study the effect of aberrant Hh signaling in premigratory ENCCs, we generated neural crest–specific Ptch1 knockout mice by crossing Ptch1f/f mice with Wnt1-Cre transgenic mice, which express Cre recombinase specifically in the neural crest, including premigratory ENCCs (10). Neural crest–specific Ptch1 mutant mice died at E12 and exhibited a significant (P < 0.001) reduction of proliferative ENCCs (p75NTR+/Ki67−) in the bowel (Figure 4). In concordance with the in vitro data, significantly more Notch-activated cells (Hes1+) were found in the mutant bowels (Figure 5, A and B), and the expressions of Hes1 and Dll1 were elevated in the ENCCs isolated from the mutant embryonic guts (Figure 5, C and D). Given that Ptch1+/− ENCCs could migrate to the bowel and cell death was barely detected (Supplemental Figure 5), the reduction of proliferating postmigratory ENCC progenitors would be a consequence of premature gliogenesis of Ptch1+/− ENCC progenitors.

Given that gliogenesis starts at E12–E12.5 or later, Ptch1f/f/Wnt1-Cre embryos did not survive long enough to allow us to examine the effect of Ptch1 deletion on gliogenesis. To overcome this problem, mice with Cre knocked into the Hoxb3 b3-IIIa enhancer region (b3-IIIa-Cre mice) were used to specifically knockout Ptch1 in vagal neural crest cells. As described previously, Cre was only expressed transiently during E9.5–E10.5 in 30% of vagal neural crest cells (18), so Ptch1 was not completely deleted in these mutants. Nevertheless, these embryos exhibited similar phenotypes to those seen in the Ptch1+/−/Wnt1-Cre embryos (Supplemental Figure 6). Importantly, E12.5 mutant embryos (Ptch1+/−, b3-IIIa-Cre) could be collected and used to examine whether deletion of Ptch1 causes premature gliogenesis in vivo. As demonstrated in the whole-mount in situ hybridization, the glial marker, fatty acid–binding protein-7 (Fabp7), was barely detected in the control embryonic guts at E12.5, while markedly increased expression of Fabp7 was observed in the mutants (b3-IIIa-Cre, Ptch1+/−) (Figure 6 and Supplemental Figure 7). Taken together, our data showed that deletion of Ptch1 induces an early onset of gliogenesis of ENCCs in mice.

Hh-induced gliogenesis is conserved among mice and humans. Only Dll1, but not Dll3, serves to integrate the Notch and Hh pathways in mouse ENCCs, while DLL3 was found to genetically interact with PTCH1 and contribute to HSCR in humans. These observations prompted us to further examine whether Hh-induced gliogenesis is conserved between mice and humans. To achieve this, human neural crest–like cells, skin-derived precursors (SKPs), were used. SKPs have been shown to express a panel of neural crest markers and to be able to generate all major neural crest derivatives (19, 20), including cells that express many markers of enteric neurons (unpublished observations). As illustrated in Figure 7, SKPs isolated from human foreskin samples grew as neurospheres and expressed the stem cell marker, vimentin (Figure 7A). They exhibited high plasticity to form neurons
Important, similar to that in mouse ENCCs, Hh could induce expression of the glial protein GFAP in human SKPs (Figure 7B). Treatment with SHH significantly increased the number of GFAP+ cells when compared with those of the untreated controls (Figure 7B).

Discussion
The ENS in mammals is derived from a small pool of progenitor cells, the ENCCs. These precursor cells extensively proliferate to expand and migrate over a long distance to fully colonize the developing gut and differentiate into millions of neurons and glia, which are organized into the network responsible for coordinating the complex behaviors of the gut (1). This developmental process involves sequential waves of neurogenesis and gliogenesis and requires an appropriate balance between the proliferation and differentiation of ENCCs and their progeny. Therefore, integration of different signaling pathways coordinating proliferation and neuronal/glial differentiation is critical for proper ENS development and defects in any of these pathway components may lead to HSCR.

Genetic screenings on patients with HSCR have identified susceptibility loci and mutations associated with the disease. Mutations in the known HSCR genes, however, can only account for approximately 15%–20% of sporadic cases. Other HSCR cases may therefore likely result from the cumulative effects of less severe ablations in genetic pathways governing ENS development. To identify novel risk loci, our group has recently conducted a GWAS on HSCR. The genotype data generated from GWAS not only allowed us to define the risk loci that we believe to be new but also to detect gene-gene interactions. Here, we have integrated genetics, statistics, and bioinformatics with experimental biology to investigate the mechanisms by which a discrete number of genes interact with each other to cause disease.
We used CCA to assess whether interactions among gene members of these signaling pathways (Hh and Notch) could contribute to HSCR. To our knowledge, this is the first time that CCA was used to assess epistasis on data sets generated from a GWAS. This approach not only illuminates the genetic architecture of a complex disease but also enhances the potential of genetic variants for risk prediction using GWAS data. To validate our genetic data, we performed functional studies to demonstrate that the gene-gene interaction between Hh and Notch is not limited at the genetic level. Indeed, there was a functional intercross among these 2 pathways during ENS development. Importantly, both of these 2 independent approaches indicated that the Notch ligand Dll serves to integrate these pathways. Therefore, Dll may play a pivotal role in ENS development and the pathogenesis of HSCR.

It is interesting to note that fewer proliferating ENCC progenitors were observed in the Ptc1 mutant embryonic guts, even though both Hh and Notch are key signaling pathways implicated in preserving a pool of undifferentiated progenitors (9, 15, 21). This implies that although proper Hh function is necessary for the formation of sufficient numbers of enteric ganglia in the bowel (9), excess Hh pathway activation in ENCCs, on the other hand, causes progenitor progression (i.e., differentiation). It is known that the ENCC-derived neuronal and glial precursor cells continue to proliferate while progressively differentiating (22, 23). Therefore, during the early events of this developmental process, Hh may control the pool sizes of neurons and glia by mediating the proliferation of neural and glial precursors and/or switching neurogenesis to gliogenesis. Given that there was no obvious change in the neuronal precursors in the Ptc1 mutant embryonic guts at E10.5 (data not shown) and E11.5 (TuJ1+, Supplemental Figure 8), the progenitor progression may not be the result of a precise regulation of neuron production (pool size of neuronal precursors). In other words, it is possible that progenitor progression depends upon feedback inhibition and/or feed-forward induction mediated by pool size of naive ENCCs. Sustaining Hh activity may induce a rapid expansion of the ENCC pool, leading to a feedback inhibition of proliferation and subsequently, a feed-forward induction of progenitor progression. Indeed, even though deletion of Ptc1 promotes proliferation, we found that the Ptc1–/– ENCCs still retain their differentiation capacity to form neurons and glia in vivo (Supplemental Figure 4 and Figure 6). Similar observations have been reported previously in mice with the Ptc1 deletion in granule neuron precursors (GNPs) and neural stem cells (NSCs). While Ptc1–/– GNPs and NSCs exhibited increased proliferation, they still remain capable of differentiating into astrocytes, oligodendrocytes, and neurons (6). Most likely, there is an additional layer of signaling refinement in the gut environment to compensate for the inhibitory effect of Hh in the process of neurogenesis.

Our functional data suggest that activation of Notch would be the feed-forward signaling for progenitor progression. In this respect, activated Notch has been shown to have an iterative role on mitotic and postmitotic precursor cells in specifying or promoting a particular cell fate, notably glia cell fate during nervous system development (11, 24, 25). Notch activity permits the early progenitor cells to remain proliferative and undifferentiated, and downregulation of Notch activity is required for the cells to exit the cell cycle. Subsequent reactivation of the Notch pathway mainly regulates the neuronal versus glial fate choice (26). This temporary release from Notch activation allows resumption of the production of temporally appropriate neurons and is crucial for proper glial differentiation (26, 27). Therefore, constitutive activation of the Hh pathway in ENCCs causes sustained Notch activity. Absence of a release from Notch activation may result in incomplete neuronal differentiation and premature gliogenesis (Supplemental Figure 9).

Our initial GWAS has already defined the glial mediator neuregulin 1 (NRG1) as a major risk locus for HSCR aside from SNPs in RET (14). Our current study further highlighted the relevance of genes governing gliogenesis in the pathogenesis of HSCR. In
summary, we have identified HSCR susceptibility genes and specific genotype constellations of PTCH1 (a Hh receptor) and DLL3 (a Notch ligand) SNPs that conferred higher risk to HSCR. In addition, we provided definitive in vivo evidence of the essential role played by the interaction between Hh and Notch pathways in coordinating neuronal and glial cell differentiation during ENS development. We also defined a molecular mechanism in which Dll integrates Hh and Notch pathways to promote glial differentiation. In concordance with our initial GWAS, our current data further indicated that premature gliogenesis may represent a new disease mechanism for HSCR, and Hh-Notch–induced premature gliogenesis may lead to HSCR.

Methods

Epistasis analysis

A GWAS on HSCR was previously conducted with 173 cases and 436 controls using Affymetrix 500K chips (14). Gene-gene interaction was interrogated on genes members of the Hh and Notch signaling pathways (Supplemental Table 1).

We decided to implement CCA, because it is the most general of multivariate techniques (28). It is also a natural statistical framework to study 2 sets of variables and see what is common among the 2 sets. CCA differs from existing methods for gene-gene interaction detection (i.e., logistic regression) in that it simultaneously considers all markers in the genes to be correlated. This can both reduce the multiple-testing burden and increase statistical power. CCA estimates the maximum correlation of 2 canonical variables, where each canonical variable is a set of linear combinations of SNPs from 1 gene. In general, there may exist more than one significant canonical correlation; however, the first canonical correlation is always the one explaining most of the relationship between the genes tested. In this study, the CCA among genes was obtained for cases and controls separately. Then we calculated the P value (evaluated via permutation) of the differences in gene-pair first canonical correlations between cases and controls. The largest significant correlation difference between patients and controls identified the 2 interacting genes. Correlations between genes for each group are illustrated in Supplemental Table 2.

To correct for population stratification, we partitioned out the population effect (EIGENSTRAT) using partial CCA (29). The partial CCA is the natural generalization of partial correlation from the univariate to the multivariate situation. Given 3 vectors of random variables, X, Y, Z, CCA allows study of the relationship between X and Y after partitioning out the effect of vector Z on both X and Y vectors (e.g., make the effect of Z constant). While regular CCA uses only the variance-covariance matrix of X and Y, the partial CCA uses the conditional variance-covariance matrix of X given Z (e.g., the residualized variance-covariance matrix) to partition out the effect of Z. Partial CCA was also used to investigate whether the epistasis identified between Hh and Notch was dependent on RET SNPs (rs2082106, rs2742234, rs17028). We next stratified the patients and controls according to the high-risk or low-risk genotypes generated by MDR to assess the relative risk to HSCR in terms of OR (30). The EIGENSOFT package and the MDR program were used.

Mice

Ptch1<sup>fl/fl</sup>, Wnt1-Cre, and b3-IIla-Cre mice have been described previously (17, 18, 31). Swiss mice (Canadian CD-1 [ICR]) were from Charles River Laboratories. Animals were maintained in the Laboratory Animal Unit at the University of Hong Kong, and all experiments were performed in accordance with procedures approved by the committee on the Use of Live Animals, the University of Hong Kong (CULTRA 1998-09).

ENCC culture

All ENCCs were isolated from E11.5 ICR, Ptch1<sup>fl/fl</sup>, or Ptch1<sup>fl/fl</sup>/Wnt1-Cre mouse embryonic guts (from stomach to hindgut), and guts were dissected in L15 medium (Invitrogen). Guts were washed with Ca<sup>2+</sup>- and Mg<sup>2+</sup>- free PBS and digested with collagenase/dispase (0.2 μg/ml each; 37°C for 10 minutes). Digested guts were triturated into single cells and filtered through cell strainers (100-μm strainers and then 40-μm strainers, Falcon, BD Biosciences). Cells were resuspended in neural crest medium, DMEM containing 15% chick embryo extract (SIL), FGF (20 ng/ml, Sigma-Aldrich), EGF (20 ng/ml, Sigma-Aldrich), Retinoic acid (35 ng/ml, Sigma-Aldrich), N2 (1%), B27 (2%, Invitrogen), and β-mercaptoethanol (50 mM, Sigma-Aldrich); plated onto poly-o-lysine–coated and fibronectin-coated (20 μg/ml each, Invitrogen) wells; and enriched by multiple replating as described previously (15, 32, 33). The ENCCs at passage 3 were harvested for quantitative RT-pCR or seeded at clonal density (1 x 10<sup>2</sup> to 1.5 x 10<sup>5</sup> cells per each 35-mm well and 1.5 x 10<sup>3</sup> cells per each 20-mm well) and used for the subsequent functional analyses.
Human SKP culture
SKPs were prepared from human foreskin as described previously (19). Briefly, dissected skin was minced into 4- to 5-mm² pieces and then digested with Accumax (Chemicon) for 2 hours at room temperature, followed by overnight incubation with Accumax at 4°C. After removal of the epidermis, the tissues were cut into 1- to 2-mm² pieces and further digested with Accumax with collagenase XI (1 mg/ml, Sigma-Aldrich) and DNase I (0.2 mg/ml, Sigma-Aldrich) at 37°C for 30 minutes, mechanically dissociated, and filtered through a 70-μm cell strainer (Falcon). Dissociated cells were pelleted and plated in DMEM/F12 (3:1; Invitrogen) containing 20 ng/ml EGF and 40 ng/ml FGF2 (both from Sigma-Aldrich). For differentiation assay, SKPs were plated on chamber slides (Nalge Nunc) coated with poly-l-lysine-laminin in DMEM/F12 (3:1) supplement with 10% FBS. Recombinant human Shh (2 mg/ml; C24II, R&D Systems) was added 1 hour before the termination of culture. Cells in 35-mm wells were also treated with Shh inhibitor, cyclopamine (10 μg/ml or human recombinant GDNF at 100 ng/ml) or in combination for 72 hours. For the signaling studies, the nonspecific siRNA control (catalog no. 1022079, AATTCTCCGAACGTGT CACGT) (Qiagen) were used. 1.5 × 10⁵ ENCCs were plated on chamber slides (Nalge Nunc) coated with poly-l-lysine-laminin in DMEM/F12 (3:1) supplement with 10% FBS. Recombinant human Shh (2 mg/ml; C24II, R&D Systems) was added 1 week after culture and incubated for 7 additional days. Cells were then fixed and stained with specific antibodies as mentioned below.

Viral transduction
Cre-GFP (Ad-GFP-Cre) or GFP (Ad-GFP) recombinant adenovirus (Vector Bioloaboratories) was added to cells in a 1:50 dilution (~1 × 10⁶ PFU/ml). The infection efficiency was estimated by counting the GFP-positive cells under the fluorescence microscope (Olympus CXX41 Inverted Microscope; Olympus). Seventy-six hours after the infection, approximately 90% of the cells expressed GFP, and then cells were harvested. Deletion of Ptch1 and the activation of the Hh signaling were confirmed by RT-PCR and Western blot as described below. Cell proliferation assays were performed 7 days after infection.

RNA interference
Dll1 siRNA (catalog no. S100980007, CGTCTATCTTGATTACTA) and a nonspecific siRNA control (catalog no. 1022079, AATTCTCCGAACGTGT CACGT) (Qiagen) were used. 1.5 × 10⁵ ENCCs were transfected with Dll1 or nonspecific siRNA using LipofectAMINE 2000 (Invitrogen) according to the manufacturer’s instructions. After 24 hours, the cells were subjected to cell proliferation assay and RNA extraction in the presence or absence of Shh.

Cell proliferation assay
Cell proliferation was measured using the In Situ Cell Proliferation Kit, Fluos (Roche). 1.5 × 10⁵ and 1.5 × 10⁶ ENCCs were seeded per slide chamber and 35-mm culture plate, respectively. The cells were treated with either human recombinant Shh at 2 μg/ml or human recombinant GDNF at 100 ng/ml (PeproTech) or in combination for 72 hours. For the signaling studies, the cells were also treated with Shh inhibitor, cyclopamine (10 μg/ml, Calbiochem); Shh agonist, purmorphamine (1 μM, Calbiochem); or Notch inhibitor, DAPT (10 mM, Calbiochem) alone or in combination with Shh or GDNF. BrdU was added 16 hours before the termination of culture. In 35-mm culture plates were then harvested, cytospun onto glass slides, and subjected to immunocytochemistry. Cells were fixed, blocked, and incubated with anti-Ret antibody (1:200, Neuronics). The immunosignal was detected using anti-goat IgG conjugated with Texas Red (1:200, Chemicon), while the glial differentiation was detected by staining with antibodies specific for S100β (rabbit 1:200, Sigma-Aldrich) or GFAP (1:200, DAKO); the immunosignal was then detected using the secondary antibody conjugated with Texas Red. All sections were counterstained with DAPI (blue) to detect nuclei. Numbers of TH⁺, Tuj1⁺, and S100β⁺ cells were counted over the total number of cells (DAPI) in both control (Ad-GFP) and mutant (Ad-GFP-Cre) groups, with or without treatment. For each treatment group, a minimum of 9 random fields under ×200 magnification with at least 250 cells in total per 35-mm well was photographed for cell counting. The values reported in bar charts represent the mean ± SEM of 2 wells (i.e., 18 random fields), and the experiments were repeated 3 times.

Immunohistochemistry. For section immunohistochemistry, embryos were fixed in 4% PFA in PBS at 4°C, dehydrated, and embedded in paraffin or cryoprotected in 30% sucrose in PBS at 4°C and embedded in OCT compound (Tissue-Tek). The sections were blocked in PBS containing 10% normal goat serum (DAKO) for 1 hour at room temperature and then incubated overnight at 4°C in a mixture of the anti-p75NTR (1:2,000, Chemicon) and anti-Ki67 (1:100, BD Pharmingen). Cryosections were used for the detection of Hes-1 expression with rabbit anti–Hes-1 antibody (1:100, a gift from Tetsuo Sudo, Toray Industries Inc., Kanagawa, Japan). After washing, the immunosignals were then detected using the secondary antibody conjugated with Texas Red or FITC.

For double staining using immunohistochemistry and in situ hybridization, paraffin sections were hybridized with digoxigenin-labeled (DIG-labeled) antisense cRNA probes (Roche). The hybridized probes were detected using anti-DIG antibody and amplified by the Renaissance TSA Biotin System (PerkinElmer) according to manufacturer’s protocol. These sections were then incubated with anti-p75NTR (Chemicon) for the detection of ENCCs, followed by a goat anti-rabbit IgG secondary antibody conjugated with Texas Red (1:200, Molecular Probes). The signals were then detected by streptavidin-fluorescein conjugate. Sections were counterstained with DAPI and mounted with aqueous mounting media. ENCCs and tissue sections were photographed using a Nikon Eclipse E600 microscope, with Sony digital camera DSM1200F, under fluorescence illumination.

Whole-mount in situ hybridization
In situ hybridization was used to examine the expression of Fabp7 (NM_ 021272, GenScript) in embryonic guts. E12.5 embryonic guts were dissected out of control (Pcth1+/+) and mutant (Pcth1−/−, bIIIIa-Cre) embryos, fixed with 4% PFA, and cryoprotected with 30% sucrose. The whole embryonic guts were hybridized with DIG-labeled RNA probe encoding the coding region of Fabp7, following the standard protocol (34). The signals were detected with BM Purple AP Substrate (Roche). The embryonic guts were then sectioned and photographed.

TUNEL
Apoptotic cells in ENCC culture and embryos were analyzed on a cytospin preparation of ENCC and paraffin sections, respectively, using the In Situ Cell Death Detection Kit, Fluorescein (Roche), following the manufacturer’s protocol.

Immunofluorescence studies
In Immunocytochemistry. To determine the lineage potential of ENCCs after deletion of Pcth1, we analyzed differentiation of adenovirus-infected Pcth1−/− ENCCs. Cells were plated at clonal density, subjected to virus infection for 72 hours, and then treated with GDNF, Dll1 (R&D Systems), or Notch inhibitor and cultured for an additional 3–10 days. Neuronal lineage commitment was monitored based on expression of TH using rabbit anti–TH antibody (1:1,000, Immunex) or Tuj1 with mouse anti-Tuj1 antibody (1:200, Chemicon), while the glial differentiation was detected by staining with antibodies specific for S100β (rabbit 1:200, Sigma-Aldrich) or GFAP (1:200, DAKO); the immunosignal was then detected using the secondary antibody conjugated with Texas Red. All cultures were counterstained with DAPI (blue) to detect nuclei. Numbers of TH⁺, Tuj1⁺, and S100β⁺ cells were counted over the total number of cells (DAPI) in both control (Ad-GFP) and mutant (Ad-GFP-Cre) groups, with or without treatment. For each treatment group, a minimum of 9 random fields under ×200 magnification with at least 250 cells in total per 35-mm well was photographed for cell counting. The values reported in bar charts represent the mean ± SEM of 2 wells (i.e., 18 random fields), and the experiments were repeated 3 times.

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**Immunoblots**

To examine the activation of cellular signaling, cells were treated with drugs. Cells from three 35-mm wells were collected and then lysed with cell lysis buffer containing 20 mM Tris (pH 7.5), 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1% Triton X-100, 2.5 mM sodium pyrophosphate, 1 mM β-glycerophosphate, 1 mM Na3VO4, 1 μg/ml leupeptin, and 1 mM phenylmethylsulfonyl fluoride. Cell lysates containing 20 μg total protein were separated on 8%–10% SDS-polyacrylamide gels and blotted onto nitrocellulose membranes. The membranes were then incubated with polyclonal antibodies against Gli1 (1:3,000, Abcam), Gli3 (1:1,000, Santa Cruz Biotechnology Inc.), Patched-1 (1:500, R&D Systems), and NICD (1:1,000, Millipore). The same membranes were stripped with stripping buffer (100 mM methanesulfonyl fluoride). Cell lysates containing 20 μg total protein were separated on 8%–10% SDS-polyacrylamide gels and blotted onto nitrocellulose membranes. The membranes were then incubated with polyclonal antibodies against Gli1 (1:3,000, Abcam), Gli3 (1:1,000, Santa Cruz Biotechnology Inc.), Patched-1 (1:500, R&D Systems), and NICD (1:1,000, Millipore).

**RT-PCR**

Total RNA was isolated from ENCCs by the RNeasy Mini Kit (Qiagen) and reverse transcribed in 20 μl reaction system using the SuperScript RNA Amplification System (Invitrogen), in accordance with the manufacturer’s instructions. PCR reactions were performed using specific primers (listed in Supplemental Table 3). Results were normalized and expressed relative to the internal control, actin.

**Quantitative RT-PCR**

RNA for RT-PCR was extracted from mouse embryonic guts or ENCCs using the RNeasy Mini Kit (Qiagen) and reverse transcribed in 20 μl reaction mix as above. Quantitative PCR was performed in the Cybergreen reaction mix (Applied Biosystems), which consisted of 1× mastermix and forward and reverse primers. The reaction mix (18 μl) was aliquoted into tubes, and 2 μl cDNA was added. Triplicated 20 μl reaction system using the SuperScript RNA mastermix and HKU765407, HKU775907, and HKU752806 to M.M. Garcia-Barceló, P.K.H. Tam, and V.C.H. Lui, respectively, from the Hong Kong Research Grants Council. The authors thank Robin Lovell-Badge (MRC National Institute for Medical Research, London, United Kingdom), S.Y. Tsai (Baylor College of Medicine, Houston, Texas, USA), and Kathryn Cheah (University of Hong Kong, Hong Kong, China) for critical reading of the manuscript.

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**Statistics**

The differences among multiple treatment groups were analyzed with a 2-sided unpaired Student t test or 1-way ANOVA, followed by Tukey post-hoc test using GraphPad Prism 5 (GraphPad Software). A P value of less than 0.05 was interpreted to represent a statistically significant difference. All experiments were replicated at least 3 times, and data are shown as mean ± SEM.

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